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Elongation of the Hydrophobic Chain as a Molecular Switch: Discovery of Capsaicin Derivatives and Endogenous Lipids as Potent Transient Receptor Potential Vanilloid Channel 2 Antagonists

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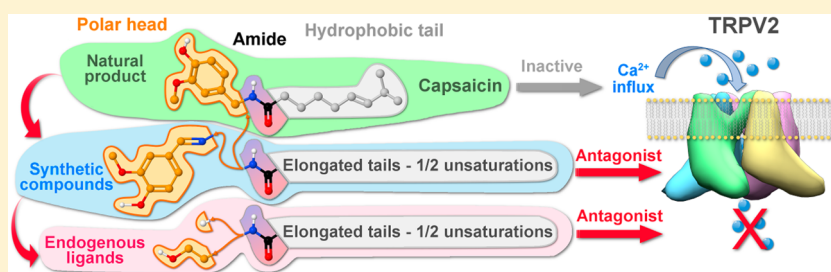
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S Supporting Information



ABSTRACT: The transient receptor potential vanilloid 2 (TRPV2) is a nonselective Ca^{2+} permeable channel member of the TRPV subfamily, still considered an orphan TRP channel due to the scarcity of available selective and potent pharmacological tools and endogenous modulators. Here we describe the discovery of novel synthetic long-chain capsaicin derivatives as potent TRPV2 antagonists in comparison to the totally inactive capsaicin, the role of their hydrophobic chain, and how the structure–activity relationships of such derivatives led, through a ligand-based approach, to the identification of endogenous long-chain fatty acid ethanolamides or primary amides acting as TRPV2 antagonists. Both synthetic and endogenous antagonists exhibited differential inhibition against known TRPV2 agonists characterized by distinct kinetic profiles. These findings represent the first example of both synthetic and naturally occurring TRPV2 modulators with efficacy in the submicromolar/low-micromolar range, which will be useful for clarifying the physiopathological roles of this receptor, its regulation, and its targeting in pathological conditions.

1. INTRODUCTION

TRPV2 belongs to the polymodal transient receptor potential (TRP) superfamily of calcium-permeable nonselective cation channels, activated by a wide variety of physical and chemical stimuli. Due to its mechanosensor property, TRPV2 is considered a stretch-modulated channel and a regulator of calcium homeostasis in different tissues and organs, in particular the heart, where it is 10-fold more abundant than in skeletal muscle.¹ Different lines of evidence suggest for TRPV2 a key role in physiological cardiac function as well as in cardiomyopathies and dystrophic diseases.^{2–4} Besides the heart, TRPV2 is also found in the brain, vascular smooth muscle cells, the gastrointestinal tract, macrophages, and the urothelial tract,⁵ and it is involved in a number of

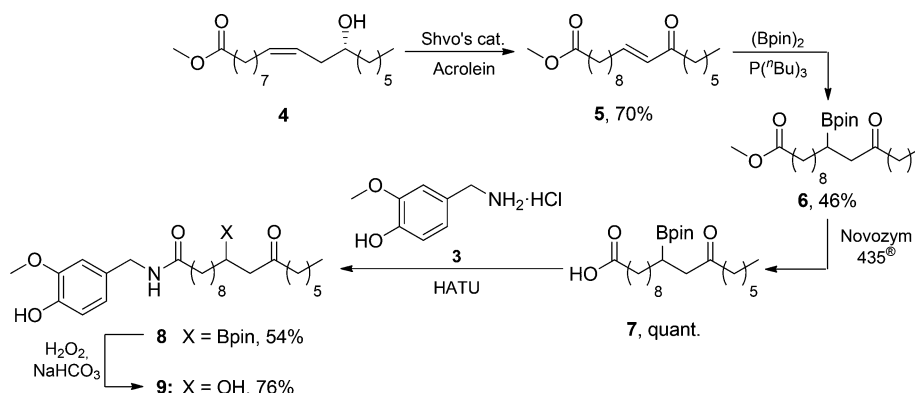
physiopathological processes,⁶ including cancer,^{7–9} particularly of the urinary tract.^{10–13}

Despite its biological and pharmacological relevance, TRPV2 is still considered an orphan TRP channel due to the scarcity of selective drugs and known endogenous ligands. The 2-aminoethoxydiphenyl borate (2APB) is one of the first nonselective activators identified for rat TRPV2 ($\text{EC}_{50} = 129 \mu\text{M}$),¹⁴ although inactive at the human orthologue, suggesting a strong species specificity.^{15,16} *Cannabis sativa* derivatives such as Δ^9 -tetrahydrocannabinol (Δ^9 -THC), cannabidiol (CBD), and Δ^9 -tetrahydrocannabivarin (Δ^9 -THCV) are TRPV2 activators^{17,18} and so is *p*-(di-*n*-propylsulfamyl)benzoic acid⁴⁸

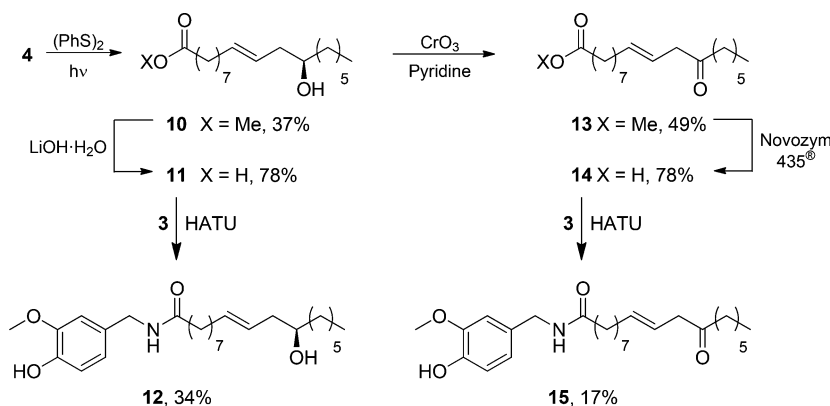
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Scheme 1. Synthesis of Compound 9



Scheme 2. Synthesis of Compounds 12 and 15



49 (probenecid).¹⁹ However, all these agonists are known to
50 modulate other TRP channels. Most TRPV channels are
51 proposed to be modulated also by phosphoinositide lipids.²⁰
52 TRPV2-mediated Ca^{2+} influx has been reported following
53 stimulation by endogenous lysophospholipids such as
54 lysophosphatidylcholine (LPC) and lysophosphatidylinositol
55 (LPI),²¹ LPC being a relatively potent activator ($\text{EC}_{50} = 3.4$
56 μM).²² To date, the nature of endogenous regulators of
57 TRPV2 activity still remains elusive.²³

58 Also synthetic inhibitors of TRPV2 are either not specific or
59 endowed with low potency, as exemplified by ruthenium red
60 ($\text{IC}_{50} = 0.6 \mu\text{M}$),²⁴ a pore blocker that inhibits other 12 ion
61 channels,²⁵ La^{3+} and Gd^{3+} ,²⁶ citral;²⁷ the alkylated imidazole
62 SKF96365;¹⁶ tetraethylammonium and 4-aminopyridine, two
63 potassium channel blockers; 1-(2-(trifluoromethyl)phenyl)-
64 imidazole, an inhibitor of capacitative Ca^{2+} entry;¹⁶ and
65 tranilast,²⁸ which has been used in several studies,^{29–34} even
66 though it has never been validated as TRPV2 antagonist.

67 TRPV2 shares high sequence identity (>50%) with TRPV1,
68 but its threshold of activation by temperature is higher (>52
69 $^{\circ}\text{C}$)²⁴ and, unlike TRPV1, is not sensitive to capsaicin. The
70 recently solved cryo-EM structures of both TRPV1 and
71 TRPV2,^{35,36} along with mutagenesis and computational
72 studies, showed that the TRPV1 binding site of capsaicin is
73 not conserved in TRPV2. Furthermore, the replacement of
74 critical residues leads to a mutant (TRPV2-Quad) against
75 which capsaicin behaves as an antagonist rather than an agonist
76 as in TRPV1.³⁷ These intriguing results prompted us to
77 investigate a series of capsaicin derivatives in which the
78 vanillylamide polar head of capsaicin bears a longer alkyl
79 chain, featuring different length, unsaturation degree, and type

of polar substituents. The structure–activity relationship
80 (SAR) of these synthetic compounds then suggested the
81 screening of structurally related endogenous lipids sharing at
82 least one functional group with the capsaicin derivatives, with
83 the aim of finding new endogenous modulators. 84

2. RESULTS

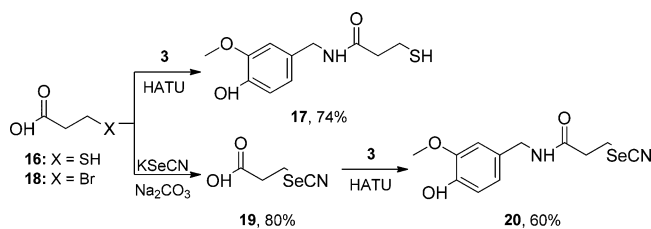
85 **2.1. Synthesis.** Commercial fatty acids such as ricinoleic
86 acid, oleic acid, and palmitic acid were used as starting material
87 to synthesize the 23 compounds tested. Scheme 1 shows the
88 synthesis of the α,β -unsaturated ketone 5 by the ruthenium-
89 catalyzed oxidation in anhydrous toluene of the homoallylic
90 alcohol of the methyl ricinoleate 4.³⁸ Shvo's catalyst and
91 acrolein were used as catalyst and hydrogen scavenger,
92 respectively.³⁹ The addition of bis(pinacolato)diboron
93 (Bpin_2) to the enone 5 in the presence of tri-*n*-butylphosphine
94 ($\text{P}(\text{nBu})_3$)⁴⁰ yielded the β -boronketone 6 in 46% yield.
95 Enzymatically controlled hydrolysis⁴¹ of the methyl ester 6
96 with Novozym 435 lipase led to the carboxylic acid 7
97 quantitatively. This acid 7 was coupled, without any further
98 purification, with 4-hydroxy-3-methoxybenzylamine hydro-
99 chloride 3 by HATU⁴² and DIPEA in anhydrous DMF,
100 achieving the amide 8. The oxidative hydrolysis of the boron
101 substituent of the compound 8 led to the β -hydroxyketone 9 in
102 a 76% yield (Scheme 1).

103 The irradiation of alcohol 4 with diphenyl sulfide⁴³ in
104 isooctane in a photochemical reactor for 3 h led to the isomer
105 10 in 37% yield after several recrystallizations at -30°C . This
106 compound was used to synthesize two new long-chain *N*-
107 vanillylamides (12, 15). The hydrolysis of the methyl ester of 107

10 led to the corresponding carboxylic acid **11**. The subsequent coupling of **11** with the 4-hydroxy-3-methoxybenzylamine hydrochloride **3** using the same conditions described above yielded compound **12** in a 34% yield. Compound **10** was also oxidized with CrO_3 in pyridine⁴⁴ to prepare the *trans* ketone **13** (49% yield), which was enzymatically hydrolyzed to synthesize the corresponding acid **14** in a 78% yield. Subsequently, **14** was coupled with the vanillyl amine **3** to yield the (*E*)-*N*-(4-hydroxy-3-methoxybenzyl)-12-oxooctadec-9-enamide **15** after purification by liquid column chromatography (17% yield) (Scheme 2).

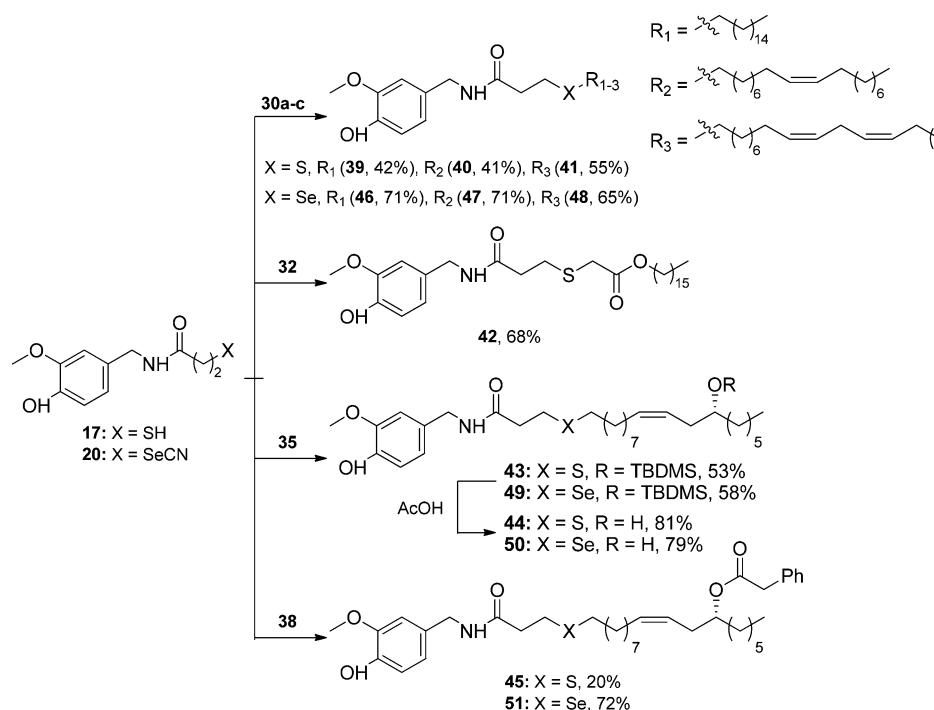
Scheme 3 shows the synthesis of the sulfur- and seleno-derivatives of **3**. Mercaptopropionic acid **16** was coupled with

Scheme 3. Synthesis of Sulfur and Seleno Intermediates



4-hydroxy-3-methoxybenzylamine hydrochloride **3** using HATU and DIPEA in anhydrous DMF, achieving the amide **17** (74% yield). The synthesis of the seleno-derivatives started with bromopropionic acid **18**, which was treated with KSeCN in water: The neutralization with Na_2CO_3 , yielded the selenocyanatopropionic acid **19** in 80% without purification. Finally, compound **19** was coupled with the 4-hydroxy-3-methoxybenzylamine hydrochloride **3** to obtain compound **20** after purification by liquid column chromatography (60% yield).

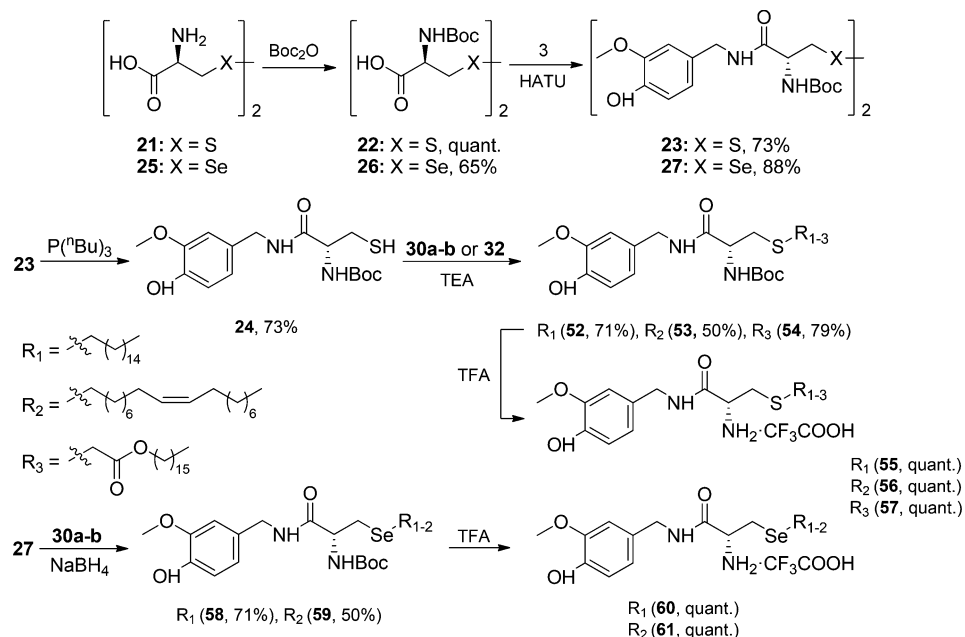
Scheme 4. Synthesis of No-Branched Sulfur- and Seleno-Derivatives



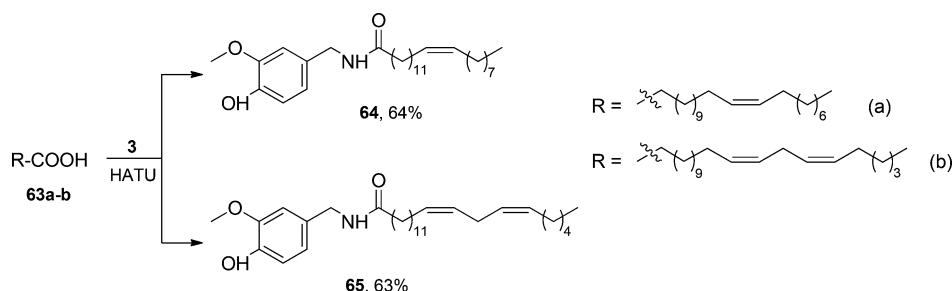
Amide **17** was S-alkylated with the previously synthesized alkylating derivatives **30a–c**, **32**, and **35** (see Supporting Information) in DMF and triethylamine, obtaining the long-chain *N*-vanillylamides **39–43** and **45** in 41–68% yield. *N*-Vanillylamide **44** was successfully achieved after removing the TBDMS protecting group with acetic acid at room temperature (81% yield). New long-chain *N*-vanillylamides were obtained from compound **20**, which was first treated with NaBH_4 in ethanol at room temperature to remove the cyano protection and regenerate the selenol group.⁴⁵ Subsequent Se-alkylation was carried out in one-pot with the addition of diverse set of alkylating reagents (**30a–c**, **35**, and **38**). *N*-Vanillylamides **46–49** and **51** were synthesized in 71–87% yields. Compound **50** was successfully prepared after removing the TBDMS protecting group with acetic acid at room temperature (79% yield) (Scheme 4).

Scheme 5 shows the synthesis of amino-branched analogues. The first step consisted of the treatment of L-cysteine **21** or L-selenocysteine **25** with Boc_2O in the presence of triethylamine to afford the protected derivatives **22**¹ and **26**² (quantitative and 65% yield, respectively).^{46,47} These compounds were coupled with 4-hydroxy-3-methoxybenzylamine hydrochloride **3** using EDCl, HOBt, and triethylamine (TEA) in anhydrous DMF, achieving the amides **23** and **27** (74% and 88% yield). The reduction of compound **23** with $\text{P}(\text{tBu})_3$ in wet dichloromethane afforded compound **24** in a 73% yield after purification by liquid column chromatography. New long-chain *N*-vanillylamides were afforded from compound **24**, which was S-alkylated with the previously synthesized alkylating derivatives **30a–c** and **32** in the presence of triethylamine, obtaining the long-chain *N*-vanillylamides **52**, **53**, and **54** in moderate yields (50–79% yield). The *N*-Boc deprotection was carried out using trifluoroacetic acid⁴⁸ in dichloromethane yielding *N*-vanillylamides **55**, **56**, and **57** as trifluoroacetic salts in quantitative yields. Compound **27** was reduced with NaBH_4

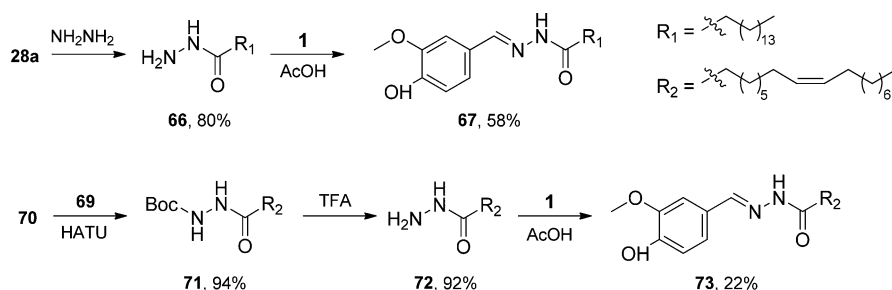
Scheme 5. Synthesis of Amino-Branched Analogues



Scheme 6. Synthesis of Compounds 64 and 65



Scheme 7. Synthesis of Compounds 67 and 73



in ethanol at room temperature to cleave the diselenium bond.⁴⁹ The Se-alkylation was carried out with the addition of the alkylating derivatives 30a,b to afford the *N*-vanillylamides 58 and 59 in 74–88% yields. Finally, The *N*-Boc deprotection was carried out using the same conditions described above to afford the *N*-vanillylamides 60 and 61 as trifluoroacetic salts. Acids 63a,b, which were previously obtained from the hydrolysis of their respective methyl esters 62a,b (see Supporting Information), were coupled with the 4-hydroxy-3-methoxybenzylamine hydrochloride 3 using HATU and DIPEA in anhydrous DMF, achieving the amides 64 and 65 after purification by liquid column chromatography (64% and 63% yield) (Scheme 6).

Methyl palmitate 28a was treated with an excess of hydrazine hydrate in ethanol to synthesize the palmitic acid hydrazide 66 (80% yield). The addition of the aromatic aldehyde vanillin 1 to compound 66 in the presence of acetic acid in reflux conditions gave the Schiff's base compound 67 in 58% yield.⁵⁰ A similar compound was synthesized starting from oleic acid 70, which was coupled to *tert*-butyl hydrazine-carboxylate 69 using HATU and DIPEA in DMF to yield the oleylhydrazide 71 in a 94% yield. The *N*-Boc deprotection of oleylhydrazide 71 with TFA in DCM for 2 h led to oleylhydrazide 72 in 92% yield. Compound 72 refluxed with vanillin 1 in the presence of acetic acid in methanol produced the Schiff base 73 in 22% yield (Scheme 7).

2.2. Biological Evaluation. 2.2.1 Capsaicin Derivatives

Activate TRPV1 Channel. The capsaicin scaffold (Figure 1)⁵¹ can be ideally divided into three regions: head, neck, and

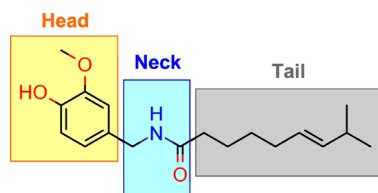


Figure 1. Chemical structure of capsaicin. The vanillyl head, the amide neck, and hydrophobic tail are shaded in yellow, cyan, and gray, respectively.

heterologously expressed in HEK-293 cells. The tested compounds did not significantly activate TRPV2-mediated Ca^{2+} elevation in transfected HEK-293 cells. Instead, preincubation (5 min) of TRPV2-HEK-293 cells with different concentrations of the tested compounds, followed by incubation with LPC (3 μM), caused inhibition of intracellular Ca^{2+} elevation due to TRPV2 response to LPC. The corresponding IC_{50} values are reported in Table 1.

The structure–activity relationships (SARs) of these compounds suggested a critical influence on the capability to exert TRPV2 antagonism of the alkyl chain and, in particular, of its hydrophobicity, length, and degree of unsaturation. Hydrophobicity is important since, as shown in Table 1, the activity dramatically dropped after introduction in the chain of polar substituents such as hydroxyl, keto, or ester groups (these groups arising from esterification of the hydroxyl group) or their combinations (42, 44, 50, 45, 51, 9, 12, 15). However, the presence of an amino group next to the amide (55, 60, 56, 61), which had marginal effects for already active compounds, by only slightly increasing their potency (60 vs 46), was instead dramatic for those inactive compounds bearing a hydroxyl or an ester moiety in the alkyl chain, whose activity was completely rescued (see 42 vs 57). The complete recovery of activity after introduction of an amino group next to the amide in derivatives bearing a polar substituent in the alkyl chain suggests that reinforcement of the polar interactions of the “head” avoids the competition with the polar-substituted alkyl chain for interaction with receptor polar residues in a region where the polar head, but not the alkyl chain, should be hosted to elicit a measurable effect. The chain is fairly more tolerant to changes not substantially affecting the hydrophobicity of the alkyl group: replacement of sulfur with selenium in the alkyl chain did not affect significantly ligand activity (39 vs 46); its replacement with a carbon atom determined an increase in potency (64 vs 40/47). While polar functionalization of the alkyl chain caused a dramatic drop of activity, amino or imino groups (67, 73) were well tolerated in the region close to the amide moiety of capsaicin. In particular, the imino derivatives were among the most active compounds within the series (IC_{50} = 0.28 and 0.12 μM , respectively). Also length and unsaturation degree of the alkyl chain significantly affected the activities of the tested compounds. The C16:0 and C18:0 saturated analogs were inactive, whereas the C20:0 derivative showed an IC_{50} = 3.1 μM . The insertion of a single double bond in C18 chain (olvanil) dramatically increased the antagonism, with IC_{50} = 0.16 μM .

Thus, the screening led to the identification of several very potent TRPV2 antagonists, exhibiting IC_{50} values in the subnanomolar to low-micromolar range. This result is quite remarkable since, despite its close homology to TRPV1, TRPV2 is insensitive to capsaicin, the residues being responsible for capsaicin binding and receptor activation in TRPV1 not conserved in TRPV2.⁵⁸

The most striking result from the SAR of capsaicin derivatives against LPC is that the elongation of the alkyl chain of capsaicin causes a switch of such scaffold from inactivity toward potent antagonism at rat recombinant TRPV2. Intriguingly, the dependence of TRPV2 modulation on the length of the ligand alkyl chain has already been observed for lysophospholipids, which require a carbon chain longer than C12 to stimulate the receptor.²¹

2.2.3. Capsaicin Derivatives Inhibit TRPV2 Channels Activated by CBD. Due to different latency in the activation

tail, formed by the vanillyl moiety, the amidic group, and the lipophilic alkyl chain, respectively. Structural variations, including incorporation of sulfur atom, into the head and the neck regions have been described in the literature.^{52–55}

Instead, the effect of a sulfur atom in the alkyl chain has been less investigated. The recent availability of the 3D structure of TRPV1⁵⁶ along with mutagenesis studies⁵⁷ allowed the identification of the capsaicin binding site, where the alkyl chain is hosted in a phenylalanine-rich hydrophobic region close to Thr550, a residue involved in H-bond interaction with the ligand amide group. The presence of a sulfur atom near the neck region should in principle lead to an increment of activity due to favorable dipole–dipole and aromatic–sulfur interactions. Since sulfur can be substituted with selenium via isosteric replacement, we also synthesized the corresponding selenium analogs. Selenium is an essential trace element whose role in medicine and biology is just starting to be elucidated. Some selenium-containing compounds have provided protection against many degenerative conditions, including cancer. Thus, a series of novel capsaicin derivatives, i.e., 9, 12, 15, 39, 46, 55, 60, 42, 57, 44, 56, 40, 45, 65, 41, 48, 64, 47, 61, 51, 50, 67, 73, whose structures are reported in Tables 1 and 2, featuring the same “head” and “neck” as capsaicin but differing in length and nature of the hydrocarbon tail, were tested on human TRPV1 heterologously expressed in human embryonic kidney (HEK)-293 cells by fluorometric assay (see Tables S1 and S2 in Supporting Information). The predicted activities as TRPV1 agonists were confirmed for many compounds within the series, exhibiting EC_{50} values from high-nanomolar to subnanomolar range. A SAR analysis of the results also disclosed the critical role of the region flanking the amide group in modulating the activity. In fact, the insertion of a positive charge next to the amide group was detrimental for activity (compounds 55–57 and 60), and the introduction of an imido group between the aromatic moiety and the amido group led to totally inactive compounds (compounds 67 and 73). Conversely, the introduction of a single polar substituent (hydroxyl, ester, or ketone) was well-tolerated, and the introduction of a sulfur or selenium atom in the hydrophobic tail even improved the activity. However, on the basis of the antagonist activity exhibited by capsaicin on TRPV2-Quad,³⁷ the new compounds were also tested on TRPV2 to determine if the elongation and the functionalization of the alkyl chain could elicit a functional response at this receptor.

2.2.2. Capsaicin Derivatives Inhibit TRPV2 Channels Activated by LPC. The activity of the synthesized capsaicin derivatives on TRPV2 was evaluated in vitro. The assays were conducted using a fluorometric assay with rat TRPV2

Table 1. Antagonist Potency of Capsaicin-like Compounds at TRPV2 against LPC (3 μM) and CBD (2 μM), Reported as IC_{50} (μM)

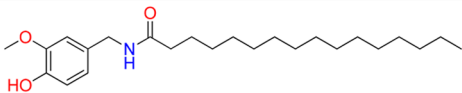
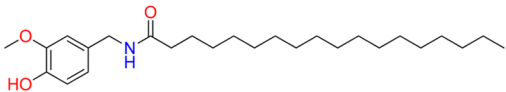
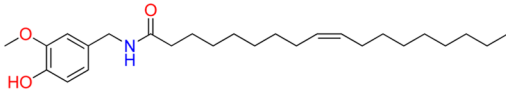
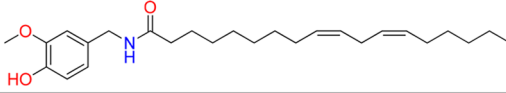
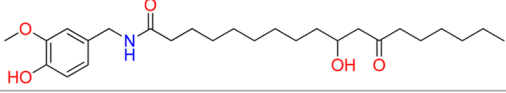
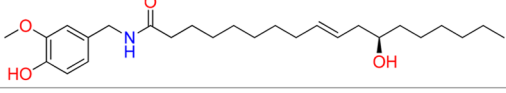
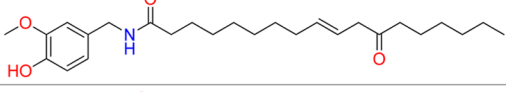
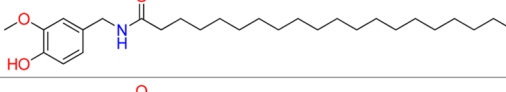
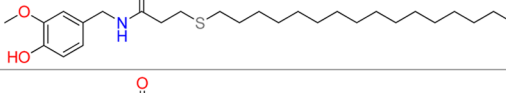
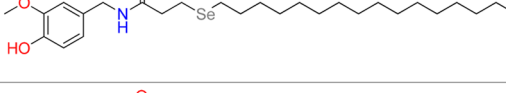
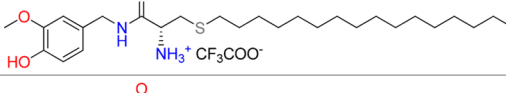
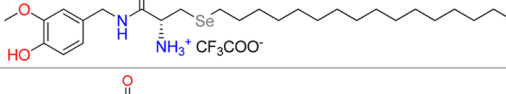
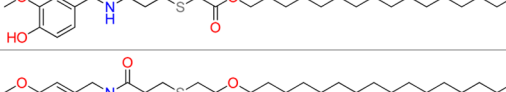
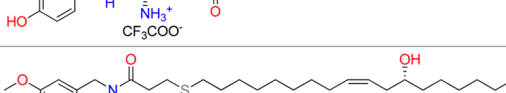
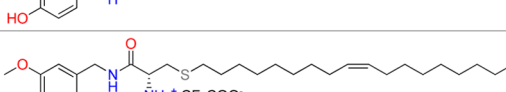
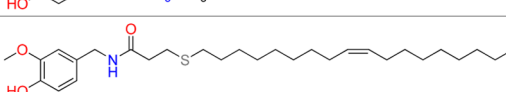
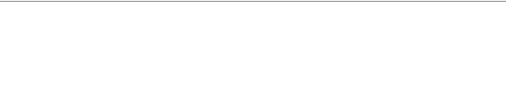
Caps-like	Structure	LPC	CBD
Palvanil (C16:0) ^a		>10	>10
Stevanil (C18:0)		>10	>10
Olvanil (C18:1)		0.16±0.02	1.7±0.1
Livanil (C18:2)		2.6±0.2	2.1±0.1
9 (C18:0)		>10	>10
12 (C18:1)		>10	7.5 ± 1.3
15 (C18:1)		>10	4.4 ± 0.3
Eicosavanillamide (C20:0)		3.1 ± 0.2	>10
39 (C19/S)		3.8 ± 0.8	nd ^b
46 (C19/Se)		4.3 ± 0.9	nd
55 (C19/S)		1.4 ± 0.2	nd
60 (C19/Se)		1.2 ± 0.03	nd
42 (C21/S/O)		>10	nd
57 (C21/S/O)		1.4 ± 0.1	nd
44 (C21/S:1)		>10	nd
56 (C21/S:1)		1.9 ± 0.1	nd
40 (C21/S:1)		2.5 ± 0.1	nd

Table 1. continued

Caps-like	Structure	LPC	CBD
45 (C21/S:1)		>10	nd
65 (C22:2)		0.82 ± 0.12	1.8 ± 0.3
41 (C22:2)		1.4 ± 0.07	2.8 ± 0.4
48 (C22:2)		1.4 ± 0.06	2.3 ± 0.1
64 (C22:1)		0.49 ± 0.07	1.5 ± 0.2
47 (C21/Se:1)		1.8 ± 0.01	3.2 ± 0.2
61 (C21/Se:1)		1.7 ± 0.01	0.98 ± 0.14
51 (C21/Se:1)		>10	2.3 ± 0.3
50 (C21/Se:1)		>10	1.4 ± 0.1

^aIn parentheses, number of C atoms in the alkyl chain followed by number of unsaturations. When heteroatom X occurs within alkyl chain, it is indicated as "/X". ^bnd: not determined.

Table 2. Antagonist Potency of Capsaicin-Imino Compounds at TRPV2 against LPC (3 μ M) and CBD (2 μ M), Reported as IC₅₀ (μ M)^a

Imino-caps	Structure	LPC	CBD
67 (16:0)		0.28 ± 0.04	6.0 ± 1.0
73 (18:1)		0.12 ± 0.01	3.0 ± 0.4

^aIn parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

profile between LPC and cannabidiol (CBD) (see Figure 2), we also investigated the effect of a representative panel of capsaicin derivatives against CBD to ascertain whether the inhibitory activity/potency would vary against agonists exhibiting different kinetics of action. Also in this case, the assays were conducted using a fluorometric assay with recombinant rat TRPV2 heterologously expressed HEK-293 cells. The preincubation (5 min) of TRPV2-HEK-293 cells with different concentrations of the tested compounds, followed by incubation with CBD (2 μ M), caused an inhibition of the Ca²⁺ elevation due to the TRPV2 response to CBD. The corresponding IC₅₀ values of the tested compounds are reported in Table 1. While the trend identified in LPC antagonism for capsaicin derivatives bearing all carbon atoms, selenium, or sulfur was substantially conserved, a

different behavior was observed with those derivatives featuring polar substituents (i.e., 50/51), since their activity against CBD was not negatively affected by these functional groups, as instead observed against LPC. The imino-derivatives 67 and 73 (see Table 2), i.e., the two most active compounds against LPC (0.28 and 0.12 μ M, respectively), were less potent against CBD (IC₅₀ = 6.0 and 3.0 μ M, respectively). The trend of activity of C16:0, C18:0, and C18:1 derivatives was similar to that observed for LPC, although C18:1 (olvanil) was less potent as an antagonist (IC₅₀ = 1.7 μ M), whereas, different from what observed with LPC, C20:0 was totally inactive. These results demonstrate a dependence of the antagonist activity on the type of agonist against which antagonism is tested.

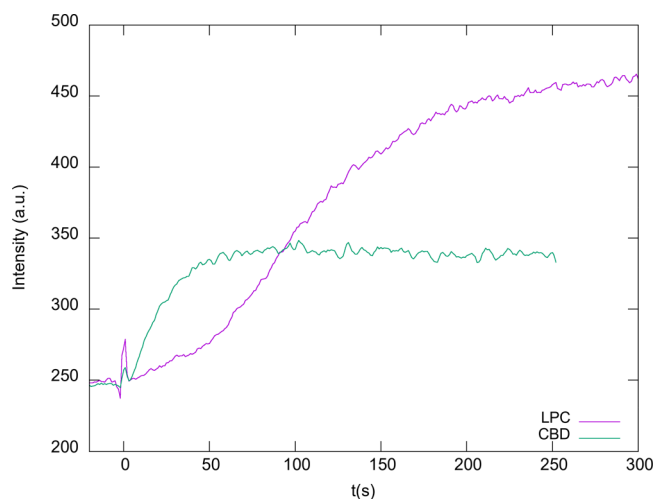


Figure 2. TRPV2 is activated by LPC (3 μM) and CBD (2 μM). The graph shows the representative traces of $[\text{Ca}^{2+}]_i$ increase evoked by the two agonists in HEK293 cells overexpressing TRPV2.

decided to ascertain the role of the head group of capsaicin, i.e., the vanillyl moiety, by testing a series of naturally occurring lipids bearing different polar heads and differing in length and unsaturation of the alkyl chain in order to determine the structural and functional requisites for TRPV2 modulation.

2.2.5. Long-Chain Ethanolamides Exhibit Differential Inhibition of TRPV2 upon Activation by LPC or CBD. To evaluate the contribution of the aromatic moiety to the overall activity, a panel of natural occurring ethanolamides differing in length and unsaturation degree was tested for both agonism and antagonism at TRPV2, using both LPC and CBD as reference activators. Ethanolamides share with the tested capsaicin derivatives the nature of both the alkyl chain and the hydrophilic groups (amide and hydroxyl moieties) in the polar head. The IC_{50} values (against CBD 2 μM and LPC 3 μM) are reported in Table 3. Ethanolamides featuring saturated alkyl chains, regardless of their lengths, were inactive against both agonists, whereas the introduction of a single double bond was sufficient to switch from inactivity to activity against both agonists (see PEA vs POEA, or SEA vs OEA), similar to what was already observed for capsaicin derivatives. However, while the C20:0 capsaicin derivative was active against LPC, the homolog ethanolamide was inactive. Moreover, while OEA was less active than the counterpart olvanil, LEA was more potent than livanil against both reference agonists. Increasing the

2.2.4. Evaluation of Endogenous Lipids as Potential TRPV2 Antagonists. Since the activity of the tested compounds appears to critically depend on the nature of alkyl chain but is less affected by changes in the polar head, we

Table 3. Potency of Fatty Ethanolamides as Functional Antagonists at TRPV2 against LPC (3 μM) and CBD (2 μM), Reported as IC_{50} (μM)

Ethanolamides	Structure	LPC	CBD
PEA ^a (C16:0) ^b		>10	>10
POEA (C16:1)		3.5 \pm 0.01	1.7 \pm 0.1
SEA (C18:0)		>10	>10
OEA (C18:1)		1.8 \pm 0.1	5.4 \pm 0.2
LEA (C18:2)		1.4 \pm 0.1	0.65 \pm 0.07
Arachidoyl-EA (C20:0)		>10	>10
AEA (C20:4)		6.6 \pm 0.1	0.96 \pm 0.09
EPEA (C20:5)		>10	2.3 \pm 0.2
Docosaenoyl-EA (C22:1)		0.74 \pm 0.02	>10
DHEA (C22:6)		>10	1.6 \pm 0.1

^aAbbreviations: EA, ethanolamide; PEA, palmitoyl ethanolamide; POEA, palmitoleoyl ethanolamide; OEA, oleoyl ethanolamide; LEA, lynoleoyl ethanolamide; arachidonyl ethanolamide; EPEA, eicosapentaenoyl ethanolamide; DHEA, docosahexaenoyl ethanolamide. ^bIn parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

Table 4. Antagonist Potency of Fatty Amides at TRPV2 against LPC (3 μ M) and CBD (2 μ M), Reported as IC₅₀ (μ M)

Amides	Structure	LPC	CBD
PA ^a (C16:0) ^b		>10	>10
SA(C18:0)		>10	>10
OA (C18:1)		2.1 \pm 0.1	2.1 \pm 0.2
LA (C18:2)		2.2 \pm 0.1	1.2 \pm 0.1
ErA (C22:1)		0.67 \pm 0.13	7.1 \pm 0.7
Eicosanamide (C20:0)		>10	>10

^aAbbreviations: PA, palmitamide; SA, stearamide; OA, oleamide; LA, linoleamide; ErA, erucamide. ^bIn parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

Table 5. Lack of Strong Antagonist Activity of Fatty Acids at TRPV2 against LPC (3 μ M) and CBD (2 μ M), Reported as IC₅₀ Values (μ M)

Acids	Structure	LPC	CBD
Palmitic acid (C16:0) ^a		>10	>10
Oleic acid (C18:1)		>10	>10
Arachidic acid (C20:0)		>10	>10
Arachidonic acid (C20:4)		>10	>10
Erucic acid (C22:1)		>10	>10
Docosadienoic acid (C22:2)		>10	>10

^aIn parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

Table 6. Slope Values from Linear Regression of Schild Analysis and *t*-Test Statistics

compd	LPC			CBD		
	slope ^a	N ^b	P ^c	slope ^a	N ^b	P ^c
61	−0.58 \pm 0.087	4	<0.0024	−0.74 \pm 0.048	4	<0.002
olvanil	−0.77 \pm 0.049	6	<0.001	−0.55 \pm 0.068	6	<0.001
docosaenoyl-EA	−0.54 \pm 0.046	6	<0.001			
50				−0.63 \pm 0.039	5	<0.001

^aMean value \pm standard deviation. ^bNumber of experiments (each one performed at least in triplicate) used for Schild regression. ^cP values calculated from *t* test values for the “slope = 1 hypothesis”.

number of double bonds increased the potency against CBD but not LPC.

2.2.6. Long-Chain Primary Amides Exhibit Differential Inhibition of TRPV2 Channels upon Activation by LPC or CBD. To also evaluate the role of the hydroxyl group, we tested a series of amide derivatives. As for capsaicin- and ethanolamine-derivatives, also for the amides the activity strongly depended upon the presence of at least one double bond. In particular, erucamide is active as TRPV2 antagonist with a potency comparable to that of its capsaicin derivative

(0.67 vs 0.49 μ M) against LPC, but it is less potent than the capsaicin counterpart against CBD (7.1 vs 1.5 μ M). As observed with the ethanolamides, also the C20:0 amide derivative was inactive against both activators (Table 4).

2.2.7. Free Fatty Acids Are Poor Inhibitors of TRPV2 Channels. Finally, to investigate the role of the amide group, we tested against both LPC and CBD a panel of long-chain fatty acids, featuring alkyl chains comparable with those occurring in the already-tested compounds. The results are reported in Table 5. Fatty acids with alkyl chains from C16 up

384 to C22 are by far less potent antagonists against both reference
385 agonists than the other classes of compounds bearing similar
386 alkyl chains, thus suggesting that the amide group is mandatory
387 for potent antagonism.

2.2.8. Schild Analysis on Selected TRPV2 Antagonists.

389 The effects of increasing concentrations of antagonist **61**,
390 olvanil, and docosaenoyl-EA vs LPC and **61**, olvanil, and **50** vs
391 CBD were tested against concentration–response curves of
392 LPC and CBD (where the effects of each concentration of
393 LPC and CBD were expressed as percent of their effect of $2 \times$
394 10^{-4} M in the absence of the antagonist) to calculate Schild's
395 plots. These compounds have been selected as representative
396 of antagonists active either against both activators (**61**, olvanil)
397 or selectively toward LPC (docosaenoyl-EA)/CBD (**50**) alone.
398 In all cases, the plots analyzed by linear regression gave slope
399 values significantly less than unity, as reported in Table 6,
400 indicative of a noncompetitive behavior. However, this result
401 may also be indicative of a nonequilibrium condition, and we
402 do not definitely rule out a competitive behavior.

3. DISCUSSION

403 Novel capsaicin derivatives, initially designed as TRPV1
404 agonists, behave as potent TRPV2 antagonists. The different
405 types of modifications introduced in these compounds
406 determine different agonist/antagonist profiles and, in
407 particular, opposite behaviors in terms of relative potency/
408 efficacy within a derivative series on the two channels. In fact,
409 the insertion of a positive charge or an imido group close the
410 amido group, detrimental for TRPV1 agonism, is well-tolerated
411 for TRPV2 antagonism and even leads in some cases to an
412 increment or a rescue of activity. Conversely, the insertion of a
413 sulfur/selenium atom and/or the presence of a polar group,
414 which increases TRPV1 agonism, leaves unaffected, or even
415 decrease, TRPV2 antagonism.

416 Given the scarcity of known endogenous ligands for TRPV2,
417 the discovery of such long-chain capsaicin derivatives as potent
418 TRPV2 antagonists prompted us to investigate the following
419 classes of long-chain fatty acid derivatives with at least one
420 functional group in common with capsaicin derivatives as
421 potential TRPV2 modulators: (i) ethanolamides, (ii) primary
422 amides, and (iii) free fatty acids, to evaluate the role of the
423 amide group itself. Antagonists were found in both the
424 ethanolamide and primary amide, but not in fatty acid, series.

425 Activities for both synthetic and endogenous ligands were
426 tested against either LPC or CBD as activators, since, on the
427 basis of their different kinetics of activation, CBD can be
428 defined as a direct TRPV2 agonist, whereas LPC induces
429 TRPV2 activation indirectly, via its G-protein-coupled
430 receptors and PI3,4 kinase mediated pathways.²¹ We found
431 that this different mode of activation is differentially counter-
432 acted by the investigated compounds, which can be classified
433 as follows: (a) compounds endowed with similar antagonist
434 efficacy against both agonists, (b) compounds selectively active
435 against LPC, (c) compounds selectively active against CBD.
436 To determine the nature of antagonism, a Schild regression
437 was carried out for the representative members of each class,
438 i.e., olvanil, docosaenoyl-EA, and compound **50**, and in all
439 three cases the antagonists behaved as noncompetitive ligands,
440 suggesting that these compounds may act as allosteric
441 antagonists. However, we cannot completely rule out a
442 competitive behavior since a Schild plot slope of <1 may
443 also suggest nonequilibrium conditions. Moreover, since the
444 hydrophobicity of the alkyl chain of the investigated

compounds is a critical requisite for LPC but not for CBD 445
inhibition, it is reasonable to speculate that a different binding 446
site is involved in LPC antagonism, with structural/functional 447
requisites different from those of CBD. This site might be 448
either on TRPV2 or on other targets activated by LPC in its 449
signaling cascade and would be the target of those compounds 450
selectively antagonizing activation by LPC. A common critical 451
requisite for activity of both ethanolamides and amides as 452
TRPV2 antagonists is the occurrence of at least one double 453
bond in the alkyl chain, since saturated lipids, regardless of the 454
length of their acyl chains, are totally inactive. This suggests 455
that a bent conformation of the alkyl chain is required for a 456
better accommodation into the active site, as previously 457
reported for other TRPV1 agonists.⁵⁹ Also C16:0 and C18:0 458
derivatives of capsaicin are inactive against both CBD and 459
LPC, whereas the C20:0 derivative is selectively active against 460
LPC. Instead, a different behavior is observed with imino- 461
capsaicin derivatives since they are active also when bearing 462
saturated alkyl chain. The aromatic moiety contributes to the 463
overall activity at TRPV2 of the compounds characterized in 464
the present work, since it occurs in the most active antagonists. 465

4. CONCLUSIONS

466 In summary, the search for structurally related synthetic or 467
endogenous lipids with structural similarity to capsaicin 468
derivatives led to identification of olvanil and **73** as potent 469
TRPV2 antagonists against LPC (0.16 and 0.12 μ M, 470
respectively) and of LEA (linoleoylethanolamide) as potent 471
TRPV2 antagonist against CBD (0.65 μ M). This finding is 472
both surprising, since all other synthetic and endogenous 473
compounds tested here on TRPV2 behave as antagonists and 474
capsaicin is inactive at this channel, and of great physiological 475
importance, since novel potent endogenous antagonists were 476
been identified following this study.

477 In conclusion, starting from the testing of a series of 478
synthetic capsaicinoids as modulators of rat TRPV2, we 479
discovered not only new tools for the pharmacological 480
manipulation of the latter but also that previously described 481
endogenous lipids, i.e., long chain fatty acid ethanolamides and 482
primary amides, behave as negative modulators of this channel. 483
These data are of great potential importance given the 484
increasingly important role assigned to TRPV2 in temperature 485
sensing, pain, insulin secretion, immune response, muscle and 486
heart function, and cancer.⁵⁸

5. EXPERIMENTAL SECTION

5.1. **Compounds.** Stevanil, livanil, ethanolamides, amides, and 487
fatty acids when not described in the synthetic section have been 488
purchased from Cayman-Vinci Biochem. Palvanil and PEA are kind 489
gifts from Epitech Group SpA, Saccolongo, Padova, Italy, whereas 490
olvanil is a precious gift from Dr. Alberto Minassi, Dipartimento di 491
Scienze del Farmaco, Università del Piemonte Orientale, Novara, 492
Italy. 493

5.2. **Synthetic Procedures.** Reactions requiring anhydrous 494
conditions were performed in flamed or oven-dried glassware using 495
anhydrous solvents and under inert atmosphere (argon). The solvents 496
and reagents were purchased from Acros Organics, Sigma-Aldrich, 497
Fluka, Merk, Panreac, Strem Chemicals, or TCI Chemicals. 498
Petroleum ether, EtOAc, DCM, and MeOH were used without 499
further purification. In the case of anhydrous reactions, solvent and 500
reagents were properly dried. Acrolein was distilled at atmospheric 501
pressure and used immediately. The reactions were monitored until 502
completion by TLC on silica gel 60F-254 precoated plates (Merck). 503
Visualization of the compounds was performed by UV light (254 nm), 504

505 and staining was performed by immersion in a 5% solution of
506 concentrated H₂SO₄ in methanol or 5% w/v phosphomolibdic acid in
507 ethanol followed by heating. Flash column chromatography was
508 performed using silica gel (technical grade, 60 Å, 40–63 µm) (Sigma-
509 Aldrich) under air pressure. NMR spectra were recorded on a
510 MERCURYplus AS400 MHz Varian spectrometer. Chemical shifts
511 are reported in parts per million (ppm, δ units). Coupling constants
512 (J) are reported and expressed in hertz (Hz). Spitting patterns are
513 designated as br (broad), s (singlet), d (doublet), dd (double
514 doublet), t (triplet), q (quartet), dt (double triplet), td (triple
515 doublet), ddd (double double doublet), p (pentuplet), and m
516 (multiplet). All ¹³C NMR spectra were proton decoupled. High
517 resolution mass spectra (HR-MS) were recorded on at the Serveis
518 Científics de l'Universitat de Lleida (SCT-UdL) and Servei de
519 Recursos Científics i Tècnics de Universitat Rovira i Virgili (URV)
520 with an Agilent G6510AA Q-TOF MS spectrometer in positive
521 electrospray ionization (ESI⁺) and Agilent LC1200 series coupled to
522 MS6210 TOF spectrometer in electrospray ionization (ESI⁺)
523 respectively. Mobile phase was composed of ACN/MeOH 50:50.
524 Flow rate: 0.6 mL/min. Infrared spectra were recorded on Jasco FT-
525 IR 6300 using a diamond ATR crystal cell. Melting points were
526 measured using Gallenkamp capillary apparatus and are uncorrected.
527 Optical rotations were measured at 20 °C with a PerkinElmer 241 nc
528 polarimeter (λ = 589 Na, path length 1 dm). Some recorded values
529 were within the error limit of the polarimeter, and therefore it was not
530 possible to determine them. It has been indicated as [α]_D²⁰ < 1°.
531 Analytical UPLC–MS was performed on a binary Acquity UPLC with
532 a Acquity PDA UPLC eLambda 800 nm triple quadrupole mass
533 spectrometer (Xevo TQ-S) using a Acquity UPLC BEH C18 50 × 2.1
534 mm, 1.7 µm C18 column. UV detection = 210–500 nm, mass
535 spectrometry = ESI⁺ (scan 100–850 m/z). Flow rate was 0.3 mL/min
536 using a solvent gradient of B 100% over 6 min (total run time with
537 equilibration back to starting conditions = 2 min) where A = MeOH
538 and B = 85/15/0.2 MeOH/H₂O/AcOH. Purities were measured by
539 UV absorption at 254 nm or TIC and are ≥95% unless otherwise
540 stated. Purity of final compounds was assessed by reversed-phase
541 UHPLC with UV diode array detection; all tested compounds were
542 >95% pure.

543 **5.2.1. Procedure I. Amine Bond Formation.** To a 0.35 M
544 solution of starting material in anhydrous DMF were added the amine
545 3 (1.1 equiv), HATU (1.5 equiv), and DIPEA (3 equiv). The mixture
546 was stirred at room temperature for 20 h. To the mixture were added
547 EtOAc and brine, and the aqueous phase was extracted with EtOAc.
548 The combined organic phases were washed with 1 M HCl, saturated
549 solution of NaHCO₃ and brine. The organic phase was dried over
550 anhydrous Na₂SO₄, filtered and the solvent was removed under
551 reduced pressure. The crude residue was purified by silica gel column
552 chromatography.

553 **5.2.2. Procedure II. Ester Hydrolysis.** To a 0.2 M solution of
554 starting material in THF/H₂O (1:1) LiOH·H₂O (3 equiv) was added.
555 The mixture was stirred at room temperature until completion of the
556 reaction. The reaction mixture was acidified with 1 M HCl until pH 1
557 and extracted with EtOAc. The organic phase was dried over
558 anhydrous Na₂SO₄, filtered, and the solvent was removed under
559 reduced pressure to afford the corresponding compound.

560 **5.2.3. Procedure III. Boc Protection.** Et₃N (1.5 equiv) was
561 added to a 0.3 M aqueous solution of starting material, cooled in an
562 ice bath. Then Boc₂O (1.5 equiv) was added dropwise and stirred
563 overnight. After completion of the reaction, the solvent was
564 evaporated under reduced pressure. The residue was dissolved in
565 EtOAc, washed with 1 M HCl and brine, dried over anhydrous
566 Na₂SO₄, filtered, and evaporated under reduced pressure. The crude
567 residue was thoroughly washed with hexane several times.

568 **5.2.4. Procedure IV. SS/SeSe Bond Cleavage. SS Bond**
569 **Cleavage.** To a 0.15 M solution of starting material in wet THF
570 was added tri-*n*-butylphosphine (P(^{*n*}Bu)₃) (1.05 equiv). The reaction
571 mixture was stirred at room temperature for 2 h. After completion of
572 the reaction, the solvent was removed under reduced pressure to
573 afford the crude product, which was purified by silica gel column
574 chromatography.

SeSe Bond Cleavage and Se-Alkylation. To a 0.13 M solution of 575
starting material in ethanol was added NaBH₄ (2.5 equiv) at 0 °C. 576
The reaction mixture was stirred for 20 min, followed by addition of 577
the respective iodinated compound. The reaction mixture was stirred 578
at room temperature for 16 h. Then, the reaction was quenched with 579
1 M HCl and extracted with EtOAc. The organic phase was dried 580
over anhydrous Na₂SO₄, filtered, and the solvent was removed under 581
reduced pressure. The crude residue was purified by silica gel column 582
chromatography. 583

5.2.5. Procedure V. Reduction of Methyl Ester. To a 0.2 M 584
solution of starting material in anhydrous THF, LiAlH₄ (2 equiv) was 585
added at 0 °C. The reaction mixture was stirred at room temperature 586
for 24 h. Then, the reaction was quenched with 1 M HCl, followed by 587
extraction with DCM. The combined organic phases were dried over 588
anhydrous Na₂SO₄, filtered, and the solvent was removed under 589
reduced pressure. The solid residue was purified by silica gel column 590
chromatography. 591

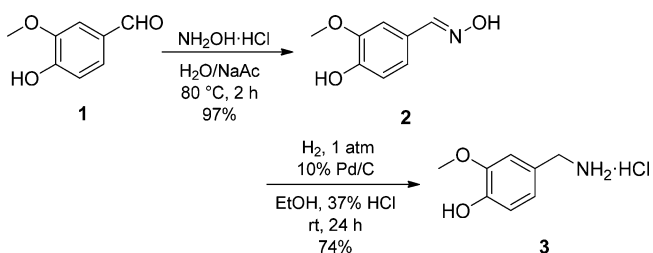
5.2.6. Procedure VI. Iodination. To a 0.25 M solution of starting 592
material in toluene iodine (1.2 equiv), imidazole (3 equiv) and PPh₃ 593
(1.2 equiv) were added. The mixture was stirred at 90 °C for 2 h. 594
The solvent was evaporated under reduced pressure. The residue was 595
dissolved in EtOAc, washed with saturated aqueous solution of 596
KMnO₄, water, and brine, dried over anhydrous Na₂SO₄, filtered, and 597
evaporated under reduced pressure. The solid residue was purified by 598
silica gel column chromatography. 599

5.2.7. Procedure VII. S-Alkylation. To a 0.2 M solution of 600
starting material in DMF, TEA (1.5 equiv) and the corresponding 601
iodinated compound (1.12 equiv) were added. The reaction mixture 602
was stirred at 90 °C overnight. To the mixture were added EtOAc and 603
brine, and the aqueous phase was extracted with EtOAc. The 604
combined organic phases were washed with 1 M HCl, saturated 605
solution of NaHCO₃, and brine. The organic phase was dried over 606
anhydrous Na₂SO₄, filtered, and the solvent was removed under 607
reduced pressure. The crude residue was purified by silica gel column 608
chromatography. 609

5.2.8. Procedure VIII. TBDMS Deprotection. A 0.25 M solution 610
of the starting material in a mixture of AcOH/THF/H₂O was stirred 611
at room temperature until deprotection was complete. The solvent 612
was evaporated under reduced pressure to obtain the reaction crude, 613
which was purified by silica gel column chromatography. 614

5.2.9. Procedure IX. Boc Deprotection. To a 0.3 M solution of 615
starting material in DCM, TFA (10 equiv) was added. The reaction 616
mixture was stirred for 1 h, followed by removal of the solvent under 617
nitrogen stream and drying in vacuo to afford the trifluoroacetate salt 618
of the compound. 619

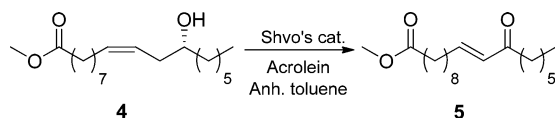
5.2.10. Procedure X. Base Schiff Formation. To a 0.03 M 620
solution of starting material in MeOH, vanillin 1 (1 equiv) was added. 621
The mixture was refluxed for 2 h in the presence of small amount of 622
glacial AcOH. After cooling, the reaction mixture was filtered to 623
recover a solid, which was recrystallized from hot MeOH to afford the 624
corresponding compound. 625 g



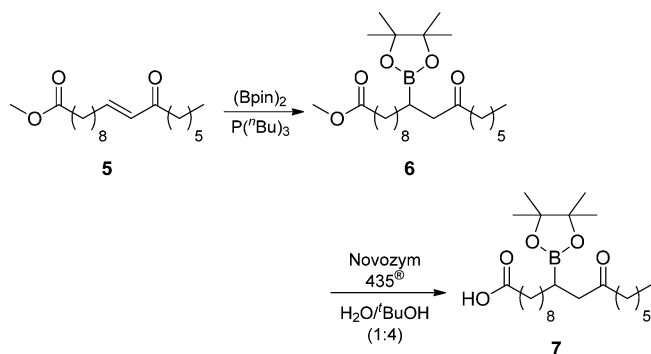
(E)-4-Hydroxy-3-methoxybenzaldehyde Oxime (2). Hydrox- 626
ylamine hydrochloride (2.37 g, 34.0 mmol) in H₂O (10 mL) and 627
sodium acetate trihydrate (4.48 g, 32.9 mmol) in H₂O (10 mL) were 628
successively added to a solution of vanillin 1 (5.00 g, 32.9 mmol) in 629
H₂O (30 mL). The reaction mixture was stirred at 80 °C for 2 h. The 630
reaction mixture was extracted with EtOAc, and the organic layer was 631
dried over anhydrous Na₂SO₄ and filtered. The solvent was 632
evaporated under reduced pressure to yield the oxime 2¹ (5.26 g, 633

634 97%) as an off-white solid. Mp = 118–119 °C. IR (ATR) ν = 3444, 3213, 3008, 2941, 1596, 1513, 1428, 1027, 969 cm^{-1} . ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ = 3.77 (s, 3H, CH_3O), 6.77 (d, 1H, J = 8.1 Hz, H_{Ar}), 6.97 (dd, 1H, J = 8.1, 2.0 Hz, H_{Ar}), 7.16 (d, 1H, J = 2.0 Hz, H_{Ar}), 7.99 (s, 1H, $\text{CH}=\text{N}$), 9.33 (s, 1H, OH), 10.84 (s, 1H, N-OH). ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ = 55.50 (CH_3O), 109.21 (C_{Ar}), 115.49 (C_{Ar}), 120.52 (C_{Ar}), 124.47 (CCHN), 147.85 (COH), 148.01 (CCH_3O), 148.10 ($\text{CH}=\text{N}$).

642 **4-Hydroxy-3-methoxybenzylamine Hydrochloride (3)**. 37% HCl (20 mL, 0.26 mol) and Pd/C (10 wt % loading) (20% w/w, 1.05 g) were added to a solution of **2** (5.2 g, 0.03 mol) in EtOH (150 mL). The reaction mixture was hydrogenated at 1 atm at room temperature for 24 h. The reaction mixture was filtered over Celite, and the solvent volume was reduced under pressure. The residue was crystallized from EtOAc and filtered to yield the amine hydrochloride salt **3**² (4.2 g, 74%) as a white solid. Mp = 219–222 °C. IR (ATR) ν = 3112, 3024, 2805, 1763, 1377, 1033, 828, 670 cm^{-1} . ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ = 3.77 (s, 3H, CH_3O), 3.83–3.90 (m, 2H, CH_2NH_2), 6.79 (d, 1H, J = 8.1 Hz, H_{Ar}), 6.85 (dd, 1H, J = 8.1, 2.0 Hz, H_{Ar}), 7.18 (d, 1H, J = 2.0 Hz, H_{Ar}), 8.40 (br, s, 3H, NH_2 , HCl), 9.19 (s, 1H, OH). ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ = 42.19 (CH_2NH_2), 55.70 (CH_3O), 113.45 (C_{Ar}), 115.27 (C_{Ar}), 121.74 (C_{Ar}), 124.64 (CCHN), 146.81 (COH), 147.51 (CCH_3O).



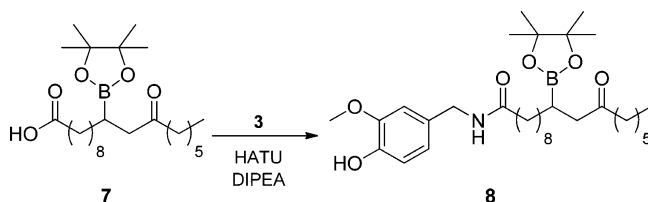
657 **Methyl 12-Oxo-10-(10E)-enoate (5)**. Shvo's catalyst (9 mg, 8 μmol) and acrolein freshly distilled (390 μL , 4.80 mmol) were added to a solution of methyl ricinoleate **4** (500 mg, 1.60 mmol) in anhydrous toluene (15 mL). The reaction mixture was purged with N_2 and stirred under reflux for 45 min. The solvent was evaporated under reduced pressure, and after the purification by silica gel column chromatography (petroleum ether/ Et_2O 95:5) the enone **5**³ (348 mg, 70%) was obtained as a yellowish oil. R_f = 0.50 (petroleum ether/ Et_2O 9:1). IR (ATR) ν = 2927, 2855, 1736, 1709, 1436, 1195, 1169, 1104, 979, 880, 752 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.86 (t, 3H, J = 6.9 Hz, CH_3), 1.23–1.33 (m, 14H, CH_2), 1.38–1.48 (m, 2H, CH_2), 1.52–1.65 (m, 4H, CH_2), 2.18 (q, 2H, J = 6.4 Hz, CH_2), 2.29 (t, 2H, J = 6.9 Hz, CH_2), 2.51 (t, 2H, J = 6.9 Hz, COCH_2), 3.65 (s, 3H, CH_3O), 6.07 (dt, 1H, J = 15.9, 1.5 Hz, $\text{CH}=\text{CH}$), 6.80 (dt, 1H, J = 15.9, 6.9 Hz, $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.01 (CH_3), 22.48 (CH_2), 24.27 (CH_2), 24.86 (CH_2), 28.04 (CH_2), 28.96 (CH_2), 29.07 ($4 \times \text{CH}_2$), 31.59 (CH_2), 32.38 (CH_2), 34.02 (CH_2), 40.08 (COCH_2), 51.41 (CH_3O), 130.28 ($\text{CH}=\text{CH}$), 147.20 ($\text{CH}=\text{CH}$), 174.24 (COO), 200.99 (COCH_2).



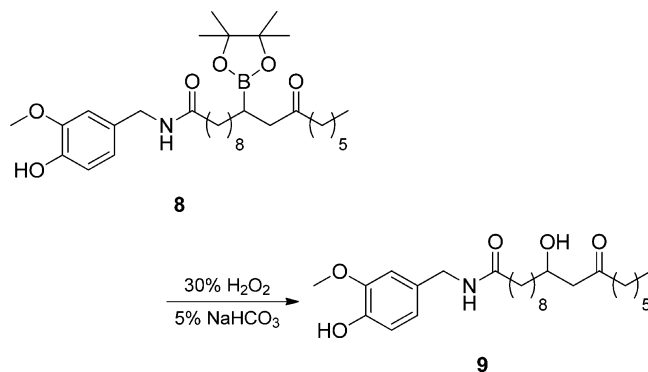
676 **Methyl 12-Oxo-10-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)octadecanoate (6)**. Tri-*n*-butylphosphine (26 μL , 0.10 mmol) was added to a solution of anhydrous CuCl (10 mg, 0.10 mmol) in anhydrous DMF (4.5 mL) under argon atmosphere. In another reaction vessel, bis(pinacolato)diboron (283 mg, 1.12 mmol) was added to a solution of methyl 12-oxooctadec-10E-enoate **5** (290 mg, 0.93 mmol) in anhydrous DMF (4.5 mL) under argon

atmosphere. This solution was transferred to the tri-*n*-butylphosphine solution. The reaction mixture was stirred at room temperature for 48 h. The crude was taken up in H_2O and extracted with petroleum ether. The organic solution was dried over anhydrous Na_2SO_4 , filtered and the solvent was evaporated under reduced pressure to yield the β -boron ketone **6** (190 mg, 46%) as a yellow oil after the purification by silica gel column chromatography (petroleum ether/ EtOAc 95:5). R_f = 0.49 (petroleum ether/ Et_2O 9:1). ^1H NMR (400 MHz, CDCl_3) δ = 0.84 (t, 3H, J = 6.9 Hz, CH_3), 1.18–1.28 (m, 30H, $(\text{CH}_3)_4$, CH_2), 1.34–1.39 (m, 1H, CHB), 1.49–1.60 (m, 4H, CH_2), 2.27 (t, 2H, J = 6.9 Hz, CH_2), 2.33 (td, 2H, J = 7.4, 3.7 Hz, COCH_2), 2.50 (d, 2H, J = 6.8 Hz, CHBCH₂CO), 3.64 (s, 3H, CH_3O).

695 **12-Oxo-10-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)octadecanoic Acid (7)**. Novozym 435 (83 mg, 50% w/w) was added to a solution of the methyl ester **6** (190 mg, 0.43 mmol) in a mixture of H_2O (308 μL) and *tert*-BuOH (922 μL). The reaction mixture was stirred at 45 °C for 24 h. The mixture was filtered and the solvent was evaporated under reduced pressure to yield the acid **7** (180 mg, quantitative) as a yellow oil. ^1H NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.20–1.34 (m, 30H, $(\text{CH}_3)_4$, CH_2), 1.38–1.44 (m, 1H, CHB), 1.51–1.58 (m, 2H, CH_2), 1.59–1.66 (m, 2H, CH_2), 2.30–2.40 (m, 4H, CH_2 , COCH_2), 2.53 (d, 2H, J = 6.8 Hz, CHBCH₂CO).

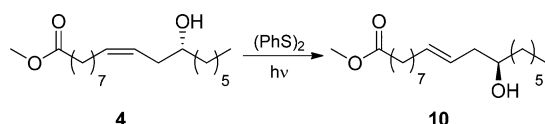


706 **N-(4'-Hydroxy-3'-methoxybenzyl)-12-oxo-10-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)octadecanamide (8)**. General procedure I was applied to a solution of the acid **7** (175 mg, 0.41 mmol) dissolved in anhydrous DMF (6 mL), amine hydrochloride salt **3** (69 mg, 0.45 mmol), DIPEA (200 μL , 1.24 mmol), and HATU (235 mg, 0.62 mmol). The amide **8** was obtained (125 mg, 54%) as a brown oil after the purification by silica gel flash column chromatography (petroleum ether/ EtOAc 6:4). R_f = 0.55 (petroleum ether/ EtOAc 3:7). ^1H NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.7 Hz, CH_3), 1.21–1.31 (m, 30H, $(\text{CH}_3)_4$, CH_2), 1.35–1.41 (m, 1H, CHB), 1.52–1.57 (m, 2H, CH_2), 1.61–1.67 (m, 2H, CH_2), 2.18 (t, 2H, J = 6.9 Hz, CH_2), 2.32–2.39 (m, 2H, COCH_2), 2.52 (d, 2H, J = 6.7 Hz, CHBCH₂CO), 3.88 (s, 3H, CH_3O), 4.35 (d, 2H, J = 5.6 Hz, CH_2NH), 5.64–5.71 (m, 1H, CH_2NH), 6.82 (ddd, 3H, J = 12.5, 9.9, 5.5 Hz, H_{Ar}).

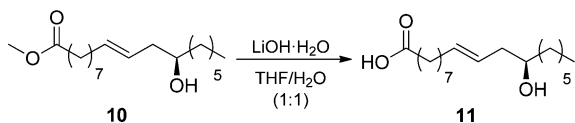


721 **N-(4'-Hydroxy-3'-methoxybenzyl)-10-hydroxy-12-oxooctadecanamide (9)**. A volume of 5% w/v NaHCO_3 (2.5 mL, 1.49 mmol) was added to a solution of compound **8** (125 mg, 0.22 mmol) and 2.5 mL of 30% H_2O_2 (0.02 mmol). The reaction mixture was stirred at room temperature for 24 h. Saturated aqueous $\text{Na}_2\text{S}_2\text{O}_4$ (0.25 mL) was added to decompose any remaining peroxide keeping the temperature below 40 °C. The reaction mixture was diluted with H_2O and extracted with EtOAc. The organic solution was dried over

729 anhydrous Na_2SO_4 and filtered. The solvent was evaporated under
 730 reduced pressure to yield the β -hydroxy ketone **9** (75 mg, 76%) as a
 731 rosaceous solid after the recrystallization from Et_2O . Mp = 73–75 °C.
 732 IR (ATR) ν = 3318, 2912, 2849, 1705, 1638, 1513, 1267, 1240, 1122,
 733 718 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.88 (t, 3H, J = 6.9 Hz,
 734 CH_3), 1.20–1.41 (m, 18H, CH_2), 1.40–1.50 (m, 2H, CH_2), 1.52–
 735 1.60 (m, 2H, CH_2), 1.60–1.68 (m, 2H, CH_2), 2.18 (t, 2H, J = 6.9 Hz,
 736 CH_2), 2.41 (t, 2H, J = 6.9 Hz, COCH_2), 2.46–2.52 (m, 1H,
 737 $\text{CHCH}_{11a}\text{CO}$), 2.59 (dd, 1H, J = 17.3, 1.8 Hz, $\text{CHCH}_{11b}\text{CO}$), 3.08
 738 (br s, 1H, CHOH), 3.87 (s, 3H, CH_3O), 3.94–4.05 (m, 1H, CHOH),
 739 4.35 (d, 2H, J = 5.7 Hz, CH_2NH), 5.69 (br s, 2H, OH, CH_2NH),
 740 6.67–6.88 (m, 3H, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.16
 741 (CH_3), 22.61 (CH_2), 23.73 (CH_2), 25.53 (CH_2), 25.87 (CH_2), 28.97
 742 (CH_2), 29.34 (CH_2), 29.35 (CH_2), 29.48 (CH_2), 29.55 (CH_2), 31.70
 743 (CH_2), 36.52 (CH_2), 36.96 (CH_2), 43.66 (CH_2NH), 43.84
 744 (COCH_2), 49.06 (CHCH_2CO), 56.08 (CH_3O), 67.77 (CHOH),
 745 110.85 (C_{Ar}), 114.53 (C_{Ar}), 120.93 (C_{Ar}), 130.56 (C_{Ar}), 145.25 (C_{Ar}),
 746 146.84 (C_{Ar}), 172.99 (NHCO), 212.84 (COCH_2). HR-MS (ESI^+),
 747 m/z : $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{26}\text{H}_{43}\text{NO}_5\text{Na}$ 472.3033; found 472.3042.

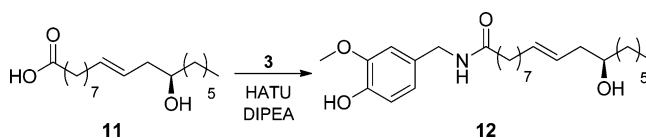


748 **Methyl (12R)-Hydroxyoctadec-(9E)-enoate (10).** Diphenyl
 749 disulfide (56 mg, 0.26 mmol) was added to a solution of methyl
 750 ricinoleate **4** (4 g, 12.8 mmol) in isooctane (120 mL). The reaction
 751 mixture was placed in a photochemical reactor and irradiated for 3 h
 752 with a Philips HP(L) 400 W medium-pressure mercury lamp. After
 753 irradiation the solvent was removed under reduced pressure and the
 754 crude reaction mixture was dissolved in hot petroleum ether (185
 755 mL). The filtrate was cooled at –30 °C, and after 48 h a white solid
 756 appeared. This solid was quickly filtered and recovered at –30 °C to
 757 yield the compound **10**⁴ (1.49 g, 37%) as a yellowish oil at room
 758 temperature. IR (ATR) ν = 3431, 2924, 2854, 1740, 1435, 1197,
 759 1171, 969, 860, 724 cm^{-1} . $[\alpha]_{\text{D}}^{20}$ –0.2° (c 2.44, CHCl_3). ^1H NMR
 760 (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.23–1.39 (m,
 761 16H, CH_2), 1.39–1.48 (m, 3H, CH_2), 1.56–1.71 (m, 2H, CH_2),
 762 1.97–2.09 (m, 3H, CH_2 , H_{11a}), 2.18–2.26 (m, 1H, H_{11b}), 2.29 (t, 2H,
 763 J = 6.9 Hz, CH_2), 3.53–3.61 (m, 1H, CHOH), 3.65 (s, 3H, CH_3O),
 764 5.47–5.56 (m, 1H, CHCH), 5.47–5.56 (m, 1H, CHCH). ^{13}C NMR
 765 (101 MHz, CDCl_3) δ = 14.22 (CH_3), 22.75 (CH_2), 25.05 (CH_2),
 766 25.79 (CH_2), 29.06 (CH_2), 29.20 (CH_2), 29.22 (CH_2), 29.49 (2 ×
 767 CH_2), 31.97 (CH_2), 32.75 (CH_2), 34.22 (CH_2), 36.88 (CH_2), 40.85
 768 (CHCH_2CHO), 51.57 (CH_3O), 71.06 (CHOH), 126.07 (CHCH),
 769 134.69 (CHCH), 174.44 (COO^-).

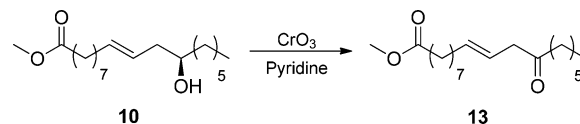


770 **(12R)-Hydroxyoctadec-(9E)-enoic Acid (11).** General proce-
 771 dure II was applied to a solution of compound **10** (200 mg, 0.64
 772 mmol) dissolved in THF/ H_2O (3 mL, 1:1) and $\text{LiOH}\cdot\text{H}_2\text{O}$ (46 mg,
 773 1.92 mmol) to yield the fatty acid **11**⁵ (150 mg, 78%) as a yellowish
 774 solid after a recrystallization in hot petroleum ether. Mp = 49–51 °C.
 775 $[\alpha]_{\text{D}}^{20}$ +6.6° (c 1, EtOH). IR (ATR) ν = 3321, 3221, 3040, 2955, 2916,
 776 2848, 1690, 1466, 1072, 959, 720, 682 cm^{-1} . ^1H NMR (400 MHz,
 777 CDCl_3) δ = 0.88 (t, 3H, J = 6.9 Hz, CH_3), 1.22–1.40 (m, 16H, CH_2),
 778 1.40–1.50 (m, 4H, CH_2), 1.58–1.68 (m, 2H, CH_2), 1.97–2.11 (m,
 779 3H, CH_2 , H_{11a}), 2.18–2.28 (m, 1H, H_{11b}), 2.33 (t, 2H, J = 6.9 Hz,
 780 CH_2), 3.54–3.63 (m, 1H, CHOH), 5.33–5.46 (m, 1H, CHCH),
 781 5.45–5.58 (m, 1H, CHCH). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.24
 782 (CH_3), 22.77 (CH_2), 24.79 (CH_2), 25.79 (CH_2), 29.02 (CH_2), 29.11
 783 (CH_2), 29.15 (CH_2), 29.47 (CH_2), 29.50 (CH_2), 31.98 (CH_2), 32.73
 784 (CH_2), 34.06 (CH_2), 36.86 (CH_2), 40.81 (CHCH_2CHO), 71.17
 785 (CHOH), 126.05 (CHCH), 134.74 (CHCH), 179.27 (COOH). HR-

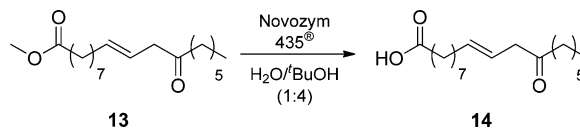
MS (ESI^+), m/z : $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{18}\text{H}_{34}\text{O}_3\text{Na}$ 321.240; found 321.2411.



N-(4'-Hydroxy-3'-methoxybenzyl)-(12R)-hydroxyoctadec-(9E)-enamide (12). General procedure I was applied to a solution of
 the acid **11** (70 mg, 0.23 mmol) dissolved in anhydrous DMF (3.3
 mL), amine hydrochloride salt **3** (53 mg, 0.28 mmol), DIPEA (122
 μL , 0.70 mmol), and HATU (133 mg, 0.35 mmol). The compound
12 was afforded (35 mg, 34%) as an off-white solid after the
 purification by silica gel flash column chromatography (petroleum
 ether/ EtOAc 6:4). $[\alpha]_{\text{D}}^{20}$ <+1° (c 0.5, DCM). R_f = 0.37 (petroleum
 ether/ EtOAc 6:4). Mp = 73–75 °C. IR (ATR) ν = 3295, 2920, 2849,
 1631, 1515, 1463, 1270, 1030, 959 cm^{-1} . ^1H NMR (400 MHz,
 CDCl_3) δ = 0.88 (t, 3H, J = 6.9 Hz, CH_3), 1.23–1.36 (m, 15H, CH_2 ,
 H_{13a}), 1.37–1.46 (m, 3H, CH_2 , H_{13b}), 1.59–1.71 (m, 2H, CH_2),
 1.96–2.09 (m, 3H, CH_2 , H_{11a}), 2.14–2.27 (m, 3H, CH_2 , H_{11b}), 3.53–
 3.61 (m, 1H, CHOH), 3.86 (s, 3H, CH_3O), 4.34 (d, J = 5.7 Hz, 2H,
 CH_2NH), 5.35–5.44 (m, 1H, CHCH), 5.47–5.56 (m, 1H, CHCH),
 5.72 (br s, 2H, CH_2NH , OH), 6.79 (ddd, 3H, J = 16.1, 9.9, 5.0 Hz,
 H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.23 (CH_3), 22.75 (CH_2),
 25.79 (CH_2), 25.86 (CH_2), 29.06 (CH_2), 29.26 (CH_2), 29.35 (CH_2),
 29.46 (CH_2), 29.49 (CH_2), 31.97 (CH_2), 32.73 (CH_2), 36.91 (CH_2),
 36.96 (CH_2), 40.82 (CHCH_2CHO), 43.65 (CH_2NH), 56.07
 (CH_3O), 71.07 (CHOH), 110.86 (C_{Ar}), 114.53 (C_{Ar}), 120.91
 (C_{Ar}), 126.12 (CHCH), 130.54 (C_{Ar}), 134.68 (CHCH), 145.26
 (C_{Ar}), 146.84 (C_{Ar}), 173.01 (NHCO). HR-MS (ESI^+), m/z : $[\text{M} +$
 $\text{H}]^+$ calcd for $\text{C}_{26}\text{H}_{44}\text{NO}_4$ 434.3265; found 434.3293.

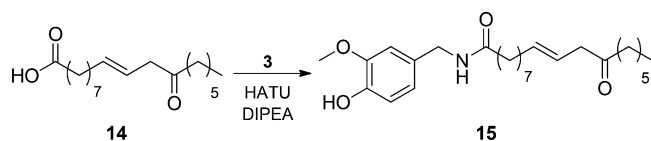


Methyl 12-Oxooctadec-(9E)-enoate (13). CrO_3 (960 mg, 9.6
 mmol) and pyridine (1.5 mL, 19.2 mmol) were added to a solution of
 compound **10** (500 mg, 1.6 mmol) in DCM (6 mL). The mixture was
 vigorously stirred at room temperature for 2 h. The reaction mixture
 was filtered over Celite and washed with 1 M HCl . The organic phase
 was dried over anhydrous Na_2SO_4 , filtered and the solvent was
 evaporated under reduced pressure to yield the ketone **13**⁶ (246 g,
 49%) as a yellowish oil after the purification by silica gel column
 chromatography (petroleum ether/ Et_2O 98:2). R_f = 0.48 (petroleum
 ether/ Et_2O 9:1). IR (ATR) ν = 2925, 2854, 1738, 1715, 1435, 1362,
 1195, 1170, 968, 725 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.87 (t,
 3H, J = 6.5 Hz, CH_3), 1.23–1.38 (m, 14H, CH_2), 1.51–1.64 (m, 4H,
 CH_2), 1.96–2.08 (m, 2H, CH_2), 2.29 (t, J = 6.9 Hz, 2H, CH_2), 2.41
 (t, 2H, J = 6.9 Hz, COCH_2), 3.07 (d, 2H, J = 5.2 Hz, CH_2CO), 3.66
 (s, 3H, CH_3O), 5.45–5.56 (m, 2H, $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz,
 CDCl_3) δ = 14.16 (CH_3), 22.63 (CH_2), 23.84 (CH_2), 25.06 (CH_2),
 29.03 (CH_2), 29.06 (CH_2), 29.21 (2 × CH_2), 29.27 (CH_2), 31.73
 (CH_2), 32.67 (CH_2), 34.22 (CH_2), 42.31 (COCH_2), 46.95 (CH_2CO),
 51.57 (CH_3O), 122.13 (CHCH), 135.16 (CHCH), 174.42 (COO^-),
 209.95 (COCH_2).

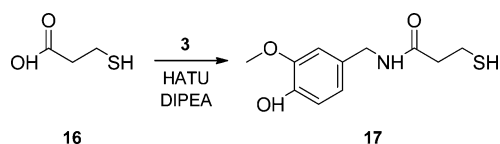


12-Oxooctadec-(9E)-enoic Acid (14). Novozym 435 (20 mg, 832
 50% w/w) was added to a solution of the methyl ester **13** (20 mg,
 0.06 mmol) in a mixture of H_2O (31 μL) and *tert*- BuOH (138 μL).
 The reaction mixture was stirred at 45 °C for 24 h. The mixture was
 filtered and the solvent was evaporated under reduced pressure to 836

yield the acid **14** (17 mg, 89%) as a white solid. Mp = 71–73 °C. IR (ATR) ν = 3121, 2954, 2918, 2848, 1701, 1263, 1082, 962, 720, 689 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.26–1.36 (m, 14H, CH_2), 1.50–1.58 (m, 2H, CH_2), 1.58–1.66 (m, 2H, CH_2), 1.98–2.08 (m, 2H, CH_2), 2.34 (t, 2H, J = 6.9 Hz, CH_2), 2.41 (t, 2H, J = 6.9 Hz, COCH_2), 3.08 (d, 2H, J = 5.2 Hz, CH_2CO), 5.44–5.57 (m, 2H, CHCH). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.17 (CH_3), 22.63 (CH_2), 23.85 (CH_2), 24.79 (CH_2), 29.03 ($2 \times \text{CH}_2$), 29.12 (CH_2), 29.18 (CH_2), 29.26 (CH_2), 31.73 (CH_2), 32.66 (CH_2), 34.09 (CH_2), 42.32 (COCH_2), 46.95 (CH_2CO), 122.13 (CHCH), 135.17 (CHCH), 179.59 (COOH), 210.13 (COCH_2). HR-MS (ESI⁺), m/z : $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{18}\text{H}_{32}\text{O}_3\text{Na}$ 319.2244; found 319.2267.

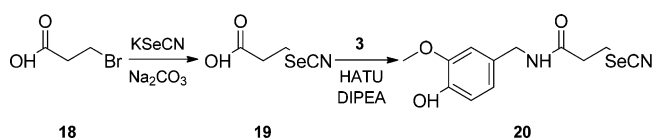


***N*-(4'-Hydroxy-3'-methoxybenzyl)-12-oxooctadec-(9*E*)-enamide (15).** General procedure I was applied to a solution of the acid **14** (210 mg, 0.71 mmol) dissolved in anhydrous DMF (10 mL), amine hydrochloride salt **3** (148 mg, 0.78 mmol), DIPEA (400 μL , 2.1 mmol), and HATU (404 mg, 1.06 mmol). The compound **15** was obtained (52 mg, 17%) as an off-white solid after the purification by silica gel flash column chromatography (petroleum ether/EtOAc 7:3). Mp = 71–73 °C. R_f = 0.36 (petroleum ether/EtOAc 7:3). IR (ATR) ν = 3393, 3312, 2917, 2850, 1703, 1636, 1554, 1509, 1242, 1125, 967, 705 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.22–1.38 (m, 14H, CH_2), 1.50–1.58 (m, 2H, CH_2), 1.59–1.69 (m, 2H, CH_2), 1.97–2.04 (m, 2H, CH_2), 2.19 (t, 2H, J = 7.4 Hz, CH_2), 2.40 (t, 2H, J = 7.4 Hz, COCH_2), 3.08 (d, 2H, J = 5.2 Hz, CH_2CO), 3.87 (s, 3H, CH_3O), 4.35 (d, 2H, J = 5.7 Hz, CH_2NH), 5.47–5.52 (m, 2H, CHCH), 5.67 (s, 1H, CH_2NH), 5.73 (br s, 1H, OH), 6.73–6.87 (6.79 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar})). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.17 (CH_3), 22.63 (CH_2), 23.86 (CH_2), 25.86 (CH_2), 29.03 (CH_2), 29.05 (CH_2), 29.23 (CH_2), 29.26 (CH_2), 29.36 (CH_2), 31.73 (CH_2), 32.64 (CH_2), 36.96 (CH_2), 42.37 (COCH_2), 43.66 (CH_2NH), 46.89 (CH_2CO), 56.07 (CH_3O), 110.83 (C_{Ar}), 114.50 (C_{Ar}), 120.92 (C_{Ar}), 122.12 (CHCH), 130.56 (C_{Ar}), 135.11 (CHCH), 145.25 (C_{Ar}), 146.82 (C_{Ar}), 172.99 (NHCO), 210.08 (COCH_2). HR-MS (ESI⁺), m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{26}\text{H}_{42}\text{NO}_4$ 432.3108; found 432.3137.



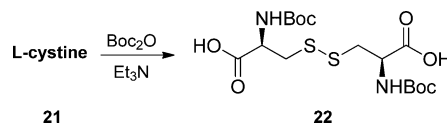
***N*-(4'-Hydroxy-3'-methoxybenzyl)-3-mercaptopropanamide (17).** General procedure I was applied to a solution of mercaptopropionic acid (1.2 mL, 12.68 mmol) dissolved in anhydrous DMF (30 mL), amine hydrochloride salt **3** (2.65 g, 13.95 mmol), DIPEA (6.63 mL, 38.04 mmol), and HATU (7.23 g, 19.02 mmol). Compound **17** was obtained after silica gel column chromatography (petroleum ether/EtOAc 5:5) as sticky oil (2.14 g, 74%). R_f = 0.60 (petroleum ether/EtOAc 4:6). IR (ATR) ν = 3425, 2922, 2853, 1515, 836 cm^{-1} . ^1H NMR (400 MHz, $(\text{CH}_3)_2\text{CO}$) δ = 1.86 (t, 1H, J = 8.2 Hz, SH), 2.54 (t, 2H, J = 6.7 Hz, CH_2), 2.70–2.82 (m, 2H, CH_2SH), 3.80 (s, 3H, CH_3O), 4.31 (d, 2H, J = 5.9 Hz, CH_2NH), 6.74 (d, 2H, J = 1.0 Hz, H_{Ar} , OH), 6.92 (s, 1H, H_{Ar}), 7.48 (s, 2H, H_{Ar} , CH_2NH). ^{13}C NMR (101 MHz, $(\text{CH}_3)_2\text{CO}$) δ = 20.10 (CH_2SH), 39.71 (CH_2), 42.47 (CH_2NH), 55.33 (CH_3O), 111.25 (C_{Ar}), 114.66 (C_{Ar}), 120.16 (C_{Ar}), 130.83 (C_{Ar}), 145.61 (C_{Ar}), 147.36 (C_{Ar}), 170.16 (NHCO). HR-MS (ESI⁺), m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{11}\text{H}_{15}\text{NO}_3\text{S}$ 242.0845; found 242.0861.

3-Selenocyanatopropanoic Acid (19). To a solution of 3-bromopropionic acid **18** (1.5 g, 9.8 mmol) in water (3 mL) was added Na_2CO_3 until pH 7. A volume of 14 mL of 10% KSeCN (1.41

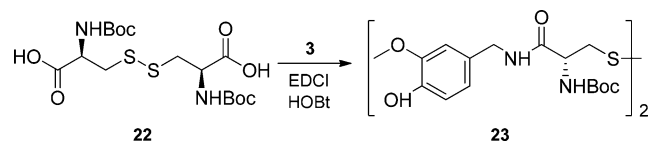


g, 9.8 mmol, 1 equiv) aqueous solution was added. The mixture stirred at room temperature for 2 days. After removing partially the solvent under reduced pressure, the crude was dissolved in Et_2O and washed with 1 M HCl, water and brine. The organic solution was dried over Na_2SO_4 , filtered and the solvent was removed under reduced pressure to yield the 3-selenocyanatopropanoic acid **19** as a yellow oil (1.39 g, 80%) which was used in the next step without further purification. IR (ATR) ν = 3024, 2649, 2152, 1703, 1401 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 3.07 (t, 2H, J = 6.4 Hz, CH_2SeCN), 3.24 (dd, 2H, J = 6.4 Hz, $\text{CH}_2\text{CH}_2\text{SeCN}$), 9.52 (br s, 1H, COOH). ^{13}C NMR (101 MHz, CDCl_3) δ = 22.89 (CH_2SeCN), 34.90 ($\text{CH}_2\text{CH}_2\text{SeCN}$), 101.68 (SeCN), 176.86 (COOH).

***N*-(4'-Hydroxy-3'-methoxybenzyl)-3-selenocyanatopropanamide (20).** General procedure I was applied to a solution of compound **19** (1.3 g, 7.30 mmol), amine hydrochloride salt **3** (1.52 g, 8.03 mmol), DIPEA (3.82 mL, 21.9 mmol), and HATU (4.16 g, 10.95 mmol) in anhydrous DMF (20 mL). Compound **20** was afforded after silica gel column chromatography (petroleum ether/EtOAc 5:5) as a white sticky solid (2.14 g, 60%). R_f = 0.65 (petroleum ether/EtOAc 4:6). IR (ATR) ν = 3315, 2924, 2853, 2148, 1638, 1235 cm^{-1} . ^1H NMR (400 MHz, $(\text{CH}_3)_2\text{CO}$) δ = 2.94 (t, 2H, J = 6.4 Hz, COCH_2), 3.34 (t, 2H, J = 6.4 Hz, CH_2SeCN), 3.81 (s, 3H, CH_3), 4.30 (d, 2H, J = 5.8 Hz, CH_2NH), 6.75 (s, 2H, H_{Ar}), 6.91 (s, 1H, H_{Ar}), 7.48 (s, 1H, OH), 7.72 (s, 1H, CH_2NH). ^{13}C NMR (101 MHz, $(\text{CH}_3)_2\text{CO}$) δ = 24.79 (CH_2SeCN), 34.84 ($\text{CH}_2\text{CH}_2\text{SeCN}$), 42.73 (CH_2NH), 55.33 (CH_3O), 104.64 (SeCN), 111.35 (C_{Ar}), 114.72 (C_{Ar}), 120.32 (C_{Ar}), 130.19 (C_{Ar}), 145.79 (C_{Ar}), 147.38 (C_{Ar}), 170.92 (NHCO). HR-MS (ESI⁺), m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{12}\text{H}_{15}\text{N}_2\text{O}_3\text{Se}$ 315.0248; found 315.0242.

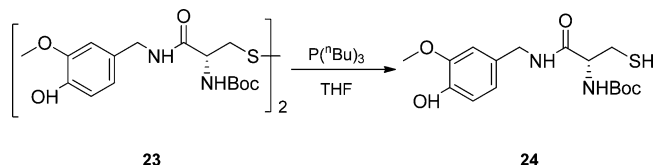


***N,N*-Di-Boc-L-cystine (22).** General procedure III was applied to L-cystine **21** (10 g, 41.67 mmol), Boc_2O (27.25 g, 124.85 mmol), and Et_3N (17.5 mL, 125.38 mmol) in water (150 mL) to yield compound **22** as a white solid, which was thoroughly washed with petroleum ether several times (17.56 g, 96%). Mp: 145–146 °C. IR (ATR) ν = 3366, 2985, 2936, 1682, 1511, 1163, 1052, 868 cm^{-1} . ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ = 1.37 (s, 18H, Boc), 2.87 (dd, 2H, J = 13.5, 10.1 Hz, CHCH_2), 3.09 (dd, 2H, J = 13.5, 4.4 Hz, CHCH_2), 4.16 (td, 2H, J = 10.1, 4.4 Hz, CHCH_2), 7.18 (d, 2H, J = 8.4 Hz, NH), 12.79 (s, 2H, COOH). ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{SO}$) δ = 28.60 ($\text{C}(\text{CH}_3)_3$), 52.96 (CHCH_2), 78.70 ($\text{C}(\text{CH}_3)_3$), 155.79 (NHCO_2), 172.82 (COOH).

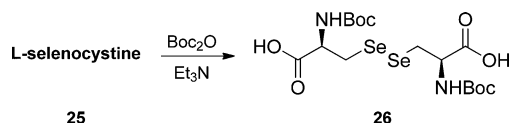


Di-[(2*R*)-*N*-Boc-amino-1-[(4'-hydroxy-3'-methoxybenzyl)-amino]-1-oxoprop-3-yl]disulfane (23). To a solution of compound **22** (5 g, 11.35 mmol) in anhydrous DMF (50 mL) were added HOBT (4.6 g, 34.05 mmol), Et_3N (4.74 mL, 34.05 mmol), and the amine hydrochloride salt **3** (5.16 g, 27.24 mmol). The mixture was stirred at 0 °C during 30 min. EDCI (6.52 g, 34 mmol) was added and the mixture stirred at room temperature during 20 h. To the mixture were added EtOAc and brine, and the aqueous phase was extracted with EtOAc. The combined organic solutions were washed with 1 M HCl, saturated NaHCO_3 , and brine. The organic solution was dried over anhydrous Na_2SO_4 , filtered and the solvent was

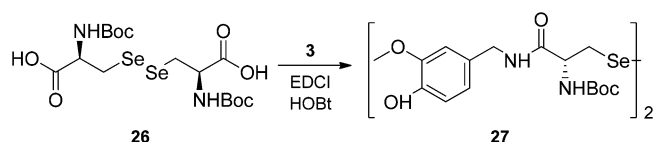
946 evaporated under reduced pressure. Compound **23** was afforded after
 947 silica gel column chromatography (PE/EtOAc 1:9) as a white solid
 948 (7.58 g, 94%). $R_f = 0.24$ (petroleum ether/EtOAc 1:9). Mp: 167–170
 949 °C. $[\alpha]_D^{20} -67.42$ (c 0.75, MeOH). IR (ATR) $\nu = 3330, 2975, 2935,$
 950 $1658, 1511, 1272, 1033\text{ cm}^{-1}$. $^1\text{H NMR}$ (400 MHz, $(\text{CD}_3)_2\text{SO}$) $\delta =$
 951 1.36 (s, 18H, $\text{C}(\text{CH}_3)_3$), 2.86 (dd, 2H, $J = 13.0, 9.9\text{ Hz}$, CHCH_2),
 952 3.07 (dd, 2H, $J = 13.0, 4.8\text{ Hz}$, CHCH_2), 3.72 (s, 6H, CH_3O), 4.02 –
 953 4.32 (m, 6H, CHCH_2 , CH_2NH), 6.55 – 6.72 (m, 4H, H_{Ar} , NHBoc),
 954 6.79 (s, 2H, H_{Ar}), 7.06 (d, 2H, $J = 8.4\text{ Hz}$, H_{Ar}), 8.31 (t, 2H, $J = 5.4$
 955 Hz , CH_2NH), 8.78 (br s, 2H, OH). $^{13}\text{C NMR}$ (101 MHz, $(\text{CD}_3)_2\text{SO}$)
 956 $\delta = 28.59$ ($\text{C}(\text{CH}_3)_3$), 40.59 (CHCH_2), 42.40 (CH_2NH), 54.17
 957 (CHCH_2), 55.92 (CH_3O), 78.73 ($\text{C}(\text{CH}_3)_3$), 111.82 (C_{Ar}), 115.53
 958 (C_{Ar}), 119.88 (C_{Ar}), 130.37 (C_{Ar}), 145.76 (C_{Ar}), 147.85 (C_{Ar}), 155.70
 959 (NHCO_2), 170.60 (NHCO). HR-MS (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd
 960 for $\text{C}_{32}\text{H}_{47}\text{N}_4\text{O}_{10}\text{S}_2$, 711.2734; found 711.2793.



961 ***N*-(4'-Hydroxy-3'-methoxy)benzyl-(2*R*)-(Boc-amino)-3-**
 962 **mercaptopropanamide (24).** General procedure IV (SS bond
 963 cleavage) was applied to compound **23** (7 g, 9.86 mmol) dissolved in
 964 THF (60 mL), $\text{P}(\text{t-Bu})_3$ (2.55 mL, 10.35 mmol) in the presence of
 965 water (1.3 mL). Compound **24** was afforded after silica gel column
 966 chromatography (petroleum ether/EtOAc 5:5) as a white solid (5.11
 967 g, 73%). $R_f = 0.42$ (petroleum ether/EtOAc 4:6). Mp: 108–110 °C.
 968 $[\alpha]_D^{20} -15.65$ (c 1.6, MeOH). IR (ATR) $\nu = 3456, 3327, 2989, 2934,$
 969 $2847, 1678, 1513, 1240\text{ cm}^{-1}$. $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 1.41$
 970 (s, 9H, $\text{C}(\text{CH}_3)_3$), 1.54 (t, 1H, $J = 10.7\text{ Hz}$, SH), 2.74 (ddd, 1H, $J =$
 971 $13.8, 10.2, 6.1\text{ Hz}$, CHCH_2), 3.09 (ddd, 1H, $J = 13.6, 7.6, 4.6\text{ Hz}$,
 972 CHCH_2), 3.84 (s, 3H, CH_3O), 4.25 – 4.44 (m, 3H, CHCH_2 ,
 973 CH_2NH), 5.48 (d, 1H, $J = 7.8\text{ Hz}$, CH_2NH), 5.81 (br s, 1H, OH),
 974 6.67 – 6.89 (m, 4H, H_{Ar} , NHBoc). $^{13}\text{C NMR}$ (101 MHz, CDCl_3) $\delta =$
 975 26.96 (CHCH_2), 28.23 ($\text{C}(\text{CH}_3)_3$), 43.47 (CH_2NH), 55.67
 976 (CHCH_2), 55.93 (CH_3O), 80.69 ($\text{C}(\text{CH}_3)_3$), 110.47 (C_{Ar}), 114.44
 977 (C_{Ar}), 120.58 (C_{Ar}), 129.66 (C_{Ar}), 145.12 (C_{Ar}), 146.74 (C_{Ar}), 155.46
 978 (NHCO_2), 169.88 (NHCO). HR-MS (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd
 979 for $\text{C}_{16}\text{H}_{24}\text{N}_2\text{O}_5\text{SNa}$, 379.1298; found 379.1326.

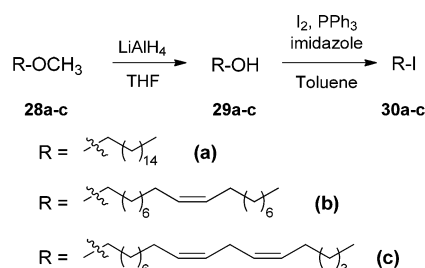


980 ***N,N*-Di-Boc-L-selenocystine (26).** General procedure III was
 981 applied to L-selenocystine **25** (1.5 g, 4.49 mmol), Boc_2O (3.24 g,
 982 13.48 mmol), and Et_3N (1.88 mL, 13.48 mmol) in water (22 mL) to
 983 yield compound **26** as a yellow solid (1.55 g, 65%), which was used
 984 in the next step without further purification. Mp: 145–147 °C. $[\alpha]_D^{20}$
 985 -75.63 (c 1.5, DCM). IR (ATR) $\nu = 3364, 2979, 2557, 1698, 1662,$
 986 1506 cm^{-1} . $^1\text{H NMR}$ (400 MHz, $(\text{CD}_3)_2\text{SO}$) $\delta = 1.37$ (s, 18H,
 987 $\text{C}(\text{CH}_3)_3$), 3.10 (dd, 2H, $J = 11.9, 10.2\text{ Hz}$, CHCH_2), 3.28 (dd, 2H, J
 988 $= 11.9, 4.7\text{ Hz}$, CHCH_2), 4.06 – 4.21 (m, 2H, CHCH_2), 7.17 (d, 2H, J
 989 $= 8.3\text{ Hz}$, NH), 12.79 (s, 2H, COOH). $^{13}\text{C NMR}$ (101 MHz,
 990 $(\text{CD}_3)_2\text{SO}$) $\delta = 28.61$ ($\text{C}(\text{CH}_3)_3$), 31.38 (CHCH_2), 54.68 (CHCH_2),
 991 78.71 ($\text{C}(\text{CH}_3)_3$), 155.71 (NHCO_2), 172.91 (COOH).



992 **Di-[(2*R*)-*N*-Boc-amino-1-((4'-hydroxy-3'-methoxybenzyl)-**
 993 **amino)-1-oxoprop-3-yl]diseleno (27).** To a solution of compound
 994 **26** (1.5 g, 2.80 mmol) in anhydrous DMF (14 mL) were added HOBT
 995 (1.14 g, 8.4 mmol), Et_3N (1.18 mL, 8.4 mmol), and the amine

hydrochloride salt **3** (1.27 g, 6.72 mmol). The mixture was stirred at 0
 °C during 30 min. EDCI (1.61 g, 8.4 mmol) was added and the
 mixture stirred at room temperature during 20 h. To the mixture were
 added EtOAc and brine, and the aqueous phase was extracted. The
 combined organic layers were washed with 1 M HCl, saturated
 NaHCO_3 , and brine. The organic phase was dried over anhydrous
 Na_2SO_4 , filtered, and the solvent was evaporated under reduced
 pressure. Compound **27** was afforded after silica gel column
 chromatography (petroleum ether/EtOAc 1:9) as a white solid
 (1.98 g, 88%). $R_f = 0.26$ (petroleum ether/EtOAc 5:5). Mp: 93–95
 °C. $[\alpha]_D^{20} 42.94$ (c 0.7, DCM). IR (ATR) $\nu = 3314, 2975, 2932, 1654,$
 $1513, 1157\text{ cm}^{-1}$. $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 1.26$ (s, 18H,
 $\text{C}(\text{CH}_3)_3$), 3.12 – 3.30 (m, 4H, CHCH_2), 3.83 (s, 6H, CH_3O), 4.25
 (dd, 2H, $J = 14.7, 5.4\text{ Hz}$, CH_2NH), 4.48 (dd, 2H, $J = 14.7, 6.5\text{ Hz}$,
 CH_2NH), 4.75 – 4.94 (m, 2H, CHCH_2), 5.58 (d, 2H, $J = 9.7\text{ Hz}$,
 NHBoc), 5.63 (s, 2H, OH), 6.77 (ddd, 6H, $J = 12.5, 9.9, 5.0$, H_{Ar}),
 8.06 (t, 2H, $J = 5.6\text{ Hz}$, CH_2NH). $^{13}\text{C NMR}$ (101 MHz, CDCl_3) $\delta =$
 28.15 ($\text{C}(\text{CH}_3)_3$), 37.43 (CHCH_2), 43.28 (CH_2NH), 55.24
 (CHCH_2), 55.86 (CH_3O), 78.98 ($\text{C}(\text{CH}_3)_3$), 110.44 (C_{Ar}), 114.24
 (C_{Ar}), 120.77 (C_{Ar}), 130.03 (C_{Ar}), 145.00 (C_{Ar}), 146.58 (C_{Ar}), 155.65
 (NHCO_2), 170.53 (NHCO). HR-MS (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd
 for $\text{C}_{32}\text{H}_{46}\text{N}_4\text{O}_{10}\text{Se}_2$, 807.1623; found 807.1621.



1-Hexadecanol (29a). General procedure V was applied to
 methyl palmitate **28a** (1 g, 3.69 mmol), LiAlH_4 (280 mg, 7.38 mmol)
 in anhydrous THF (20 mL). Compound **29a**¹⁰ was afforded after
 silica gel column chromatography (petroleum ether/ Et_2O 9:1) as a
 white solid (875 mg, 98%). $R_f = 0.88$ (petroleum ether/ Et_2O 9:1).
 Mp: 50–52 °C. IR (ATR) $\nu = 3320, 3226, 2915, 2919, 2847, 1462$
 cm^{-1} . $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 0.87$ (t, 3H, $J = 6.9\text{ Hz}$, CH_3),
 1.15 – 1.41 (m, 24H, CH_2), 1.45 – 1.64 (m, 4H, CH_2 , HOCH_2CH_2),
 3.62 (t, 2H, $J = 6.9\text{ Hz}$, HOCH_2CH_2). $^{13}\text{C NMR}$ (101 MHz, CDCl_3)
 $\delta = 14.08$ (CH_3), 22.67 (CH_2), 25.74 (CH_2), 29.35 (CH_2), 29.43
 (CH_2), 29.60 (CH_2), 29.61 (CH_2), 29.65 ($2 \times \text{CH}_2$), 29.67 (CH_2),
 29.68 ($3 \times \text{CH}_2$), 31.91 (CH_2), 32.78 (HOCH_2CH_2), 62.99
 (HOCH_2CH_2).

(9*Z*)-Octadecen-1-ol (29b). General procedure V was applied to
 methyl oleate **28b** (2.5 g, 8.43 mmol), LiAlH_4 (640 mg, 16.86 mmol)
 in anhydrous THF (50 mL). Compound **29b**¹¹ was afforded after
 silica gel column chromatography (petroleum ether/ Et_2O 9:1) as a
 brown oil (2.19 g, 97%). $R_f = 0.88$ (petroleum ether/ Et_2O 9:1). IR
 (ATR) $\nu = 3320, 2921, 2852, 1463, 1055\text{ cm}^{-1}$. $^1\text{H NMR}$ (400 MHz,
 CDCl_3) $\delta = 0.87$ (t, 3H, $J = 6.9\text{ Hz}$, CH_3), 1.16 – 1.41 (m, 22H, CH_2),
 1.47 – 1.62 (m, 2H, HOCH_2CH_2), 1.73 (s, 1H, OH), 2.00 (q, 4H, $J =$
 6.4 Hz , CH_2CH , CHCH_2), 3.61 (t, 2H, $J = 6.9\text{ Hz}$, HOCH_2CH_2),
 5.25 – 5.47 (m, 2H, $\text{CH}=\text{CH}$). $^{13}\text{C NMR}$ (101 MHz, CDCl_3) $\delta =$
 14.07 (CH_3), 22.65 (CH_2), 25.73 (CH_2), 27.16 (CH_2CH), 27.18
 (CHCH_2), 29.22 (CH_2), 29.30 ($2 \times \text{CH}_2$), 29.40 (CH_2), 29.49
 (CH_2), 29.50 (CH_2), 29.72 (CH_2), 29.74 (CH_2), 31.88 (CH_2), 32.75
 (HOCH_2CH_2), 62.93 (HOCH_2CH_2), 129.76 ($\text{CH}=\text{CH}$), 129.90
 ($\text{CH}=\text{CH}$).

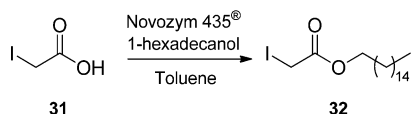
(9*Z*,12*Z*)-Octadecadien-1-ol (29c). General procedure V was
 applied to methyl linoleate **28b** (1 g, 3.39 mmol), LiAlH_4 (257 mg,
 6.79 mmol) in anhydrous THF (30 mL). Compound **29c**¹² was
 afforded after silica gel column chromatography (petroleum ether/
 Et_2O 9:1) as a colorless oil (885 mg, 98%). $R_f = 0.88$ (petroleum
 ether/ Et_2O 9:1). IR (ATR) $\nu = 3373, 2926, 2855, 1719, 1463\text{ cm}^{-1}$.
 $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 0.89$ (t, 3H, $J = 6.9\text{ Hz}$, CH_3),
 1.19 – 1.48 (m, 16H, CH_2), 1.51 – 1.61 (m, 2H, HOCH_2CH_2), 2.05

1054 (q, 4H, $J = 6.4$ Hz, $\text{CH}_2\text{CH}(\text{CHCH}_2)$), 2.77 (t, 2H, $J = 6.9$ Hz, CHCH_2CH), 3.59–3.67 (m, 2H, HOCH_2CH_2), 5.14–5.52 (m, 4H, 2 \times $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 14.04$ (CH_3), 22.55 (CH_2), 25.61 (CHCH_2CH), 25.71 (CH_2), 27.18 (CH_2CH), 27.20 (CHCH_2), 29.22 (CH_2), 29.33 (CH_2), 29.38 (CH_2), 29.48 (CH_2), 29.63 (CH_2), 31.51 (CH_2), 32.78 (HOCH_2CH_2), 63.03 (HOCH_2CH_2), 127.89 ($\text{CH}=\text{CH}$), 127.97 ($\text{CH}=\text{CH}$), 130.08 ($\text{CH}=\text{CH}$), 130.08 ($\text{CH}=\text{CH}$).

1062 **1-Iodohexadecane (30a)**. General procedure VI was applied to compound **29a** (1 g, 4.12 mmol), iodine (1.25 g, 4.95 mmol), PPh_3 (1.3 g, 4.95 mmol), and imidazole (0.85 g, 12.36 mmol) in toluene (15 mL). Compound **30a**¹³ was afforded after silica gel column chromatography (petroleum ether) as a yellow oil (1.08 g, 75%). $R_f = 0.1$ (petroleum ether). IR (ATR) $\nu = 2920, 2851, 1464, 1376, 1171, 719$ cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.88$ (t, 3H, $J = 6.9$ Hz, CH_3), 1.26 (s, 24H, CH_2), 1.34–1.41 (m, 2H, $\text{ICH}_2\text{CH}_2\text{CH}_2$), 1.75–1.87 (m, 2H, ICH_2CH_2), 3.18 (t, 2H, $J = 6.9$ Hz, ICH_2). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 7.21$ (ICH_2), 14.11 (CH_3), 22.69 (CH_2), 28.55 (CH_2), 29.36 (CH_2), 29.42 (CH_2), 29.55 (CH_2), 29.61 (CH_2), 29.73 ($2 \times \text{CH}_2$), 29.68 ($2 \times \text{CH}_2$), 29.69 (CH_2), 30.51 (CH_2), 31.92 (CH_2), 33.58 (ICH_2CH_2).

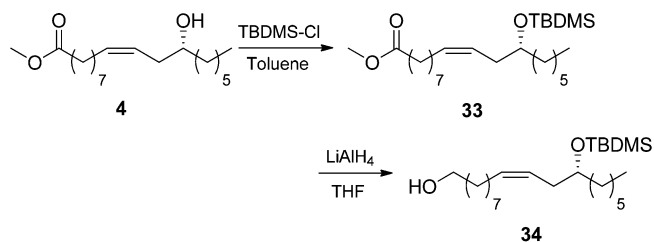
1075 **1-Iodo-(9Z)-octadecene (30b)**. General procedure VI was applied to compound **29b** (2 g, 7.45 mmol), iodine (2.27 g, 8.94 mmol), PPh_3 (2.34 g, 8.94 mmol), and imidazole (1.52 g, 22.35 mmol) in toluene (30 mL). Compound **30b**¹⁴ was afforded after silica gel column chromatography (petroleum ether/ Et_2O 9:1) as a yellow oil (2.42 g, 86%). $R_f = 0.1$ (petroleum ether/ Et_2O 9:1). IR (ATR) $\nu = 2921, 2852, 1462, 1181$ cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.88$ (t, 3H, $J = 6.9$ Hz, CH_3), 1.16–1.48 (m, 22H, CH_2), 1.72–1.91 (m, 2H, ICH_2CH_2), 2.01 (q, 4H, $J = 6.4$ Hz, $\text{CH}_2\text{CH}(\text{CHCH}_2)$), 3.18 (t, 2H, $J = 6.9$ Hz, ICH_2), 5.21–5.48 (m, 2H, $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 7.24$ (ICH_2), 14.10 (CH_3), 22.67 (CH_2), 27.15 (CH_2CH), 27.21 (CHCH_2), 28.50 (CH_2), 29.16 (CH_2), 29.29 (CH_2), 29.31 (CH_2), 29.51 (CH_2), 29.68 (CH_2), 29.75 (CH_2), 30.48 (CH_2), 31.89 (CH_2), 33.55 (ICH_2CH_2), 129.73 ($\text{CH}=\text{CH}$), 129.98 ($\text{CH}=\text{CH}$).

1090 **18-Iodo-(6Z,9Z)-octadecadiene (30c)**. General procedure VI was applied to compound **29c** (850 mg, 3.18 mmol), iodine (968 mg, 3.81 mmol), PPh_3 (1 g, 3.81 mmol), and imidazole (650 mg, 9.54 mmol) in toluene (15 mL). Compound **30c**¹⁴ was afforded after silica gel column chromatography (petroleum ether) as a yellow oil (1.13 g, 95%). $R_f = 0.1$ (petroleum ether/ Et_2O 9:1). IR (ATR) $\nu = 3439, 2926, 2855, 1707, 1458, 1175$ cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.89$ (t, 3H, $J = 6.9$ Hz, CH_3), 1.18–1.50 (m, 16H, CH_2), 1.78–1.86 (m, 2H, ICH_2CH_2), 2.05 (q, 4H, $J = 6.4$ Hz, $\text{CH}_2\text{CH}(\text{CHCH}_2)$), 2.77 (t, 2H, $J = 6.9$ Hz, CHCH_2CH), 3.18 (t, 2H, $J = 6.9$ Hz, ICH_2CH_2), 5.25–5.50 (m, 2 \times $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 7.20$ (ICH_2), 14.07 (CH_3), 22.57 (CH_2), 25.63 (CHCH_2CH), 27.18 (CH_2CH), 27.20 (CHCH_2), 28.50 (CH_2), 29.17 (CH_2), 29.30 (CH_2), 29.34 (CH_2), 29.59 (CH_2), 30.48 (CH_2), 31.52 (CH_2), 33.55 (ICH_2CH_2), 127.89 ($\text{CH}=\text{CH}$), 128.02 ($\text{CH}=\text{CH}$), 130.02 ($\text{CH}=\text{CH}$), 130.18 ($\text{CH}=\text{CH}$).



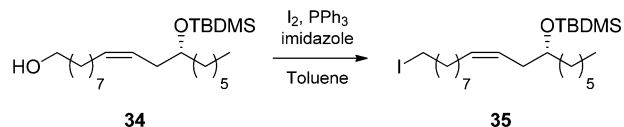
1106 **Hexadecyl 2-Iodoacetate (32)**. To a solution of iodoacetic acid **31** (500 mg, 2.69 mmol) in toluene (5 mL) were added 1-hexadecanol (978 mg, 4.03 mmol, 1.5 equiv) and Novozym 435 (150 mg). The reaction mixture was stirred at 50 $^{\circ}\text{C}$ for 2 days. The mixture was filtered off, EtOAc was added, and the organic phase was washed with saturated solution of NaHCO_3 , water, and brine. The organic solution was then dried over Na_2SO_4 , and the solvent was removed under reduced pressure. Compound **32**¹⁵ was afforded after silica gel column chromatography (petroleum ether/ Et_2O 9:1) as a yellow oil (562 mg, 51%). $R_f = 0.36$ (petroleum ether/ Et_2O 9:1). IR (ATR) $\nu = 2920, 2851, 1733, 1259, 1089$ cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.86$ (t, 3H, $J = 6.9$ Hz, CH_3), 1.14–1.41 (m, 26H, CH_2),

1.54–1.74 (m, 2H, $\text{COOCH}_2\text{CH}_2$), 3.68 (s, 2H, ICH_2), 4.13 (t, 2H, $J = 6.9$ Hz, $\text{COOCH}_2\text{CH}_2$). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 5.19$ (ICH_2), 14.27 (CH_3), 22.84 (CH_2), 25.90 (CH_2), 28.50 (CH_2), 29.33 (CH_2), 29.51 (CH_2), 29.63 (CH_2), 29.70 (CH_2), 29.78 (CH_2), 29.80 (CH_2), 29.82 (CH_2), 29.84 (3 \times CH_2), 32.07 (CH_2), 66.41 (COOCH_2), 169.00 (COOCH_2).



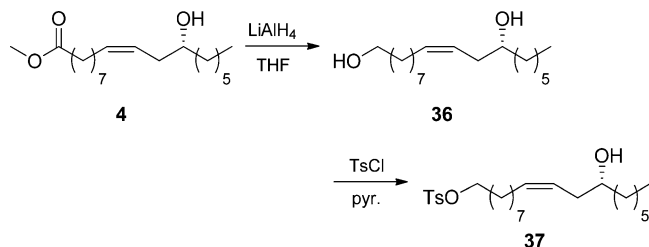
Methyl (12R)-[(tert-Butyldimethylsilyl)oxy]octadec-(9Z)-enoate (33). To a solution of methyl ricinoleate **4** (2 g, 6.4 mmol) in DCM (40 mL) were added DMAP (31 mg, 0.25 mmol) and Et_3N (2.23 mL, 16 mmol). TBDMS-Cl was slowly added (1.5 g, 9.92 mmol). The mixture was stirred at room temperature for 2 days. Then, the organic phase was washed with 1 M HCl, water, and brine, dried over anhydrous NaSO_4 and the solvent was removed under reduced pressure. Compound **33**¹⁶ was afforded after silica gel column chromatography (petroleum ether/ Et_2O 9:1) as a colorless oil (2.37 g, 87%). $R_f = 0.1$ (petroleum ether/ Et_2O 9:1). $[\alpha]_D^{20}$ 9.98 (c 2.8, DCM). IR (ATR) $\nu = 2927, 2855, 1742, 1461, 1251$ cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.04$ (s, 6H, $\text{Si}(\text{CH}_3)_2$), 0.78–0.95 (m, 12H, $\text{Si}(\text{CH}_3)_3$, CH_3), 1.16–1.46 (m, 18H, CH_2), 1.51–1.68 (m, 2H, COCH_2CH_2), 2.01 (q, 2H, $J = 6.4$ Hz, CH_2CH), 2.17 (t, 2H, $J = 6.9$ Hz, CHCH_2), 2.29 (t, 2H, $J = 6.9$ Hz, COCH_2CH_2), 3.59–3.73 (m, 4H, CH_3O , CH_2CHO), 5.29–5.51 (m, 2H, $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz, CDCl_3) $\delta = -4.59$ (SiCH_3), -4.38 (SiCH_3), 14.06 (CH_3), 18.11 ($\text{Si}(\text{CH}_3)_3$), 22.61 (CH_2), 24.92 (COCH_2CH_2), 25.38 (CH_2), 25.89 ($\text{Si}(\text{CH}_3)_3$), 27.40 (CH_2CH), 29.10 (CH_2), 29.12 (CH_2), 29.14 (CH_2), 29.45 (CH_2), 29.58 (CH_2), 31.87 (CH_2), 34.06 (COCH_2CH_2), 35.23 (CHCH_2), 36.84 (CH_2), 51.38 (CH_3O), 72.37 (CH_2CHO), 125.95 ($\text{CH}=\text{CH}$), 131.28 ($\text{CH}=\text{CH}$), 174.23 (COOH).

(12R)-[(tert-Butyldimethylsilyl)oxy]octadec-(9Z)-en-1-ol (34). General procedure V was applied to compound **33** (2.20 g, 5.15 mmol) with anhydrous LiAlH_4 (390 mg, 10.30 mmol) in dry THF (50 mL). Compound **34**¹⁷ was afforded after silica gel column chromatography (petroleum ether/ Et_2O 9:1) as a brown oil (1.91 g, 93%). $R_f = 0.86$ (petroleum ether/ Et_2O 9:1). $[\alpha]_D^{20}$ 13.21 (c 2.6, DCM). IR (ATR) $\nu = 3330, 2926, 2854, 1461, 1253, 1054$ cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.04$ (s, 6H, $\text{Si}(\text{CH}_3)_2$), 0.78–0.93 (m, 12H, $\text{Si}(\text{CH}_3)_3$, CH_3), 1.14–1.50 (m, 20H, CH_2), 1.51–1.62 (m, 2H, HOCH_2CH_2), 2.04 (q, 2H, $J = 6.4$ Hz, CH_2CH), 2.18 (t, 2H, $J = 6.9$ Hz, CHCH_2), 3.54–3.74 (m, 3H, HOCH_2CH_2 , CH_2CHO), 5.30–5.50 (m, 2H, $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz, CDCl_3) $\delta = -4.58$ (SiCH_3), -4.37 (SiCH_3), 14.07 (CH_3), 18.12 ($\text{Si}(\text{CH}_3)_3$), 22.61 (CH_2), 25.39 (CH_2), 25.72 (CH_2), 25.90 ($\text{Si}(\text{CH}_3)_3$), 27.43 (CH_2CH), 29.26 (CH_2), 29.38 (CH_2), 29.46 (CH_2), 29.49 (CH_2), 29.64 (CH_2), 31.87 (CH_2), 32.77 (HOCH_2CH_2), 35.24 (CHCH_2), 36.84 (CH_2), 63.00 (HOCH_2CH_2), 72.40 (CH_2CHO), 125.91 ($\text{CH}=\text{CH}$), 131.36 ($\text{CH}=\text{CH}$).



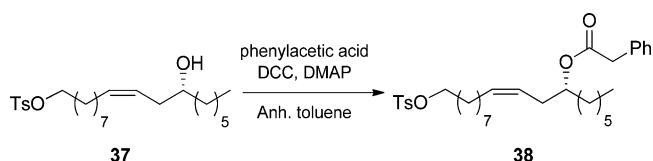
(12R)-[(tert-Butyldimethylsilyl)oxy]-1-iodooctadec-(9Z)-ene (35). General procedure VI was applied to compound **34** (1.8 g, 4.51 mmol), iodine (1.37 g, 5.42 mmol), PPh_3 (1.42 g, 5.42 mmol), and imidazole (921 mg, 13.53 mmol) in toluene (20 mL). Compound **35** was afforded after silica gel column chromatography (petroleum ether) as a colorless oil (1.86 g, 81%). $R_f = 0.1$ (petroleum ether/ Et_2O 9:1). $[\alpha]_D^{20}$ 7.12 (c 0.6, DCM). IR (ATR) $\nu = 2925, 2854, 1461, 1252$,

1172 1063 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.05 (s, 6H, $\text{Si}(\text{CH}_3)_2$),
 1173 0.80–0.97 (m, 12H, $\text{Si}(\text{CH}_3)_3$, CH_3), 1.15–1.49 (m, 20H, CH_2),
 1174 1.71–1.92 (m, 2H, ICH_2CH_2), 2.02 (q, 2H, J = 6.4 Hz, CH_2CH),
 1175 2.18 (t, 2H, J = 6.9 Hz, CHCH_2), 3.18 (t, 2H, J = 7.1 Hz, ICH_2CH_2),
 1176 3.57–3.75 (m, 1H, CH_2CHO), 5.29–5.52 (m, 2H, $\text{CH}=\text{CH}$). ^{13}C
 1177 NMR (101 MHz, CDCl_3) δ = –4.56 ($\text{Si}(\text{CH}_3)_3$), –4.35 ($\text{Si}(\text{CH}_3)_3$), 7.19
 1178 (ICH_2), 14.09 (CH_3), 18.13 ($\text{Si}(\text{CH}_3)_3$), 22.63 (CH_2), 25.40 (CH_2),
 1179 25.91 ($\text{Si}(\text{CH}_3)_3$), 27.42 (CH_2CH), 28.50 (CH_2), 29.21 (CH_2),
 1180 29.31 (CH_2), 29.47 (CH_2), 29.61 (CH_2), 30.48 (CH_2), 31.89 (CH_2),
 1181 33.55 (ICH_2CH_2), 35.25 (CHCH_2), 36.86 (CH_2), 72.38 (CH_2CHO),
 1182 125.97 ($\text{CH}=\text{CH}$), 131.30 ($\text{CH}=\text{CH}$).

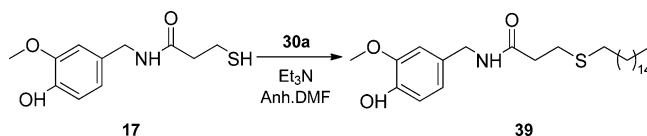


1183 **Octadec-9Z-ene-1-(12R)-diol (36).** General procedure V was
 1184 applied to methyl ricinoleate 4 (2.50 g, 8 mmol) with LiAlH_4 (607
 1185 mg, 16 mmol) in anhydrous THF (40 mL). Compound 36¹⁸ was
 1186 afforded after silica gel column chromatography (petroleum ether/
 1187 Et_2O 9:1) as a colorless oil (1.95 g, 86%). R_f = 0.82 (petroleum ether/
 1188 Et_2O 9:1). IR (ATR) ν = 3329, 2923, 2853, 1458, 1053 cm^{-1} . ^1H
 1189 NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.19–
 1190 1.39 (m, 18H, CH_2), 1.40–1.49 (m, 2H, CH_2), 1.51–1.58 (m, 2H,
 1191 HOCH_2CH_2), 1.59 (br s, 2H, OH), 2.04 (q, 2H, J = 6.4 Hz,
 1192 CH_2CH), 2.20 (t, 2H, J = 6.9 Hz, CHCH_2), 3.62 (m, 3H,
 1193 HOCH_2CH_2 , CH_2CHO), 5.29–5.47 (m, 1H, $\text{CH}=\text{CH}$), 5.47–
 1194 5.66 (m, 1H, $\text{CH}=\text{CH}$). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.06
 1195 (CH_3), 22.59 (CH_2), 25.68 (CH_2), 25.69 (CH_2), 27.36 (CH_2CH),
 1196 29.17 (CH_2), 29.31 (CH_2), 29.33 (CH_2), 29.40 (CH_2), 29.59 (CH_2),
 1197 31.81 (CH_2), 32.73 (HOCH_2CH_2), 35.32 (CHCH_2), 36.81 (CH_2),
 1198 62.96 (HOCH_2CH_2), 71.49 (CH_2CHO), 125.14 ($\text{CH}=\text{CH}$), 133.39
 1199 ($\text{CH}=\text{CH}$).

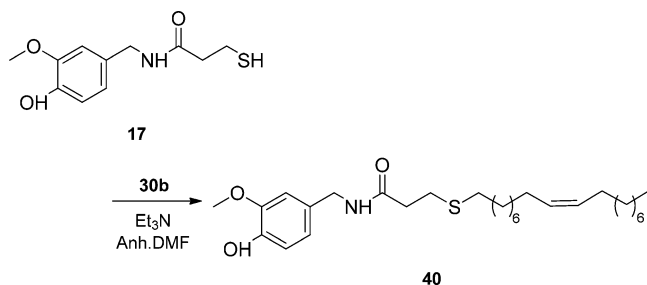
1200 **(12'R)-Hydroxyoctadec-9'Z-en-1-yl-4-methylbenzene-**
 1201 **sulfonate (37).** To a solution of compound 36 (1.6 g, 5.62 mmol) in
 1202 a mixture of DCM and pyridine (6 mL, 5:5) were added TsCl (1.07 g,
 1203 5.62 mmol, 1 equiv) in portions and DMAP (27 mg, 0.22 mmol). The
 1204 mixture was stirred at room temperature for 20 h. The mixture was
 1205 washed with 1 M HCl and extracted with EtOAc . The organic phase
 1206 was dried over Na_2SO_4 , and the solvent was removed under reduced
 1207 pressure. Compound 37¹⁹ was afforded after silica gel column
 1208 chromatography (petroleum ether/ Et_2O 7:3) as a yellow oil (1.11 g,
 1209 45%). R_f = 0.84 (petroleum ether/ Et_2O 7:3). $[\alpha]_D^{20}$ 4.40 (c 1.4,
 1210 DCM). IR (ATR) ν = 2924, 2854, 1458, 1358 cm^{-1} . ^1H NMR (400
 1211 MHz, CDCl_3) δ = 0.88 (t, 3H, J = 6.9 Hz, CH_3), 1.11–1.39 (m, 18H,
 1212 CH_2), 1.39–1.54 (m, 2H, CH_2), 1.53–1.70 (m, 2H, OCH_2CH_2) 2.03
 1213 (q, 2H, J = 6.4 Hz, CH_2CH), 2.20 (t, 2H, J = 6.9 Hz, CHCH_2), 2.44
 1214 (s, 3H, CH_3C), 3.54–3.71 (m, 1H, CH_2CHO), 4.01 (t, 2H, J = 6.9
 1215 Hz, OCH_2CH_2), 5.31–5.47 (m, 1H, $\text{CH}=\text{CH}$), 5.48–5.68 (m, 1H,
 1216 $\text{CH}=\text{CH}$), 7.33 (d, 2H, J = 8.5 Hz, H_{Ar}), 7.78 (d, 2H, J = 7.9 Hz,
 1217 H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.06 (CH_3), 21.60 (CH_3C),
 1218 22.59 (CH_2), 25.28 (CH_2), 25.69 (CH_2), 27.35 (CH_2CH), 28.78
 1219 (OCH_2CH_2), 28.84 (CH_2), 29.10 (CH_2), 29.22 (CH_2), 29.32 (CH_2),
 1220 29.56 (CH_2), 31.81 (CH_2), 35.34 (CHCH_2), 36.83 (C-CH_2), 70.64
 1221 (OCH_2CH_2), 71.45 (CH_2CHO), 125.23 ($\text{CH}=\text{CH}$), 127.84 (2 \times
 1222 C_{Ar}), 129.76 (2 \times C_{Ar}), 133.22 (C_{Ar}), 133.27 ($\text{CH}=\text{CH}$), 144.58
 1223 (C_{Ar}).



1224 **1'-Hexyl-12'-(Tosyloxy)dodec-(3'Z)-en-(1'R)-yl-2-phenyla-**
 1225 **acetate (38).** To a solution of compound 37 (900 mg, 2.05 mmol) in
 1226 anhydrous toluene (10 mL), phenylacetic acid (307 mg, 2.25 mmol),
 1227 1.1 equiv), DCC (1.02 g, 5.13 mmol, 2.5 equiv), and DMAP (500 mg,
 1228 4.1 mmol, 2 equiv) were added. The mixture was left stirred at room
 1229 temperature overnight and then filtered off to remove DCU. The
 1230 solvent was partially evaporated; the crude was dissolved in EtOAc
 1231 and washed with 1 M HCl, water, and brine. The organic phase was
 1232 dried over Na_2SO_4 and the solvent was removed under reduced
 1233 pressure. Compound 38 was afforded after silica gel column
 1234 chromatography (petroleum ether/ EtOAc 8:2) as a colorless oil
 1235 (935 mg, 82%). R_f = 0.53 (petroleum ether/ EtOAc 8:2). $[\alpha]_D^{20}$ 16.91
 1236 (c 5, DCM). IR (ATR) ν = 2925, 2855, 1730, 1361, 1187 cm^{-1} . ^1H
 1237 NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.11–
 1238 1.39 (m, 18H, CH_2), 1.42–1.56 (m, 2H, CH_2), 1.58–1.67 (m, 2H,
 1239 OCH_2CH_2), 1.97 (q, 2H, J = 6.4, CH_2CH), 2.13–2.38 (m, 2H,
 1240 CHCH_2), 2.44 (s, 3H, CH_3C), 3.58 (s, 2H, COCH_2), 4.01 (t, 2H, J =
 1241 6.9 Hz, OCH_2CH_2), 4.87 (p, 1H, J = 6.1 Hz, CH_2CHO), 5.19–5.37
 1242 (m, 1H, $\text{CH}=\text{CH}$), 5.37–5.55 (m, 1H, $\text{CH}=\text{CH}$), 7.19–7.43 (m,
 1243 7H, H_{Ar}), 7.79 (d, 2H, J = 8.0 Hz, H_{Ar}). ^{13}C NMR (101 MHz,
 1244 CDCl_3) δ = 14.04 (CH_3), 21.61 (CH_3C), 22.50 (CH_2), 25.17 (CH_2),
 1245 25.31 (CH_2), 27.27 (CH_2CH), 28.80 (OCH_2CH_2), 28.88 (CH_2),
 1246 29.04 (CH_2), 29.13 (CH_2), 29.27 (CH_2), 29.49 (CH_2), 31.66 (CH_2),
 1247 31.89 (CHCH_2), 33.53 (CH_2), 41.74 (COCH_2), 70.64 (OCH_2CH_2),
 1248 74.44 (CH_2CHO), 124.15 ($\text{CH}=\text{CH}$), 126.92 (C_{Ar}), 127.85 (2 \times
 1249 C_{Ar}), 128.44 (2 \times C_{Ar}), 129.20 (2 \times C_{Ar}), 129.76 (2 \times C_{Ar}), 132.57
 1250 ($\text{CH}=\text{CH}$), 133.25 (C_{Ar}), 134.31 (C_{Ar}), 144.57 (C_{Ar}), 171.27
 1251 (OCOCH_2). HR-MS (ESI^+), m/z : $[\text{M}+\text{NH}_4]^+$ calcd for $\text{C}_{33}\text{H}_{52}\text{NO}_3\text{S}$,
 1252 574.3561; found 573.3563.

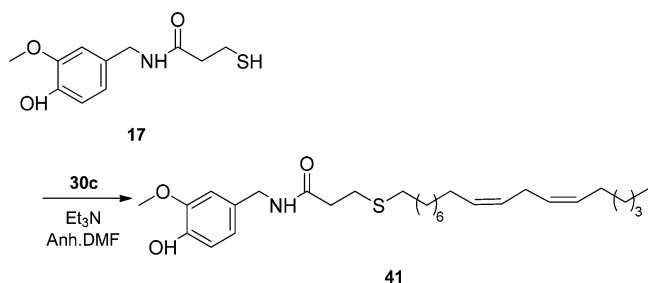


1253 **3-(Hexadecylthio)-N-(4'-hydroxy-3'-methoxybenzyl)-**
 1254 **propanamide (39).** General procedure VII was applied to 32 (150
 1255 mg, 0.62 mmol), compound 30a (245 mg, 0.70 mmol), and Et_3N
 1256 (175 μL , 1.24 mmol) dissolved in anhydrous DMF (4 mL).
 1257 Compound 39 was afforded after silica gel column chromatography
 1258 (petroleum ether/ EtOAc 7:3) as a white solid (136 mg, 42%). Mp =
 1259 72–73 $^\circ\text{C}$. R_f = 0.48 (petroleum ether/ EtOAc 5:5). IR (ATR) ν =
 1260 2925, 2855, 1730, 1361, 1187 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ =
 1261 0.88 (t, 3H, J = 6.9 Hz, CH_3), 1.23–1.32 (m, 24H, CH_2), 1.56–1.60
 1262 (m, 4H, SCH_2CH_2), 2.40–2.58 (m, 4H, COCH_2S , SCH_2CH_2), 2.84
 1263 (t, 2H, J = 6.9 Hz, CH_2S), 3.88 (s, 3H, CH_3O), 4.37 (d, 2H, J = 5.7
 1264 Hz, CH_2NH), 5.59 (s, 1H, CH_2NH), 5.90 (br s, 1H, OH), 6.81 (ddd,
 1265 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) δ =
 1266 14.28 (CH_3), 22.85 (CH_2), 28.04 (CH_2S), 29.05 (CH_2), 29.40 (CH_2),
 1267 29.52 (CH_2), 29.69 (CH_2), 29.77 (CH_2), 29.81 (3 \times CH_2), 29.85 (4
 1268 \times CH_2), 32.08 (CH_2), 32.63 (COCH_2), 37.07 (SCH_2CH_2), 43.80
 1269 (CH_2NH), 56.13 (CH_3O), 110.80 (C_{Ar}), 114.49 (C_{Ar}), 120.97 (C_{Ar}),
 1270 130.24 (C_{Ar}), 145.28 (C_{Ar}), 146.84 (C_{Ar}), 171.12 (NHCO). HR-MS
 1271 (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{48}\text{NO}_3\text{S}$, 466.3355; found
 1272 466.3378.

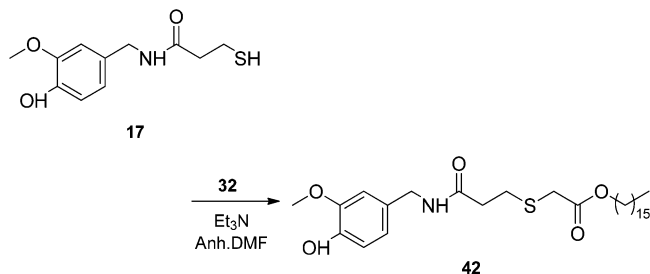


1273 **N-(4'-Hydroxy-3'-methoxybenzyl)-3-(octadec-9'Z-en-1-**
 1274 **ylthio)propanamide (40).** General procedure VII was applied to
 1275 compound 17 (100 mg, 0.41 mmol), compound 30b (174 mg, 0.46

1276 mmol), and Et₃N (115 μ L, 0.82 mmol) dissolved in anhydrous DMF
1277 (2 mL). Compound **40** was afforded after silica gel column
1278 chromatography (petroleum ether/EtOAc 5:5) as a white sticky
1279 solid (83 mg, 41%). R_f = 0.73 (petroleum ether/EtOAc 5:5). IR
1280 (ATR) ν = 3505, 3323, 2919, 2851, 1640, 1519 cm^{-1} . ¹H NMR (400
1281 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 6.9 Hz, CH₃), 1.23–1.37 (m, 22H,
1282 CH₂), 1.51–1.61 (m, 2H, SCH₂CH₂), 2.01 (q, 4H, J = 6.4 Hz,
1283 CH₂CH, CHCH₂), 2.44–2.55 (m, 4H, COCH₂, SCH₂CH₂), 2.83 (t,
1284 2H, J = 6.9 Hz, COCH₂CH₂), 3.88 (s, 3H, CH₃O), 4.37 (d, 2H, J =
1285 5.7 Hz, CH₂NH), 5.28–5.40 (m, 2H, CH=CH), 5.64 (s, 1H, OH),
1286 5.94 (br s, 1H, CH₂NH), 6.81 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}).
1287 ¹³C NMR (101 MHz, CDCl₃) δ = 14.26 (CH₃), 22.82 (CH₂), 27.33
1288 (CH₂CH), 27.36 (CHCH₂), 28.03 (CH₂S), 29.03 (CH₂), 29.35
1289 (CH₂), 29.39 (CH₂), 29.46 (2 \times CH₂), 29.57 (CH₂), 29.66 (CH₂),
1290 29.76 (CH₂), 29.88 (CH₂), 29.91 (CH₂), 32.04 (CH₂), 32.61
1291 (COCH₂), 37.08 (SCH₂CH₂), 43.77 (CH₂NH), 56.11 (CH₃O),
1292 110.80 (C_{Ar}), 114.49 (C_{Ar}), 120.93 (C_{Ar}), 129.93 (CH=CH), 130.11
1293 (CH=CH), 130.21 (C_{Ar}), 145.27 (C_{Ar}), 146.83 (C_{Ar}), 171.13
1294 (NHCO). HR-MS (ESI⁺), m/z : [M + H]⁺ calcd for C₂₉H₅₀NO₃S,
1295 492.3511; found 492.3502.

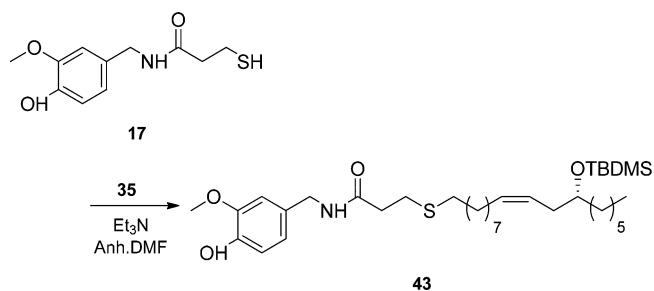


1296 **N-(4'-Hydroxy-3'-methoxybenzyl)-3-(octadeca-(9Z,12Z)-**
1297 **dien-1-ylthio)propanamide (41)**. General procedure VII was
1298 applied to compound **17** (100 mg, 0.41 mmol), compound **30c**
1299 (173 mg, 0.46 mmol), and Et₃N (115 μ L, 0.82 mmol) dissolved in
1300 anhydrous DMF (2 mL). Compound **41** was afforded after silica gel
1301 column chromatography (petroleum ether/EtOAc 7:3) as a yellow oil
1302 (110 mg, 55%). R_f = 0.66 (petroleum ether/EtOAc 5:5). IR (ATR) ν
1303 = 2923, 2854, 1643, 1515, 1273 cm^{-1} . ¹H NMR (400 MHz, CDCl₃) δ
1304 = 0.89 (t, 3H, J = 6.9 Hz, CH₃), 1.25–1.39 (m, 16H, CH₂), 1.51–
1305 1.62 (m, 2H, SCH₂CH₂), 2.04 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂),
1306 2.42–2.59 (m, 4H, SCH₂CH₂), 2.69–2.90 (m, 4H, COCH₂CH₂,
1307 CHCH₂CH), 3.87 (s, 3H, CH₃O), 4.36 (d, 2H, J = 5.7 Hz, CH₂NH),
1308 5.26–5.43 (m, 4H, 2 \times CH=CH), 5.66 (s, 1H, OH), 5.96 (s, 1H,
1309 CH₂NH), 6.80 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101
1310 MHz, CDCl₃) δ = 14.21 (CH₃), 22.71 (CH₂), 25.77 (CHCH₂CH),
1311 27.34 (CH₂CH), 27.35 (CHCH₂), 28.02 (CH₂S), 29.02 (CH₂), 29.34
1312 (CH₂), 29.38 (CH₂), 29.48 (CH₂), 29.56 (CH₂), 29.78 (CH₂), 29.66
1313 (CH₂), 31.59 (CH₂), 32.59 (COCH₂), 37.03 (SCH₂CH₂), 43.77
1314 (CH₂NH), 56.10 (CH₃O), 110.80 (C_{Ar}), 114.49 (C_{Ar}), 120.92 (C_{Ar}),
1315 128.04 (CH=CH), 128.14 (CH=CH), 130.19 (C_{Ar}), 130.22 (CH=
1316 CH), 130.34 (CH=CH), 145.27 (C_{Ar}), 146.83 (C_{Ar}), 171.14
1317 (NHCO). HR-MS (ESI⁺), m/z : [M + H]⁺ calcd for C₂₉H₄₈NO₃S,
1318 490.3355; found 490.3351.

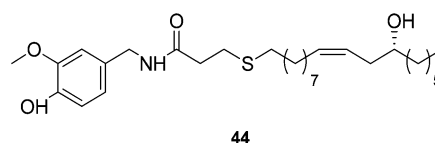


1319 **Hexadecyl 2-[(3'-(4'-Hydroxy-3'-methoxybenzyl)amino)-**
1320 **3'-oxopropyl]thioacetate (42)**. General procedure VII was
1321 applied to compound **17** (50 mg, 0.21 mmol), compound **32** (95

1322 mg, 0.23 mmol), and Et₃N (60 μ L, 0.42 mmol) dissolved in
1323 anhydrous DMF (2 mL). Compound **42** was afforded after silica gel
1324 column chromatography (petroleum ether/EtOAc 6:4) as a white
1325 solid (75 mg, 68%). Mp: 59–60 °C. R_f = 0.61 (petroleum ether/
1326 EtOAc 5:5). IR (ATR) ν = 3370, 3278, 2955, 2917, 2849, 1726, 1269
1327 cm^{-1} . ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, J = 6.9 Hz, 3H, CH₃),
1328 1.24–1.33 (m, 26H, CH₂), 1.57–1.65 (m, 2H, COOCH₂CH₂), 2.53
1329 (t, 2H, J = 6.9 Hz, COCH₂), 2.97 (t, 2H, J = 6.9 Hz, COCH₂CH₂),
1330 3.24 (s, 2H, SCH₂), 3.88 (s, 3H, CH₃OH), 4.06 (t, 2H, J = 6.9 Hz,
1331 COOCH₂CH₂), 4.37 (d, 2H, J = 5.7 Hz, CH₂NH), 5.63 (br s, 1H,
1332 OH), 6.09 (br s, 1H, CH₂NH), 6.80 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz,
1333 H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.26 (CH₃), 22.83 (CH₂),
1334 25.96 (CH₂), 28.65 (CH₂), 29.26 (CH₂S), 29.36 (CH₂), 29.50 (CH₂),
1335 29.65 (CH₂), 29.72 (CH₂), 29.79 (CH₂), 29.79 (CH₂), 29.82 (CH₂),
1336 29.83 (3 \times CH₂), 32.06 (CH₂), 34.40 (SCH₂), 36.55 (COCH₂),
1337 43.76 (CH₂NH), 56.12 (CH₃O), 65.91 (COOCH₂), 110.77 (C_{Ar}),
1338 114.44 (C_{Ar}), 120.91 (C_{Ar}), 130.22 (C_{Ar}), 145.23 (C_{Ar}), 146.83 (C_{Ar}),
1339 170.75 (NHCO), 170.80 (COOCH₂). HR-MS (ESI⁺), m/z : [M +
1340 H]⁺ calcd for C₂₉H₅₀NO₃S, 524.3404; found 524.3437.

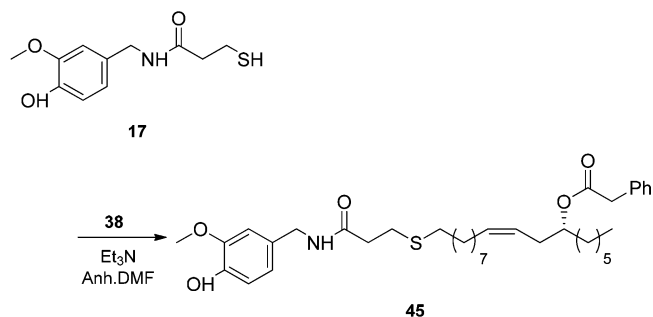


1341 **N-(4'-Hydroxy-3'-methoxybenzyl)-3-(((12Z)-tert-butyl-**
1342 **dimethylsilyl)oxy)octadec-(9Z)-en-1-ylthio)propanamide**
1343 **(43)**. General procedure VII was applied to compound **17** (100 mg,
1344 0.41 mmol), compound **35** (236 mg, 0.46 mmol), and Et₃N (120 μ L,
1345 0.82 mmol) dissolved in DMF (2 mL). Compound **43** was afforded
1346 after silica gel column chromatography (petroleum ether/EtOAc 5:5)
1347 as a yellow oil (135 mg, 53%). R_f = 0.45 (petroleum ether/EtOAc
1348 5:5). $[\alpha]_D^{20}$ = -4.71 (c 0.45, DCM). IR (ATR) ν = 3370, 3278, 2955,
1349 2917, 2849, 1726, 1269 cm^{-1} . ¹H NMR (400 MHz, CDCl₃) δ = 0.03
1350 (s, 6H, Si(CH₃)₂), 0.73–0.94 (m, 12H, SiC(CH₃)₃, CH₃), 1.14–1.42
1351 (m, 20H, CH₂), 1.47–1.67 (m, 2H, SCH₂CH₂), 2.00 (q, 2H, J = 6.4
1352 Hz, CH₂CH), 2.11–2.26 (m, 2H, CHCH₂), 2.41–2.57 (m, 4H,
1353 COCH₂, SCH₂CH₂), 2.83 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.55–
1354 3.74 (m, 1H, CH₂CHO), 3.86 (s, 3H, CH₃O), 4.34 (d, 2H, J = 5.7
1355 Hz, CH₂NH), 5.27–5.51 (m, 2H, CH=CH), 5.76 (s, 1H, OH), 6.03
1356 (s, 1H, CH₂NH), 6.79 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR
1357 (101 MHz, CDCl₃) δ = 4.57 (Si(CH₃)₃), -4.36 (Si(CH₃)₃), 14.09 (CH₃),
1358 18.13 (SiC(CH₃)₃), 22.62 (CH₂), 25.38 (CH₂), 25.91 (SiC(CH₃)₃),
1359 27.44 (CH₂CH), 27.87 (CH₂S), 28.87 (CH₂), 29.20 (CH₂), 29.28
1360 (CH₂), 29.44 (CH₂), 29.46 (CH₂), 29.60 (CH₂), 29.65 (CH₂), 31.87
1361 (CH₂), 32.43 (CH₂), 35.24 (CHCH₂), 36.84 (COCH₂, SCH₂CH₂),
1362 43.59 (CH₂NH), 55.93 (CH₃O), 72.38 (CH₂CHO), 110.66 (C_{Ar}),
1363 114.36 (C_{Ar}), 120.74 (C_{Ar}), 125.93 (CH=CH), 130.02 (C_{Ar}), 131.34
1364 (CH=CH), 145.12 (C_{Ar}), 146.71 (C_{Ar}), 171.04 (NHCO). HR-MS
1365 (ESI⁺), m/z : [M + H]⁺ calcd for C₃₅H₆₄NO₄SSi, 622.4307; found
1366 g

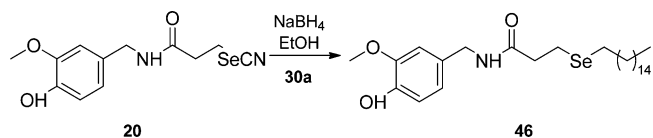


1367 **N-(4'-Hydroxy-3'-methoxybenzyl)-3-(((12Z)-hydroxy)-**
1368 **octadec-(9Z)-en-1-ylthio)propanamide (44)**. General procedure
1369 VIII was applied to compound **43** (100 mg, 0.16 mmol) in AcOH/
1370 THF/H₂O (1 mL, 6:2:2). Compound **44** was afforded after silica gel
1371 column chromatography (petroleum ether/EtOAc 6:4) as a colorless
1372 oil (66 mg, 81%). R_f = 0.62 (petroleum ether/EtOAc 5:5). $[\alpha]_D^{20}$

1373 –1.37 (c 0.4, DCM). IR (ATR) ν = 3290, 2923, 2852, 1645, 1514, 1374 1273 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.88 (t, 3H, J = 6.9 Hz, 1375 CH_3), 1.21–1.38 (m, 18H, CH_2), 1.41–1.49 (m, 4H, CH_2), 1.51–1376 1.60 (m, 2H, SCH_2CH_2), 2.04 (q, 2H, J = 6.4 Hz, CH_2CH), 2.22 (t, 1377 2H, J = 6.9 Hz, CHCH_2), 2.43–2.55 (m, 4H, COCH_2 , SCH_2CH_2), 1378 2.83 (t, 2H, J = 6.9 Hz, COCH_2CH_2), 3.56–3.65 (m, 1H, CH_2CHO), 1379 3.88 (s, 3H, CH_3O), 4.37 (d, 2H, J = 5.7 Hz, CH_2NH), 5.34–5.46 1380 (m, 1H, $\text{CH}=\text{CH}$), 5.50–5.60 (m, 1H, $\text{CH}=\text{CH}$), 6.00 (s, 1H, 1381 CH_2NH), 6.80 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ^{13}C NMR (101 1382 MHz, CDCl_3) δ = 14.23 (CH_3), 22.76 (CH_2), 25.86 (CH_2), 27.53 1383 (CH_2CH), 28.04 (CH_2S), 28.95 (CH_2), 29.28 (CH_2), 29.35 (CH_2), 1384 29.49 ($2 \times \text{CH}_2$), 29.71 (CH_2), 29.76 (CH_2), 31.98 (CH_2), 32.59 1385 (SCH_2CH_2), 35.49 (CHCH_2), 36.98 (COCH_2), 36.99 (SCH_2CH_2), 1386 43.81 (CH_2NH), 56.12 (CH_3O), 71.67 (CH_2CHO), 110.83 (C_{Ar}), 1387 114.52 (C_{Ar}), 120.94 (C_{Ar}), 125.31 ($\text{CH}=\text{CH}$), 130.13 (C_{Ar}), 133.59 1388 ($\text{CH}=\text{CH}$), 145.30 (C_{Ar}), 146.86 (C_{Ar}), 171.25 (NHCO). HR-MS 1389 (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{29}\text{H}_{50}\text{NO}_4\text{Si}$, 508.3461; found 1390 508.3451.

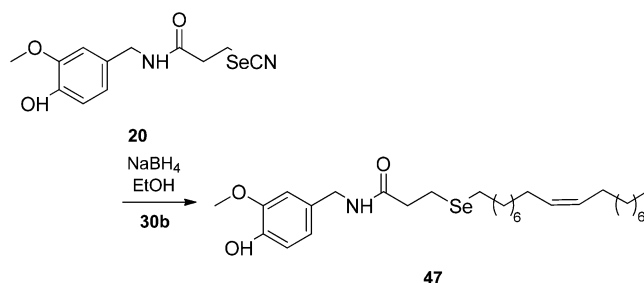


1391 **1''-Hexyl-12''-[(3'''-((4'''-hydroxy-3'''-methoxybenzyl)-**
1392 **amino)-3'''-oxopropylthio]dodec-(3''Z)-en-(1''R)-yl 2-phenyl-**
1393 **acetate (45).** General procedure VII was applied to compound 17
1394 (100 mg, 0.41 mmol), compound 38 (255 mg, 0.46 mmol), and Et_3N
1395 (115 μL , 0.82 mmol) dissolved in anhydrous DMF (2 mL).
1396 Compound 45 was afforded after silica gel column chromatography
1397 (petroleum ether/EtOAc 6:4) as a yellow oil (51 mg, 20%). R_f = 0.78
1398 (petroleum ether/EtOAc 6:4). $[\alpha]_D^{20}$ 7.90 (c 0.4, DCM). IR (ATR) ν
1399 = 3290, 2924, 2853, 1729, 1646, 1514 cm^{-1} . ^1H NMR (400 MHz,
1400 CDCl_3) δ = 0.86 (t, 3H, J = 6.9 Hz, CH_3), 1.06–1.40 (m, 18H, CH_2),
1401 1.46–1.60 (m, 4H, CH_2 , SCH_2CH_2), 1.99 (q, 2H, J = 6.4 Hz,
1402 CH_2CH), 2.19–2.35 (m, 2H, CHCH_2), 2.44–2.56 (m, 4H, COCH_2 ,
1403 SCH_2CH_2), 2.83 (t, 2H, J = 6.9 Hz, COCH_2CH_2), 3.58 (s, 2H,
1404 OCOCH_2), 3.87 (s, 3H, CH_3O), 4.36 (d, 2H, J = 5.7 Hz, CH_2NH),
1405 4.86 (p, 1H, J = 6.2 Hz, CH_2CHO), 5.22–5.32 (m, 1H, $\text{CH}=\text{CH}$),
1406 5.39–5.48 (m, 1H, $\text{CH}=\text{CH}$), 6.04 (br s, 1H, CH_2NH), 6.80 (ddd,
1407 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}), 7.21–7.34 (m, 5H, H_{Ar}). ^{13}C NMR
1408 (101 MHz, CDCl_3) δ = 14.20 (CH_3), 22.66 (CH_2), 25.33 (CH_2),
1409 27.45 (CH_2CH), 27.01 (CH_2S), 28.99 (CH_2), 29.20 (CH_2), 29.32
1410 (CH_2), 29.37 (CH_2), 29.55 (CH_2), 29.69 (CH_2), 29.73 (CH_2), 31.82
1411 (CH_2), 32.04 (CHCH_2), 32.57 (COCH_2), 33.69 (CH_2), 36.91
1412 (SCH_2CH_2), 41.90 (OCOCH_2), 43.84 (CH_2NH), 56.11 (CH_3O),
1413 74.65 (CH_2CHO), 110.81 (C_{Ar}), 114.50 (C_{Ar}), 120.94 (C_{Ar}), 124.25
1414 ($\text{CH}=\text{CH}$), 127.09 (C_{Ar}), 128.60 ($2 \times \text{C}_{\text{Ar}}$), 129.36 ($2 \times \text{C}_{\text{Ar}}$),
1415 130.06 (C_{Ar}), 132.80 ($\text{CH}=\text{CH}$), 134.46 (C_{Ar}), 145.30 (C_{Ar}), 146.84
1416 (C_{Ar}), 171.37 (NHCO), 171.48 (OCOCH_2). HR-MS (ESI^+), m/z :
1417 $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{37}\text{H}_{56}\text{NO}_5\text{S}$, 626.3879; found 626.3870.

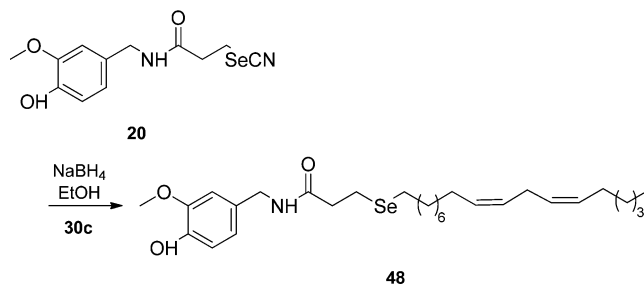


1418 **3-(Hexadecylseleno)-N-(4'-hydroxy-3'-methoxybenzyl)-**
1419 **propanamide (46).** General procedure IV was applied to compound
1420 20 (100 mg, 0.32 mmol), NaBH_4 (30 mg, 0.8 mmol), and compound
1421 30a (126 mg, 0.36 mmol) dissolved in EtOH (2 mL). Compound 46
1422 was afforded after silica gel column chromatography (petroleum

ether/EtOAc 7:3) as a yellow sticky solid (166 mg, 71%). R_f = 0.55 1423 (petroleum ether/EtOAc 7:3). IR (ATR) ν = 3504, 3317, 2917, 2848, 1424 1645, 1519 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.88 (t, 3H, J = 1425 6.9 Hz, CH_3), 1.22–1.36 (m, 26H, CH_2), 1.59–1.68 (m, 2H, 1426 SeCH_2CH_2), 2.53–2.62 (m, 4H, COCH_2 , SeCH_2CH_2), 2.83 (t, 2H, J 1427 = 6.9 Hz, CH_2Se), 3.88 (s, 3H, CH_3O), 4.36 (d, 2H, J = 5.7 Hz, 1428 CH_2NH), 5.66 (s, 1H, CH_2NH), 5.88 (br s, 1H, OH), 6.80 (ddd, 3H, 1429 J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.26 1430 (CH_3), 18.69 (CH_2Se), 22.83 (CH_2), 24.84 (SeCH_2CH_2), 29.31 1431 (CH_2), 29.49 (CH_2), 29.68 (CH_2), 29.75 (CH_2), 29.79 ($2 \times \text{CH}_2$), 1432 29.83 ($4 \times \text{CH}_2$), 30.08 (CH_2), 30.74 (CH_2), 32.06 (CH_2), 38.03 1433 (COCH_2), 43.78 (CH_2NH), 56.12 (CH_3O), 110.83 (C_{Ar}), 114.49 1434 (C_{Ar}), 120.96 (C_{Ar}), 130.20 (C_{Ar}), 145.28 (C_{Ar}), 146.84 (C_{Ar}), 171.41 1435 (NHCO). HR-MS (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{48}\text{NO}_3\text{Se}$, 1436 514.2799; found 514.2795. 1437 g

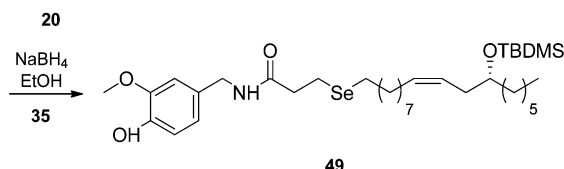
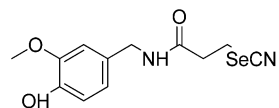


N-(4'-Hydroxy-3'-methoxybenzyl)-3-(octadec-(9''Z)-en-1- 1438 **ylseleno)propanamide (47).** General procedure IV was applied to 1439 compound 20 (200 mg, 0.64 mmol), NaBH_4 (59 mg, 1.6 mmol), and 1440 compound 30b (271 mg, 0.72 mmol) dissolved in EtOH (2 mL). 1441 Compound 47 was afforded after silica gel column chromatography 1442 (petroleum ether/EtOAc 7:3) as a yellow sticky solid (244 mg, 71%). 1443 R_f = 0.71 (petroleum ether/EtOAc 7:3). IR (ATR) ν = 3509, 3321, 1444 2919, 2850, 1646, 1519 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.88 1445 (t, 3H, J = 6.9 Hz, CH_3), 1.24–1.37 (m, 22H, CH_2), 1.60–1.68 (m, 1446 2H, SeCH_2CH_2), 2.01 (q, 4H, J = 6.4 Hz, CH_2CH , CHCH_2), 2.54– 1447 2.61 (m, 4H, COCH_2 , SeCH_2CH_2), 2.84 (t, 2H, J = 6.9 Hz, 1448 COCH_2CH_2), 3.88 (s, 3H, CH_3O), 4.37 (d, 2H, J = 5.7 Hz, 1449 CH_2NH), 5.29–5.40 (m, 2H, $\text{CH}=\text{CH}$), 5.61 (s, 1H, OH), 5.83 (br 1450 s, 1H, CH_2NH), 6.82 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ^{13}C NMR 1451 (101 MHz, CDCl_3) δ = 14.27 (CH_3), 18.70 (CH_2Se), 22.83 (CH_2), 1452 24.84 (SeCH_2CH_2), 27.35 (CH_2CH), 27.37 (CHCH_2), 29.29 (CH_2), 1453 29.40 (CH_2), 29.47 ($2 \times \text{CH}_2$), 29.58, (CH_2) 29.67 (CH_2), 29.89 1454 (CH_2), 29.92 (CH_2), 30.08 (CH_2), 30.74 (CH_2), 32.05 (CH_2), 38.06 1455 (COCH_2), 43.80 (CH_2NH), 56.14 (CH_3O), 110.83 (C_{Ar}), 114.48 1456 (C_{Ar}), 120.99 (C_{Ar}), 129.94 ($\text{CH}=\text{CH}$), 130.11 ($\text{CH}=\text{CH}$), 130.22 1457 (C_{Ar}), 145.29 (C_{Ar}), 146.84 (C_{Ar}), 171.37 (NHCO). HR-MS (ESI^+), 1458 m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{29}\text{H}_{50}\text{NO}_3\text{Se}$, 540.2956; found 540.2957. 1459 g

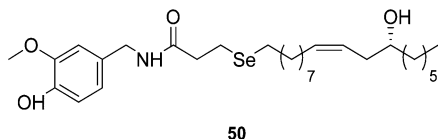


N-(4'-Hydroxy-3'-methoxybenzyl)-3-(octadeca-(9''Z,12''Z)- 1460 **dien-1-ylseleno)propanamide (48).** General procedure IV was 1461 applied to compound 20 (100 mg, 0.32 mmol), NaBH_4 (30 mg, 0.80 1462 mmol), and compound 30c (135 mg, 0.36 mmol) dissolved in EtOH 1463 (2 mL). Compound 48 was afforded after silica gel column 1464 chromatography (petroleum ether/EtOAc 7:3) as a yellowish oil 1465 (111 mg, 65%). R_f = 0.7 (petroleum ether/EtOAc 7:3). IR (ATR) ν = 1466 3288, 3008, 2923, 2852, 1644, 1514 cm^{-1} . ^1H NMR (400 MHz, 1467 CDCl_3) δ = 0.88 (t, 3H, J = 6.9 Hz, CH_3), 1.25–1.38 (m, 16H, CH_2), 1468

1469 1.59–1.68 (m, 2H, SeCH_2CH_2), 2.04 (q, 4H, $J = 6.4$ Hz, CH_2CH ,
1470 CHCH_2), 2.54–2.61 (m, 4H, COCH_2 , SeCH_2CH_2), 2.77 (t, 2H, $J =$
1471 6.9 Hz, 2H, CHCH_2CH), 2.83 (t, 2H, $J = 6.9$ Hz, COCH_2CH_2), 3.88
1472 (s, 3H, CH_3O), 4.36 (d, 2H, $J = 5.7$ Hz, CH_2NH), 5.28–5.42 (m, 4H,
1473 $2 \times \text{CH}=\text{CH}$), 5.66 (s, 1H, OH), 5.88 (br s, 1H, CH_2NH), 6.80
1474 (ddd, 3H, $J = 12.5, 9.9, 5.0$ Hz, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) δ
1475 = 14.21 (CH_3), 18.69 (CH_2), 22.70 (CH_2), 24.81 (SeCH_2CH_2), 25.77
1476 (CHCH_2CH), 27.33 (CH_2CH), 27.35 (CHCH_2), 29.26 (CH_2), 29.38
1477 (CH_2), 29.48 (CH_2), 29.56 (CH_2), 29.77 (CH_2), 30.06 (CH_2), 30.72
1478 (CH_2), 31.66 (CH_2), 38.02 (COCH_2), 43.78 (CH_2NH), 56.12
1479 (CH_3O), 110.82 (C_{Ar}), 114.48 (C_{Ar}), 120.95 (C_{Ar}), 128.04 ($\text{CH}=\text{CH}$),
1480 128.14 ($\text{CH}=\text{CH}$), 130.19 (C_{Ar}), 130.22 ($\text{CH}=\text{CH}$), 130.34
1481 ($\text{CH}=\text{CH}$), 145.28 (C_{Ar}), 146.83 (C_{Ar}), 171.39 (NHCO). HR-MS
1482 (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{29}\text{H}_{48}\text{NO}_3\text{Se}$, 538.2799; found
1483 538.2761.

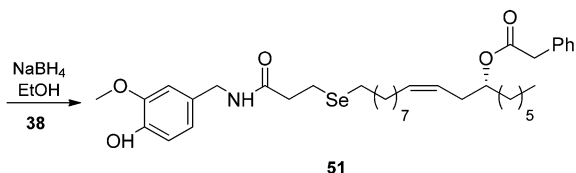
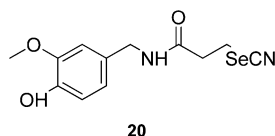


1484 **N-(4'-Hydroxy-3'-methoxybenzyl)-3-[(11''R)-tert-**
1485 **butylidimethylsilyloxy]octadec-9''Z-en-1-ylseleno]-**
1486 **propanamide (49).** General procedure IV was applied to compound
1487 **20** (100 mg, 0.32 mmol), NaBH_4 (30 mg, 0.80 mmol), and
1488 compound **35** (233 mg, 0.46 mmol) dissolved in EtOH (2 mL).
1489 Compound **49** was afforded after silica gel column chromatography
1490 (petroleum ether/EtOAc 7:3) as a yellow oil (124 mg, 58%). $R_f =$
1491 0.54 (petroleum ether/EtOAc 7:3). $[\alpha]_{\text{D}}^{20} -2.21$ (c 0.7, DCM). IR
1492 (ATR) $\nu = 3288, 2924, 2853, 1645, 1514$ cm^{-1} . ^1H NMR (400 MHz,
1493 CDCl_3) $\delta = 0.04$ (s, 6H, $\text{Si}(\text{CH}_3)_2$), 0.80–0.97 (m, 12H, $\text{Si}(\text{CH}_3)_3$,
1494 CH_3), 1.15–1.32 (m, 20H, CH_2), 1.52–1.71 (m, 2H, SeCH_2CH_2),
1495 2.01 (q, 2H, $J = 6.4$ Hz, CH_2CH), 2.18 (t, 2H, $J = 6.9$ Hz, CHCH_2),
1496 2.58 (t, 4H, $J = 6.9$ Hz, COCH_2 , SeCH_2CH_2), 2.84 (s, 2H,
1497 COCH_2CH_2), 3.58–3.70 (m, 1H, CH_2CHO), 3.89 (s, 3H, CH_3O),
1498 4.37 (d, 2H, $J = 5.7$ Hz, CH_2NH), 5.32–5.49 (m, 2H, $\text{CH}=\text{CH}$),
1499 5.58 (s, 1H, OH), 5.80 (s, 1H, CH_2NH), 6.81 (ddd, 3H, $J = 12.5, 9.9,$
1500 5.0 Hz, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) $\delta = -4.56$ (SiCH_3),
1501 -4.36 (SiCH_3), 14.09 (CH_3), 18.14 ($\text{Si}(\text{CH}_3)_3$), 18.55 (CH_2Se),
1502 22.62 (CH_2), 24.71 (SeCH_2CH_2), 25.39 (CH_2), 25.91 ($\text{Si}(\text{CH}_3)_3$),
1503 27.45 (CH_2CH), 29.13 (CH_2), 29.29 (CH_2), 29.44 (CH_2), 29.46
1504 (CH_2), 29.65 (CH_2), 29.68 (CH_2), 29.93 (CH_2), 31.88 (CH_2), 35.25
1505 (CHCH_2), 36.85 (CH_2), 37.90 (COCH_2), 43.64 (CH_2NH), 55.97
1506 (CH_3O), 72.39 (CH_2CHO), 110.65 (C_{Ar}), 114.31 (C_{Ar}), 120.83
1507 (C_{Ar}), 125.93 ($\text{CH}=\text{CH}$), 130.05 (C_{Ar}), 131.35 ($\text{CH}=\text{CH}$), 145.12
1508 (C_{Ar}), 146.66 (C_{Ar}), 171.19 (NHCO).

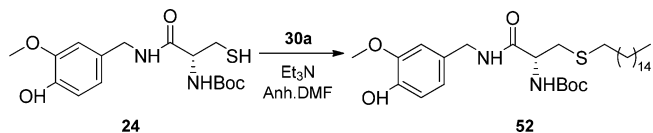


1509 **N-(4'-Hydroxy-3'-methoxybenzyl)-3-[(11''R)-hydroxy]-**
1510 **octadec-9''Z-en-1-ylseleno]propanamide (50).** General proce-
1511 dure VIII was applied to compound **49** (100 mg, 0.18 mmol) in
1512 AcOH/THF/ H_2O (1 mL, 6:2:2). Compound **50** was afforded after
1513 silica gel column chromatography (petroleum ether/EtOAc 5:5) as a
1514 pale yellow oil (79 mg, 79%). $R_f = 0.77$ (petroleum ether/EtOAc 7:3).
1515 $[\alpha]_{\text{D}}^{20} -7.88$ (c 0.3, DCM). IR (ATR) $\nu = 3288, 2923, 2852, 1646,$
1516 1514 1273 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.88$ (t, 3H, $J =$
1517 6.9 Hz, CH_3), 1.21–1.39 (m, 18H, CH_2), 1.42–1.48 (m, 2H,
1518 COHCH_2), 1.58–1.67 (m, 2H, SeCH_2CH_2), 2.04 (q, 2H, $J = 6.4$ Hz,
1519 CH_2CH), 2.20 (t, 2H, $J = 6.9$ Hz, CHCH_2), 2.53–2.61 (m, 4H,

COCH_2 , SeCH_2CH_2), 2.83 (t, 2H, $J = 6.9$ Hz, COCH_2CH_2), 3.57–
3.65 (m, 1H, CH_2CHO), 3.87 (s, 3H, CH_3O), 4.36 (d, 2H, $J = 5.7$
Hz, CH_2NH), 5.34–5.45 (m, 1H, $\text{CH}=\text{CH}$), 5.49–5.60 (m, 1H,
 $\text{CH}=\text{CH}$), 5.73 (br s, 1H, OH), 5.93 (br s, 1H, CH_2NH), 6.80 (ddd,
3H, $J = 12.5, 9.9, 5.0$ Hz, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) $\delta =$
14.23 (CH_3), 18.71 (CH_2Se), 22.76 (CH_2), 24.81 (SeCH_2CH_2),
25.86 (CH_2), 27.53 (CH_2), 29.21 (CH_2), 29.35 (CH_2), 29.49 ($2 \times$
 CH_2), 29.75 (CH_2), 30.00 (CH_2), 30.69 (SeCH_2CH_2), 31.98 (CH_2),
35.50 (CHCH_2), 36.98 (CH_2), 38.04 (COCH_2), 43.79 (CH_2NH),
56.13 (CH_3O), 71.65 (CH_2CHO), 110.85 (C_{Ar}), 114.51 (C_{Ar}),
120.97 (C_{Ar}), 125.31 ($\text{CH}=\text{CH}$), 130.21 (C_{Ar}), 133.58 ($\text{CH}=\text{CH}$),
145.29 (C_{Ar}), 146.85 (C_{Ar}), 171.39 (NHCO). HR-MS (ESI^+), m/z :
1531 $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{29}\text{H}_{50}\text{NO}_4\text{Se}$, 556.2905; found 556.2901. 1532 g

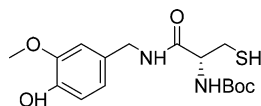


1''-Hexyl-12''-[(3'''-((4'''-hydroxy-3'''-methoxybenzyl)-
amino)-3'''-oxopropyl)seleno]dodec-(3''Z)-en-(1''R)-yl 2-phe-
nylacetaate (51). General procedure IV was applied to compound
20 (100 mg, 0.32 mmol), NaBH_4 (30 mg, 0.80 mmol), and
compound **38** (200 mg, 0.36 mmol) dissolved in EtOH (2 mL).
Compound **51** was afforded after silica gel column chromatography
(petroleum ether/EtOAc 5:5) as a yellow oil (155 mg, 72%). $R_f =$
0.58 (petroleum ether/EtOAc 5:5). $[\alpha]_{\text{D}}^{20} 14.78$ (c 1.8, DCM). IR
(ATR) $\nu = 3291, 2924, 2853, 1729, 1645, 1514$ cm^{-1} . ^1H NMR (400
MHz, CDCl_3) $\delta = 0.89$ (t, 3H, $J = 6.9$ Hz, CH_3), 1.20–1.40 (m, 18H,
 CH_2), 1.50–1.58 (m, 2H, SeCH_2CH_2), 1.61–1.71 (m, 2H,
 COHCH_2), 2.01 (q, 2H, $J = 6.4$ Hz, CH_2CH), 2.23–2.37 (m, 2H,
 CHCH_2), 2.60 (t, 4H, $J = 6.9$ Hz, COCH_2 , SeCH_2CH_2), 2.86 (t, 2H,
 $J = 6.9$ Hz, COCH_2CH_2), 3.61 (s, 2H, OCOCH_2), 3.89 (s, 3H,
 CH_3O), 4.38 (d, 2H, $J = 5.7$ Hz, CH_2NH), 4.90 (p, 1H, $J = 6.3$ Hz,
 CH_2CHO), 5.26–5.35 (m, 1H, $\text{CH}=\text{CH}$), 5.42–5.51 (m, 1H,
 $\text{CH}=\text{CH}$), 5.75 (s, 1H, OH), 5.98 (br s, 1H, CH_2NH), 6.83 (ddd,
3H, $J = 12.5, 9.9, 5.0$ Hz, H_{Ar}), 7.16–7.42 (m, 5H, H_{Ar}). ^{13}C NMR
(101 MHz, CDCl_3) $\delta = 14.18$ (CH_3), 18.68 (CH_2), 22.63 (CH_2),
24.76 (SeCH_2CH_2), 25.30 (CH_2), 27.43 (CH_2CH), 29.18 (CH_2),
29.23 (CH_2), 29.35 (CH_2), 29.53 (CH_2), 29.66 (CH_2), 30.02 (CH_2),
30.68 (CH_2), 31.79 (CH_2), 32.01 (CHCH_2), 33.66 (CH_2), 37.97
(COCH_2), 41.87 (OCOCH_2), 43.74 (CH_2NH), 56.09 (CH_3O),
74.62 (CH_2CHO), 110.82 (C_{Ar}), 114.48 (C_{Ar}), 120.91 (C_{Ar}), 124.22
($\text{CH}=\text{CH}$), 127.06 (C_{Ar}), 128.57 ($2 \times \text{C}_{\text{Ar}}$), 129.33 ($2 \times \text{C}_{\text{Ar}}$),
130.17 (C_{Ar}), 132.78 ($\text{CH}=\text{CH}$), 134.42 (C_{Ar}), 145.26 (C_{Ar}), 146.83
(C_{Ar}), 171.41 (NHCO), 171.46 (OCOCH_2). HR-MS (ESI^+), m/z :
1559 $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{37}\text{H}_{56}\text{NO}_5\text{Se}$, 674.3324; found 674.3315. 1560 g

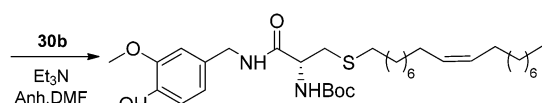


(2R)-Boc-amino-3-(hexadecylthio)-N-(4'-hydroxy-3'-
methoxybenzyl)propanamide (52). General procedure VII was
applied to compound **24** (200 mg, 0.56 mmol), compound **30a** (220
mg, 0.63 mmol), and Et_3N (0.16 mL, 1.12 mmol) in anhydrous DMF
(5 mL). Compound **52** was afforded after silica gel column
chromatography (petroleum ether/EtOAc 6:4) as a white solid
(230 mg, 71%). $R_f = 0.29$ (petroleum ether/EtOAc 5:5). Mp: 76–77
°C. $[\alpha]_{\text{D}}^{20} -2.28$ (c 0.6, DCM). IR (ATR) $\nu = 3449, 3336, 2918, 2850,$
1681, 1659, 1513 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.87$ (t, 3H,
1569

1570 $J = 6.9$ Hz, CH_3), 1.15–1.35 (m, 26H, CH_2), 1.42 (s, 9H, $\text{C}(\text{CH}_3)_3$),
 1571 1.47–1.60 (m, 2H, SCH_2CH_2), 2.52 (td, 2H, $J = 6.9$, 1.7 Hz,
 1572 SCH_2CH_2), 2.84 (dd, 1H, $J = 13.7$, 6.9 Hz, CHCH_2S), 2.98 (dd, 1H J
 1573 $= 13.7$, 5.5 Hz, CHCH_2S), 3.86 (s, 3H, CH_3O), 4.25 (d, 1H, $J = 5.7$
 1574 Hz, CH_2NH), 4.29–4.45 (m, 2H, CHCH_2S), 5.39 (d, 1H, $J = 5.7$ Hz,
 1575 CH_2NH), 5.70 (s, 1H, OH), 6.67 (t, $J = 5.5$ Hz, 1H, NHBOC), 6.78
 1576 (ddd, 3H, $J = 12.5$, 9.9, 5.0 Hz, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) δ
 1577 $= 14.25$ (CH_3), 22.82 (CH_2), 28.39 ($\text{C}(\text{CH}_3)_3$), 28.92 (CH_2), 29.36
 1578 (CH_2), 29.49 (CH_2), 29.65 (CH_2), 29.74 (CH_2), 29.78 ($2 \times \text{CH}_2$),
 1579 29.81 (CH_2), 29.82 ($4 \times \text{CH}_2$), 32.05 (CH_2), 32.82 (SCH_2CH_2),
 1580 34.61 (CHCH_2S), 43.68 (CH_2NH), 54.25 (CHCH_2S), 56.08
 1581 (CH_3O), 80.59 ($\text{C}(\text{CH}_3)_3$), 110.63 (C_{Ar}), 114.50 (C_{Ar}), 120.76
 1582 (C_{Ar}), 129.81 (C_{Ar}), 145.24 (C_{Ar}), 146.83 (C_{Ar}), 155.51 (NHCO_2),
 1583 170.58 (NHCO). HR-MS (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd for
 1584 $\text{C}_{32}\text{H}_{57}\text{N}_2\text{O}_5\text{S}$, 581.3988; found 581.3978.

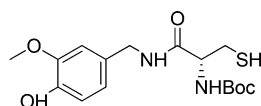


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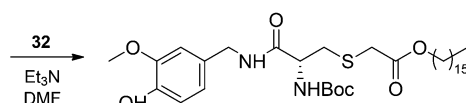


53

1585 **(2R)-Boc-amino-N-(4'-hydroxy-3'-methoxybenzyl)-3-(octa-**
 1586 **dec-9'Z-en-1-ylthio)propanamide (53).** General procedure VII
 1587 was applied to compound 24 (100 mg, 0.42 mmol), compound 30b
 1588 (179 mg, 0.47 mmol), and Et_3N (117 μL , 0.84 mmol) dissolved
 1589 in DMF (2 mL). Compound 53 was afforded after silica gel column
 1590 chromatography (petroleum ether/EtOAc 7:3) as a white solid (127
 1591 mg, 50%). Mp: 43–44 °C. $R_f = 0.58$ (petroleum ether/EtOAc 7:3).
 1592 $[\alpha]_D^{20}$ 0.26 (c 1.2, DCM). IR (ATR) $\nu = 3450$, 3333, 2918, 2850,
 1593 1514, 1240 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta = 0.88$ (t, 3H, $J =$
 1594 6.9 Hz, CH_3), 1.18–1.38 (m, 22H, CH_2), 1.42 (s, 9H, $\text{C}(\text{CH}_3)_3$),
 1595 1.48–1.61 (m, 2H, SCH_2CH_2), 2.01 (q, 4H, $J = 6.4$ Hz, CH_2CH ,
 1596 CHCH_2), 2.45–2.58 (m, 2H, SCH_2CH_2), 2.84 (dd, 1H, $J = 13.7$, 6.9
 1597 Hz, CHCH_2S), 3.00 (dd, 1H, $J = 13.7$, 5.5 Hz, CHCH_2S), 3.88 (s,
 1598 3H, CH_3O), 4.24 (dd, 1H, $J = 12.5$, 6.1 Hz, CH_2NH), 4.30–4.48 (m,
 1599 2H, CHCH_2S), 5.22–5.44 (m, 3H, $\text{CH}=\text{CH}$, CH_2NH), 5.59 (s, 1H,
 1600 OH), 6.61 (t, 1H, $J = 5.5$ Hz, NHBOC), 6.80 (ddd, 3H, $J = 12.5$, 9.9,
 1601 5.0 Hz, H_{Ar}). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 14.10$ (CH_3), 22.66
 1602 (CH_2), 27.18 (CH_2CH), 27.20 (CHCH_2), 28.24 ($\text{C}(\text{CH}_3)_3$), 28.76
 1603 (CH_2), 29.18 (CH_2), 29.23 (CH_2), 29.29 (CH_2), 29.30 (CH_2), 29.40
 1604 (SCH_2CH_2), 29.50 (CH_2), 29.59 (CH_2), 29.68 (CH_2), 29.73 (CH_2),
 1605 29.75 (CH_2), 31.88 (CH_2), 32.66 (SCH_2CH_2), 34.44 (CHCH_2S),
 1606 43.55 (CH_2NH), 54.12 (CHCH_2S), 55.94 (CH_3O), 80.57
 1607 ($\text{C}(\text{CH}_3)_3$), 110.45 (C_{Ar}), 114.31 (C_{Ar}), 120.64 (C_{Ar}), 129.68
 1608 (C_{Ar}), 129.76 ($\text{CH}=\text{CH}$), 129.95 ($\text{CH}=\text{CH}$), 145.10 (C_{Ar}),
 1609 146.65 (C_{Ar}), 155.55 (NHCO_2), 170.37 (NHCO). HR-MS (ESI^+),
 1610 m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{34}\text{H}_{59}\text{N}_2\text{O}_5\text{S}$, 607.4145; found 607.4138.



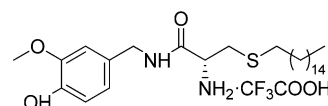
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54

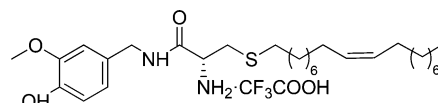
1611 **Hexadecyl 2-[(2R)-Boc-amino-3'-((4'-hydroxy-3'-**
 1612 **methoxybenzyl)amino)-3'-oxopropyl]thiolacetate (54).** Gen-
 1613 eral procedure VII was applied to compound 24 (200 mg, 0.56 mmol),
 1614 compound 35 (258 mg, 0.63 mmol), and Et_3N (160 μL , 1.12 mmol)
 1615 dissolved in anhydrous DMF (2 mL). Compound 54 was afforded

after silica gel column chromatography (petroleum ether/EtOAc 7:3) 1616
 as a white solid (282 mg, 79%). Mp: 74–75 °C. $R_f = 0.75$ (petroleum 1617
 ether/EtOAc 7:3). $[\alpha]_D^{20} -8.04$ (c 1, MeOH). IR (ATR) $\nu = 3493$, 1618
 3326, 2917, 2849, 1655, 1518 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta =$ 1619
 0.88 (t, $J = 6.9$ Hz, 3H, CH_3), 1.17–1.35 (m, 26H, CH_2), 1.42 (s, 9H, 1620
 $\text{C}(\text{CH}_3)_3$), 1.55–1.65 (m, 2H, $\text{COOCH}_2\text{CH}_2$), 2.88 (dd, 1H, $J =$ 1621
 13.7, 6.9 Hz, CHCH_2S), 3.07 (dd, 1H, $J = 13.7$, 6.9 Hz, CHCH_2S), 1622
 3.35 (s, 2H, SCH_2), 3.87 (s, 3H, CH_3O), 4.07 (t, 2H, $J = 6.9$ Hz, 1623
 $\text{COOCH}_2\text{CH}_2$), 4.25–4.49 (m, 3H, COCHCH_2 , CH_2NH), 5.47– 1624
 5.69 (m, 2H, CH_2NH , OH), 6.73–6.87 (m, 3H, H_{Ar}), 7.04 (t, 1H, $J =$ 1625
 5.0 Hz, NHBOC). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 14.09$ (CH_3), 1626
 22.66 (CH_2), 25.78 (CH_2), 28.26 ($\text{C}(\text{CH}_3)_3$), 28.44 (CH_2), 29.20 1627
 CH_2), 29.33 (CH_2), 29.48 (CH_2), 29.55 (CH_2), 29.62 (CH_2), 29.63 1628
 CH_2), 29.65 (CH_2), 29.67 ($3 \times \text{CH}_2$), 31.90 (CH_2), 34.70 1629
 SCH_2CH_2), 35.89 (CHCH_2S), 43.50 (CH_2NH), 53.59 (CHCH_2S), 1630
 55.93 (CH_3O), 66.07 (COOCH_2), 80.35 ($\text{C}(\text{CH}_3)_3$), 110.42 (C_{Ar}), 1631
 114.28 (C_{Ar}), 120.61 (C_{Ar}), 129.70 (C_{Ar}), 145.00 (C_{Ar}), 146.62 (C_{Ar}), 1632
 155.46 (NHCO_2), 170.00 (NHCO), 171.34 (COOCH_2). HR-MS 1633
 (ESI^+) , m/z : $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{34}\text{H}_{59}\text{N}_2\text{O}_7\text{S}$, 639.4043; found 1634
 639.4040. 1635 g



55

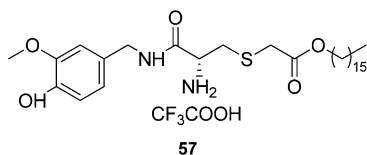
2-(Hexadecylthio)-1-[N-(4'-hydroxy-3'-methoxybenzyl)- 1636
carbamoyl]-(1R)-ethylammonium Trifluoroacetate (55). Gen- 1637
 eral procedure IX was applied to compound 52 (200 mg, 0.34 mmol), 1638
 TFA (0.26 mL, 3.4 mmol) in DCM (1 mL). Compound 55 was 1639
 afforded after flushing nitrogen and drying in vacuo as a yellow oil 1640
 (195 mg, quantitative). $[\alpha]_D^{20} -6.67$ (c 0.6, DCM). IR (ATR) $\nu =$ 1641
 3093, 2921, 2852, 1779, 1667, 1153 cm^{-1} . ^1H NMR (400 MHz, 1642
 CDCl_3) $\delta = 0.88$ (t, 3H, $J = 6.9$ Hz, CH_3), 1.21–1.31 (m, 26H, CH_2), 1643
 1.45–1.54 (m, 2H, SCH_2CH_2), 2.48 (t, 2H, $J = 6.9$ Hz, SCH_2CH_2), 1644
 2.85–3.03 (m, 2H, CHCH_2S), 3.83 (s, CH_3O), 4.22–4.38 (m, 3H, 1645
 CHCH_2S , CH_2NH), 6.52 (br s, 2H, NH_2), 6.68–6.85 (m, 4H, OH, 1646
 H_{Ar}), 7.55 (t, 1H, $J = 5.0$ Hz, CH_2NH). ^{13}C NMR (101 MHz, 1647
 CDCl_3) $\delta = 14.26$ (CH_3), 22.85 (CH_2), 28.82 (CH_2), 29.30 (CH_2), 1648
 29.32 (CH_2), 29.52 (CH_2), 29.65 (CH_2), 29.74 ($2 \times \text{CH}_2$), 29.84 1649
 CH_2), 29.86 ($4 \times \text{CH}_2$), 32.08 (CH_2), 32.50 (SCH_2CH_2), 33.06 1650
 CHCH_2S), 44.38 (CH_2NH), 52.72 (CHCH_2S), 56.01 (CH_3O), 1651
 110.67 (C_{Ar}), 114.71 (C_{Ar}), 116.86 (CF_3COOH), 120.92 (C_{Ar}), 1652
 128.31 (C_{Ar}), 145.52 (C_{Ar}), 146.95 (C_{Ar}), 161.37 (CF_3COOH), 1653
 167.54 (NHCO). HR-MS (ESI^+), m/z : $[\text{M} + \text{H}]^+$ calcd for 1654
 $\text{C}_{27}\text{H}_{49}\text{N}_2\text{O}_3\text{S}$, 481.3458; found 481.3497. 1655 g



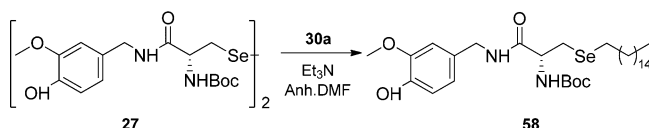
56

1-[N-(4'-Hydroxy-3'-methoxybenzyl)carbamoyl]-2-(octa- 1656
dec-9'Z-en-1-ylthio)-(1R)-ethylammonium Trifluoroacetate 1657
(56). General procedure IX was applied to compound 53 (100 mg, 1658
 0.16 mmol), TFA (120 μL , 1.64 mmol) in DCM (1 mL). Compound 1659
 56 was afforded after flushing nitrogen and drying in vacuo as a yellow 1660
 oil (98 mg, quantitative). $[\alpha]_D^{20}$ 0.62 (c 2.2, DCM). IR (ATR) $\nu =$ 1661
 2922, 2853, 1662, 1199, 1133 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) $\delta =$ 1662
 0.87 (t, 3H, $J = 6.9$ Hz, CH_3), 1.21–1.35 (m, 22H, CH_2), 1.43–1.51 1663
 (m, 2H, SCH_2CH_2), 2.00 (q, 4H, $J = 6.4$ Hz, CH_2CH , CHCH_2), 2.45 1664
 (t, 2H, $J = 6.9$ Hz, SCH_2CH_2), 2.94 (d, 2H, $J = 6.0$ Hz, CHCH_2S), 1665
 3.78 (s, 3H, CH_3O), 4.13–4.34 (m, 3H, CHCH_2S , CH_2NH), 5.26– 1666
 5.43 (m, 2H, $\text{CH}=\text{CH}$), 6.70 (ddd, 3H, $J = 12.5$, 9.9, 5.0 Hz, H_{Ar}), 1667
 7.87 (t, 1H, $J = 5.0$ Hz, CH_2NH). ^{13}C NMR (101 MHz, CDCl_3) $\delta =$ 1668
 14.25 (CH_3), 22.83 (CH_2), 27.37 (CH_2CH , CHCH_2), 28.86 (CH_2), 1669
 29.34 (CH_2), 29.41 (CH_2), 29.44 (CH_2), 29.46 (CH_2), 29.47 (CH_2), 1670
 29.61 (CH_2), 29.68 (CH_2), 29.82 (CH_2), 29.85 (CH_2), 29.92 (CH_2), 1671
 32.05 (CH_2), 32.66 (SCH_2CH_2), 32.96 (CHCH_2S), 44.0 (CH_2NH), 1672

1673 52.77 (CHCH₂S), 55.96 (CH₃O), 110.71 (C_{Ar}), 114.67 (C_{Ar}), 120.74
1674 (C_{Ar}), 128.82 (C_{Ar}), 129.90 (CH=CH), 130.11 (CH=CH), 145.27
1675 (C_{Ar}), 146.93 (C_{Ar}), 167.76 (NHCO). HR-MS (ESI⁺), *m/z*: [M +
1676 H]⁺ calcd for C₃₁H₅₁N₂O₃S, 507.3615; found 507.3616.

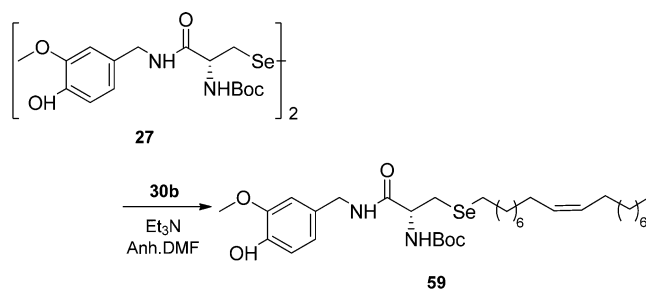


1677 **2'-Hexadecyloxy-1-[N-(4'-hydroxy-3'-methoxybenzyl)]-**
1678 **carbamoyl-2-[(oxoethyl)thio]ethan-(1R)-ammonium Trifluor-**
1679 **oacetate (57).** General procedure IX was applied to compound 54
1680 (200 mg, 0.31 mmol), TFA (240 μ L, 3.1 mmol) in DCM (1 mL).
1681 Compound 57 was afforded after flushing nitrogen and drying *in*
1682 *vacuo* as a yellow oil (201 mg, quantitative). [α]_D²⁰ -7.53 (*c* 0.4,
1683 MeOH). IR (ATR) ν = 2917, 2850, 1662, 1176, 1131 cm⁻¹. ¹H NMR
1684 (400 MHz, CDCl₃) δ = 0.88 (t, *J* = 6.9 Hz, 3H, CH₃), 1.18–1.34 (m,
1685 26H, CH₂), 1.53–1.64 (m, 2H, COOCH₂CH₂), 2.98–3.14 (m, 2H,
1686 CHCH₂S), 3.37 (s, 2H, SCH₂), 3.82 (s, 3H, CH₃O), 3.99–4.11 (m,
1687 2H, COOCH₂CH₂), 4.22–4.43 (m, 3H, COCHCH₂, H₂NH), 6.67–
1688 6.83 (m, 3H, H_{Ar}), 7.94 (t, 1H, *J* = 5.0 Hz, CH₂NH). ¹³C NMR (101
1689 MHz, CDCl₃) δ = 14.26 (CH₃), 22.84 (CH₂), 25.87 (CH₂), 28.43
1690 (CH₂), 29.35 (CH₂), 29.51 (2 \times CH₂), 29.64 (CH₂), 29.73 (CH₂),
1691 29.81 (CH₂), 29.83 (CH₂), 29.85 (3 \times CH₂), 32.08 (CH₂), 34.65
1692 (CH₂), 34.95 (CH₂), 44.24 (CH₂NH), 53.08 (CHCH₂S), 55.99
1693 (CH₃O), 67.26 (COOCH₂), 110.62 (C_{Ar}), 114.64 (C_{Ar}), 120.80
1694 (C_{Ar}), 128.61 (C_{Ar}), 145.35 (C_{Ar}), 146.91 (C_{Ar}), 167.33 (NHCO),
1695 172.72 (COOCH₂). HR-MS (ESI⁺), *m/z*: [M + H]⁺ calcd for
1696 C₂₉H₅₁N₂O₅S, 539.3513; found 539.3557.

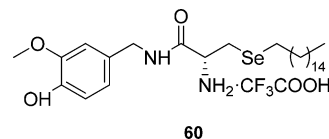


1697 **(2R)-Boc-amino-3-(hexadecylseleno)-N-(4'-hydroxy-3'-**
1698 **methoxybenzyl)propanamide (58).** General procedure III was
1699 applied to compound 27 (200 mg, 0.25 mmol), NaBH₄ (24 mg, 0.62
1700 mmol), and compound 30a (197 mg, 0.56 mmol) dissolved in EtOH
1701 (2 mL). Compound 58 was afforded after silica gel column
1702 chromatography (petroleum ether/EtOAc 7:3) as a white solid
1703 (231 mg, 74%). *R*_f = 0.37 (petroleum ether/EtOAc 6:4). Mp: 75–76
1704 °C. [α]_D²⁰ -5.24 (*c* 1.3, DCM). IR (ATR) ν = 3281, 3008, 2924, 2854,
1705 1666, 1516 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 1H, *J* =
1706 6.9 Hz, CH₃), 1.17–1.38 (m, 26H, CH₂), 1.42 (s, 9H, *J* = 4.9 Hz,
1707 C(CH₃)₃), 1.58–1.69 (m, 2H, SeCH₂CH₂), 2.46–2.67 (m, 2H,
1708 SeCH₂CH₂), 2.83 (dd, 1H, *J* = 12.8, 6.9 Hz, CHCH₂Se), 3.05 (dd,
1709 1H, *J* = 12.8, 5.2 Hz, CHCH₂Se), 3.88 (s, 3H, CH₃O), 4.22–4.36 (m,
1710 1H, CHCH₂Se), 4.37 (d, 2H, *J* = 5.7 Hz, CH₂NH), 5.33 (s, 1H,
1711 CH₂NH), 5.58 (s, 1H, OH), 6.55 (t, 1H, *J* = 5.5 Hz, NHBoc), 6.80
1712 (ddd, 3H, *J* = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ
1713 = 14.10 (CH₃), 22.67 (CH₂), 25.37 (SeCH₂CH₂), 25.88 (CHCH₂Se),
1714 28.24 (C(CH₃)₃), 29.13 (CH₂), 29.34 (CH₂), 29.51 (CH₂), 29.59
1715 (CH₂), 29.63 (3 \times CH₂), 29.66 (CH₂), 29.67 (2 \times CH₂), 29.81
1716 (CH₂), 30.51 (CH₂), 31.90 (CH₂), 43.54 (CH₂NH), 54.63
1717 (CHCH₂Se), 55.95 (CH₃O), 80.37 (C(CH₃)₃), 110.49 (C_{Ar}),
1718 114.32 (C_{Ar}), 120.65 (C_{Ar}), 129.68 (C_{Ar}), 145.10 (C_{Ar}), 146.67
1719 (C_{Ar}), 155.30 (NHCO₂), 170.46 (NHCO). HR-MS (ESI⁺), *m/z*: [M
1720 + H]⁺ calcd for C₃₂H₅₇N₂O₅Se, 629.3433; found 629.3431.

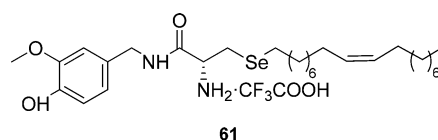
1721 **(2R)-Boc-amino-N-(4'-hydroxy-3'-methoxybenzyl)-3-(octa-**
1722 **dec-(9'Z)-en-1-ylseleno)propanamide (59).** General procedure
1723 III was applied to compound 27 (200 mg, 0.25 mmol), NaBH₄ (24
1724 mg, 0.62 mmol) and compound 30b (212 mg, 0.56 mmol) dissolved
1725 in EtOH (2 mL). Compound 59 was afforded after silica gel column
1726 chromatography (petroleum ether/EtOAc 6:4) as a yellow oil (287
1727 mg, 88%). *R*_f = 0.66 (petroleum ether/EtOAc 7:3). [α]_D²⁰ -4.90 (*c*
1728 1.4, DCM). IR (ATR) ν = 3444, 3337, 2919, 2850, 1676, 1511 cm⁻¹.
1729 ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, *J* = 6.9 Hz, CH₃),



1.16–1.39 (m, 22H, CH₂), 1.42 (s, 9H, C(CH₃)₃), 1.57–1.68 (m, 1730
2H, SeCH₂CH₂), 2.01 (q, 4H, *J* = 6.4 Hz, CH₂CH, CHCH₂), 2.44–
1731 2.70 (m, 2H, SeCH₂CH₂), 2.83 (dd, 1H, *J* = 12.8, 6.9 Hz, CHCH₂Se),
1732 3.05 (dd, 1H, *J* = 12.8, 5.2 Hz, CHCH₂Se), 3.88 (s, 3H, CH₃O),
1733 4.26–4.35 (m, CHCH₂Se), 4.37 (d, 2H, *J* = 5.7 Hz, CH₂NH),
1734 5.23–5.43 (m, 3H, CH=CH, CH₂NH), 5.60 (s, 1H, OH),
1735 6.56 (t, 1H, *J* = 5.5 Hz, NHBoc), 6.79 (ddd, 3H, *J* = 12.5, 9.9, 5.0 Hz,
1736 H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.10 (CH₃), 22.66 (CH₂),
1737 25.36 (SeCH₂CH₂), 25.90 (CHCH₂Se), 27.18 (CH₂CH), 27.20
1738 (CHCH₂), 28.24 (C(CH₃)₃), 29.11 (CH₂), 29.23 (CH₂), 29.30 (2 \times 1739
1740 CH₂), 29.41 (CH₂), 29.50 (CH₂), 29.72 (CH₂), 29.75 (CH₂), 29.80
1741 (CH₂), 30.50 (CH₂), 31.88 (CH₂), 43.55 (CH₂NH), 54.42
1742 (CHCH₂Se), 55.95 (CH₃O), 80.57 (C(CH₃)₃), 110.48 (C_{Ar}),
1743 114.31 (C_{Ar}), 120.66 (C_{Ar}), 129.68 (C_{Ar}), 129.76 (CH=CH),
1744 129.95 (CH=CH), 145.10 (C_{Ar}), 146.65 (C_{Ar}), 155.54 (NHCO₂),
1745 170.43 (NHCO). HR-MS (ESI⁺), *m/z*: [M + H]⁺ calcd for
1746 C₃₄H₅₉N₂O₅Se, 655.3589; found 655.3583.



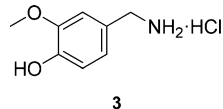
1747 **2-(Hexadecylseleno)-1-[N-(4'-hydroxy-3'-methoxybenzyl)-**
1748 **carbamoyl]-(1R)-ethylammonium Trifluoroacetate (60).** Gen-
1749 eral procedure IX was applied to compound 58 (200 mg, 0.32 mmol),
1750 TFA (240 μ L, 3.2 mmol) in DCM (1 mL). Compound 60 was
1751 afforded after flushing nitrogen and drying *in vacuo* as a yellow oil
1752 (201 mg, quantitative). [α]_D²⁰ 0.65 (*c* 1.4, MeOH). IR (ATR) ν = 1752
1753 3425, 3316, 2916, 2849, 1658, 1187 cm⁻¹. ¹H NMR (400 MHz,
1754 CDCl₃) δ = 0.88 (t, 3H, *J* = 6.9 Hz, CH₃), 1.20–1.34 (m, 26H, CH₂),
1755 1.53–1.61 (m, 2H, SeCH₂CH₂), 2.55 (t, 2H, *J* = 6.9 Hz, SeCH₂CH₂),
1756 2.85–3.01 (m, 2H, CHCH₂Se), 3.82 (s, 3H, CH₃O), 4.21–4.37 (m,
1757 3H, CHCH₂Se, CH₂NH), 6.73 (ddd, 3H, *J* = 12.5, 9.9, 5.0 Hz, H_{Ar}),
1758 7.53 (t, 1H, *J* = 5.0 Hz, CH₂NH), 7.98 (br s, 1H, OH), 9.42 (br s, 2H,
1759 NH₂). ¹³C NMR (101 MHz, CDCl₃) δ = 14.25 (CH₃), 22.84 (CH₂),
1760 23.51 (CHCH₂Se), 25.89 (CH₂), 27.72 (CH₂), 29.22 (CH₂), 29.51
1761 (CH₂), 29.64 (CH₂), 29.73 (CH₂), 29.80 (CH₂), 29.81 (CH₂), 29.83
1762 (CH₂), 29.85 (3 \times CH₂), 30.19 (CH₂), 32.08 (CH₂), 44.50
1763 (CH₂NH), 53.54 (CHCH₂Se), 55.94 (CH₃O), 110.72 (C_{Ar}),
1764 114.77 (C_{Ar}), 116.78 (CF₃COOH), 120.96 (C_{Ar}), 128.09 (C_{Ar}),
1765 145.43 (C_{Ar}), 146.96 (C_{Ar}), 160.81–162.0 (CF₃COOH), 167.72
1766 (NHCO). HR-MS (ESI⁺), *m/z*: [M + H]⁺ calcd for C₂₇H₄₉N₂O₃Se,
1767 529.2903; found 529.2905.



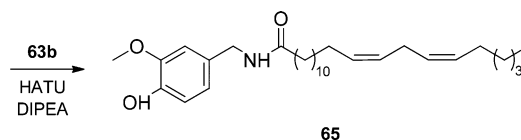
1768 **1-[N-(4'-Hydroxy-3'-methoxybenzyl)carbamoyl]-2-(octa-**
1769 **dec-(9'Z)-en-1-ylseleno)-(1R)-ethylammonium Trifluoroac-**
1770 **tate (61).** General procedure IX was applied to compound 59 (200
1771 mg, 0.30 mmol), TFA (230 μ L, 3 mmol) in DCM (1 mL).
1772 Compound 61 was afforded after flushing nitrogen and drying *in*
1773 *vacuo* as a yellow oil (199 mg, quantitative). [α]_D²⁰ -2.58 (*c* 0.3,
1774 DCM). IR (ATR) ν = 2922, 2853, 1666, 1199 cm⁻¹. ¹H NMR (400

1775 MHz, CDCl₃) δ = 0.87 (t, 3H, J = 6.9 Hz, CH₃), 1.22–1.34 (m, 22H, CH₂), 1.51–1.61 (m, 2H, SeCH₂CH₂), 2.00 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.54 (t, 2H, J = 6.9 Hz, SeCH₂CH₂), 2.93 (d, 2H, J = 6.4 Hz, CHCH₂Se), 3.81 (s, 3H, CH₃O), 4.17–4.34 (m, 3H, CHCH₂Se, CH₂NH), 5.28–5.42 (m, 2H, CH=CH), 6.72 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}), 7.64 (t, 1H, J = 5.5 Hz, CH₂NH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.26 (CH₃), 22.83 (CH₂), 23.56 (CHCH₂Se), 25.89 (SeCH₂CH₂), 27.37 (CH₂CH, CHCH₂), 29.24 (CH₂), 29.42 (CH₂), 29.47 (CH₂), 29.47 (CH₂), 29.59 (CH₂), 29.68 (CH₂), 29.87 (CH₂), 29.91 (CH₂), 29.92 (CH₂), 30.26 (CH₂), 32.06 (CH₂), 44.17 (CH₂NH), 53.40 (CHCH₂Se), 56.00 (CH₃O), 110.73 (C_{Ar}), 114.68 (C_{Ar}), 120.87 (C_{Ar}), 128.61 (C_{Ar}), 129.90 (CH=CH), 178.77 (CH=CH), 145.38 (C_{Ar}), 146.93 (C_{Ar}), 167.76 (NHCO). HR-MS (ESI⁺), m/z : [M + H]⁺ calcd for C₂₉H₅₁N₂O₃Se, 555.3059; found 555.3067.

MHz, CDCl₃) δ = 14.25 (CH₃), 22.82 (CH₂), 25.94 (COCH₂CH₂), 27.35 (CH₂CH, CHCH₂), 29.46 (3 × CH₂), 29.50 (CH₂), 29.66 (2 × CH₂), 29.69 (CH₂), 29.75 (2 × CH₂), 29.83 (CH₂), 29.91 (CH₂), 29.92 (CH₂), 32.04 (CH₂), 37.00 (COCH₂CH₂), 43.66 (CH₂NH), 56.05 (CH₃O), 110.82 (C_{Ar}), 114.50 (C_{Ar}), 120.92 (C_{Ar}), 130.00 (CH=CH), 130.04 (CH=CH), 130.51 (C_{Ar}), 145.26 (C_{Ar}), 146.83 (C_{Ar}), 173.04 (COCH₂CH₂). HR-MS (ESI⁺), m/z : [M + Na]⁺ calcd for C₃₀H₅₁NO₃Na, 496.3767; found 496.3756.

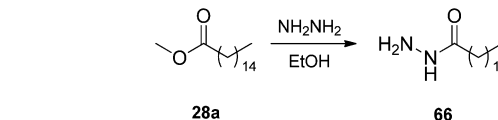


3



65

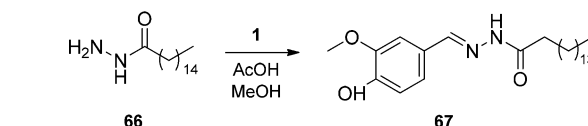
N-(4'-Hydroxy-3'-methoxybenzyl)docosa-(13Z,16Z)-dienamide (65). General procedure I was applied to a solution of compound 63b (23 mg, 0.07 mmol) dissolved in DMF (1 mL), amine hydrochloride salt 3 (15 mg, 0.08 mmol), DIPEA (38 μ L, 0.21 mmol), and HATU (39 mg, 0.10 mmol). Compound 65 was afforded after silica gel column chromatography (petroleum ether/EtOAc 6:4) as a sticky oil (21 mg, 63%). R_f = 0.40 (petroleum ether/EtOAc 5:5). IR (ATR) ν = 3489, 3316, 3302, 2919, 2849, 1639, 1518 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.89 (t, 3H, J = 6.9 Hz, CH₃), 1.24–1.38 (m, 22H, CH₂), 1.59–1.70 (m, 2H, COCH₂CH₂), 2.05 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.19 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 2.77 (t, 2H, J = 6.9 Hz, CHCH₂CH), 3.87 (s, 3H, CH₃O), 4.35 (d, 2H, J = 5.7 Hz, CH₂NH), 5.28–5.43 (m, 4H, 2 × CH=CH), 5.59–5.72 (m, 2H, OH, CH₂NH), 6.79 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.22 (CH₃), 22.72 (CH₂), 25.78 (CHCH₂CH), 25.94 (COCH₂CH₂), 27.35 (CH₂CH), 27.39 (CHCH₂), 29.48 (2 × CH₂), 29.50 (2 × CH₂), 29.65 (CH₂), 29.70 (CH₂), 29.75 (2 × CH₂), 29.83 (CH₂), 31.68 (CH₂), 37.03 (COCH₂CH₂), 43.68 (CH₂NH), 56.08 (CH₃O), 110.82 (C_{Ar}), 114.49 (C_{Ar}), 120.95 (C_{Ar}), 128.09 (2 × CH=CH), 130.31 (CH=CH), 130.34 (CH=CH), 130.53 (C_{Ar}), 145.26 (C_{Ar}), 146.82 (C_{Ar}), 173.05 (COCH₂CH₂). HR-MS (ESI⁺), m/z : [M + Na]⁺ calcd for C₃₀H₄₉NO₃Na, 494.3610; found 494.3606.



28a

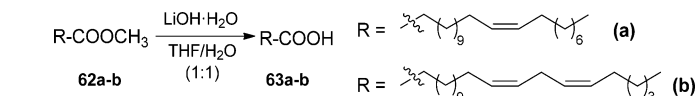
66

Hexadecanohydrazide (66). To a suspension of methyl palmitate 28a (1 g, 3.69 mmol) in ethanol (20 mL), hydrazine hydrate (64%, 370 μ L, 7.38 mmol, 2 equiv) was added. Then, the mixture was heated at 150 °C for 3 h. The mixture was cooled, and the solid precipitated was recovered by filtration to yield compound 66²¹ as a white solid (800 mg, 80%). Mp: 110–111 °C. IR (ATR) ν = 3315, 3288, 3199, 2956, 2917, 2848, 1627, 1535 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 6.9 Hz, CH₃), 1.06–1.42 (m, 24H, CH₂), 1.55–1.74 (m, 2H, NHCOCH₂CH₂), 2.08–2.23 (m, 2H, NHCOCH₂CH₂), 3.89 (br s, 2H, NH₂NH), 6.66 (s, 1H, NH₂NH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.10 (CH₃), 22.67 (CH₂), 25.48 (NHCOCH₂CH₂), 29.25 (CH₂), 29.27 (CH₂), 29.34 (CH₂), 29.44 (CH₂), 29.57 (CH₂), 29.62 (CH₂), 29.63 (CH₂), 29.64 (CH₂), 29.66 (CH₂), 29.67 (CH₂), 31.90 (CH₂), 34.59 (NHCOCH₂CH₂), 173.97 (NHCOCH₂).



66

67



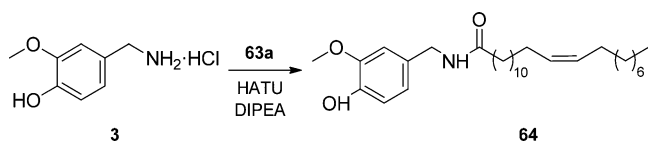
62a-b

63a-b

R =

(13Z)-Docosenoic Acid (63a). General procedure II was applied to a solution of methyl (13Z)-docosenoate 62a (500 μ L, 1.23 mmol) dissolved in THF/H₂O (6 mL, 1:1) and LiOH·H₂O (155 mg, 3.70 mmol) to yield compound 63a as a white solid (360 mg, 86%). Mp: 30–32 °C. IR (ATR) ν = 2916, 2849, 1691, 1471 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 6.9 Hz, CH₃), 1.17–1.39 (m, 28H, CH₂), 1.58–1.70 (m, 2H, OHCOCH₂CH₂), 2.02 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.34 (t, 2H, J = 6.9 Hz, OHCOCH₂CH₂), 5.24–5.42 (m, 2H, CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.09 (CH₃), 22.67 (CH₂), 24.67 (OHCOCH₂CH₂), 27.20 (CH₂CH, CHCH₂), 29.05 (CH₂), 29.23 (CH₂), 29.30 (CH₂), 29.31 (2 × CH₂), 29.42 (CH₂), 29.51 (CH₂), 29.53 (CH₂), 29.57 (CH₂), 29.59 (CH₂), 29.76 (2 × CH₂), 31.90 (CH₂), 34.01 (OHCOCH₂CH₂), 129.86 (CH=CH), 129.89 (CH=CH), 179.89 (OHCOCH₂CH₂).

(13Z,16Z)-Docosadienoic Acid (63b). General procedure II was applied to a solution of methyl (13Z,16Z)-docosadienoate 62b (25 μ L, 0.07 mmol) in THF/H₂O (1 mL, 1:1) and LiOH·H₂O (9 mg, 0.21 mmol) to yield compound 63b²⁰ as a sticky solid (23 mg, quantitative). IR (ATR) ν = 2922, 2853, 1708, 1458 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.89 (t, 3H, J = 6.9 Hz, CH₃), 1.17–1.45 (m, 22H, CH₂), 1.53–1.72 (m, 2H, COCH₂CH₂), 2.05 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.34 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 2.77 (t, 2H, J = 6.9 Hz, CHCH₂CH), 5.24–5.44 (m, 4H, 2 × CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.07 (CH₃), 22.58 (CH₂), 24.68 (OHCOCH₂CH₂), 25.63 (CHCH₂CH), 27.20 (CH₂CH), 27.24 (CHCH₂), 29.07 (CH₂), 29.24 (CH₂), 29.32 (CH₂), 29.36 (CH₂), 29.43 (CH₂), 29.54 (CH₂), 29.58 (CH₂), 29.60 (CH₂), 29.68 (CH₂), 31.53 (CH₂), 34.05 (OHCOCH₂CH₂), 127.94 (2 × CH=CH), 130.17 (2 × CH=CH), 179.96 (OHCOCH₂CH₂).

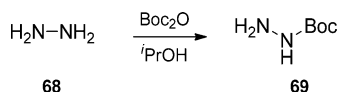


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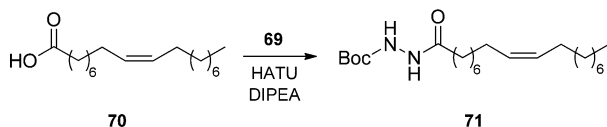
64

N-(4'-Hydroxy-3'-methoxybenzyl)docosa-(13Z)-enamide (64). General procedure I was applied to a solution of compound 63a (200 mg, 0.59 mmol) in anhydrous DMF (5 mL), amine hydrochloride salt 3 (123 mg, 0.65 mmol), DIPEA (309 μ L, 1.77 mmol), and HATU (337 mg, 0.88 mmol). Compound 64 was afforded after silica gel column chromatography (petroleum ether/EtOAc 6:4) as a sticky solid (179 mg, 64%). R_f = 0.42 (petroleum ether/EtOAc 5:5). IR (ATR) ν = 3489, 3315, 3304, 2918, 2849, 1627, 1648, 1465 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 6.9 Hz, CH₃), 1.23–1.36 (m, 28H, CH₂), 1.59–1.69 (m, 2H, COCH₂CH₂), 2.01 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.19 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.87 (s, 3H, CH₃O), 4.34 (d, 2H, J = 5.7 Hz, CH₂NH), 5.29–5.39 (m, 2H, CH=CH), 5.69 (s, 2H, OH, CH₂NH), 6.79 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101

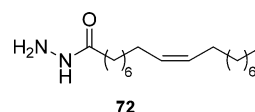
N'-(4'-Hydroxy-3'-methoxybenzylidene)hexadecano-hydrazide (67). General procedure X was applied to compound **66** (280 mg, 1.03 mmol), vanillin **1** (157 mg, 1.03 mmol), AcOH (60 μ L, 1.03 mmol) in MeOH (30 mL). Compound **67** was afforded as a white solid (242 mg, 58%) after recrystallization from hot MeOH. The ^1H NMR analysis confirmed the presence of the *cis* isomer of the imine as the minor product. Mp: 109–110 $^\circ\text{C}$. IR (ATR) ν = 3202, 3054, 2917, 2849, 1659, 1510 cm^{-1} . *Trans* isomer: ^1H NMR (400 MHz, CDCl_3) δ = 0.88 (t, 3H, J = 6.9 Hz, CH_3), 1.23–1.42 (m, 24H, CH_2), 1.69–1.78 (m, 2H, $\text{NHCOCH}_2\text{CH}_2$), 2.74 (t, 2H, J = 6.9 Hz, $\text{NHCOCH}_2\text{CH}_2$), 3.95 (s, 3H, CH_3O), 5.86 (s, 1H, OH), 6.93 (d, 1H, J = 8.2 Hz, H_{Ar}), 7.09 (dd, 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.25 (d, 1H, J = 1.8 Hz, H_{Ar}), 7.65 (s, 1H, $\text{HC}=\text{NNH}$), 9.02 (s, 1H, NHCO). *Cis* isomer: ^1H NMR (400 MHz, CDCl_3) δ = 2.28 (t, 2H, J = 6.9 Hz, $\text{NHCOCH}_2\text{CH}_2$), 3.94 (s, 1H, CH_3OH), 5.91 (br s, 1H, OH), 6.89 (d, 1H, J = 8.2 Hz, H_{Ar}), 6.98 (dd, 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.49 (br s, 1H, H_{Ar}), 8.00 (s, 1H, $\text{HC}=\text{NNH}$), 8.46 (s, 1H, NHCO). The rest of signals are common to *trans* isomer. *Trans* isomer: ^{13}C NMR (101 MHz, CDCl_3) δ = 14.27 (CH_3), 22.85 ($\text{NHCOCH}_2\text{CH}_2$), 24.97 (CH_2), 29.51 (CH_2), 29.59 (CH_2), 29.64 (CH_2), 29.72 (CH_2), 29.81 ($2 \times \text{CH}_2$), 29.85 ($4 \times \text{CH}_2$), 32.08 (CH_2), 32.96 ($\text{NHCOCH}_2\text{CH}_2$), 56.09 (CH_3O), 107.97 (C_{Ar}), 114.61 (C_{Ar}), 122.37 (C_{Ar}), 126.49 (C_{Ar}), 143.20 ($\text{HC}=\text{NNH}$), 147.07 (C_{Ar}), 147.90 (C_{Ar}), 176.00 (NHCO). *Cis* isomer: ^{13}C NMR (101 MHz, CDCl_3) δ = 56.38 (CH_3O), 107.86 (C_{Ar}), 114.13 (C_{Ar}), 123.80 (C_{Ar}), 126.20 (C_{Ar}). The rest of signals are common to *trans* isomer. HR-MS (ESI^+), m/z : [M + Na] $^+$ calcd for $\text{C}_{48}\text{H}_{80}\text{N}_4\text{O}_6\text{Na}$, 831.5976; found 831.5968.



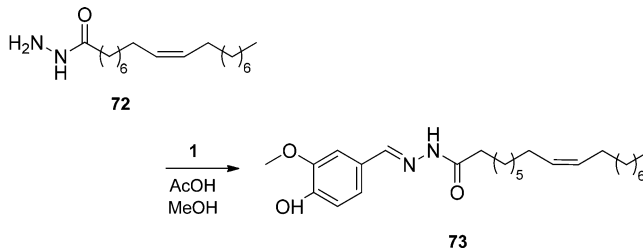
tert-Butyl Hydrazinecarboxylate (69). Hydrazine hydrate **68** (64%, 1.52 mL, 31.2 mmol) was mixed with isopropanol (3 mL) at 0 $^\circ\text{C}$. Then, a solution of Boc_2O (6.8 g, 31.2 mmol, 1 equiv) in isopropanol (6 mL) was added dropwise. The reaction mixture turned cloudy upon addition and was stirred at room temperature for 2 h. The solvent was removed under reduced pressure and the residue was dissolved in DCM, washed with 1 M HCl and brine. The organic phase was dried over Na_2SO_4 and the solvent was removed under reduced pressure. The residue was recrystallized from hexane to yield compound **69**²² as a white solid (1.94 g, 47%). Mp: 38–40 $^\circ\text{C}$. IR (ATR) ν = 3374, 3324, 2981, 1692, 1627, 1502 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 1.44 (s, 9H, $\text{C}(\text{CH}_3)_3$), 3.57 (s, 2H, NH_2), 6.00 (s, 1H, NHCO). ^{13}C NMR (101 MHz, CDCl_3) δ = 28.28 ($\text{C}(\text{CH}_3)_3$), 80.42 ($\text{C}(\text{CH}_3)_3$), 158.22 (COO).



N'-(tert-Butyloxycarbonyl)octadec-(9Z)-enohydrazide (70). General procedure I was applied to a solution of oleic acid **70** (1 g, 3.54 mmol) dissolved in DMF (30 mL), compound **69** (524 mg, 3.96 mmol), DIPEA (1.85 mL, 10.62 mmol), and HATU (2.02 g, 5.31 mmol). Compound **71**²³ was afforded after silica gel column chromatography (petroleum ether/EtOAc 7:3) as a yellow oil (1.32 g, 94%). R_f = 0.47 (petroleum ether/EtOAc 6:4). IR (ATR) ν = 3280, 2924, 2854, 1729, 1673, 1242 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.86 (t, 3H, J = 6.9 Hz, CH_3), 1.16–1.40 (m, 20H, CH_2), 1.44 (s, 9H, $\text{C}(\text{CH}_3)_3$), 1.57–1.74 (m, 2H, $\text{NHCOCH}_2\text{CH}_2$), 1.90–2.07 (m, 4H, CH_2CH , CHCH_2), 2.11–2.28 (m, 2H, $\text{NHCOCH}_2\text{CH}_2$), 5.22–5.43 (m, 2H, $\text{CH}=\text{CH}$), 6.85 (s, 1H, NHNH), 8.06 (s, 1H, NHNH). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.07 (CH_3), 22.64 (CH_2), 25.25 ($\text{NHCOCH}_2\text{CH}_2$), 27.14 (CH_2CH), 27.18 (CHCH_2), 28.11 ($\text{C}(\text{CH}_3)_3$), 29.08 (CH_2), 29.17 (CH_2), 29.19 (CH_2), 29.27 (CH_2), 29.29 (CH_2), 29.48 (CH_2), 29.67 (CH_2), 29.72 (CH_2), 31.86 (CH_2), 33.97 ($\text{NHCOCH}_2\text{CH}_2$), 81.66 ($\text{C}(\text{CH}_3)_3$), 129.68 ($\text{CH}=\text{CH}$), 129.93 ($\text{CH}=\text{CH}$), 155.85 ($\text{COC}(\text{CH}_3)_3$), 172.80 (NHCOCH₂).



Oleylhydrazine (72). To a solution of compound **71** (1 g, 2.52 mmol) in DCM (3 mL), TFA (1.93 mL, 25.2 mmol, 10 equiv) was added. The mixture was stirred for 2 h at room temperature. Then, the solvent was partially evaporated. Water was added, and the pH was adjusted to 7 with saturated solution of NaHCO_3 . The aqueous phase was extracted with DCM, and the organic solution was dried over Na_2SO_4 and filtered. The solvent was removed under reduced pressure to yield the compound **72** as a yellow solid (687 mg, 92%). Mp: 109–110 $^\circ\text{C}$. IR (ATR) ν = 3316, 3214, 2919, 2849, 1628, 1596 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.12–1.42 (m, 20H, CH_2), 1.53–1.74 (m, 2H, $\text{NHCOCH}_2\text{CH}_2$), 1.88–2.05 (m, 4H, CH_2CH , CHCH_2), 2.08–2.24 (m, 2H, $\text{NHCOCH}_2\text{CH}_2$), 3.97 (s, 2H, H_2N), 5.20–5.43 (m, 2H, $\text{CH}=\text{CH}$), 6.84 (s, 1H, NH). ^{13}C NMR (101 MHz, CDCl_3) δ = 14.08 (CH_3), 22.65 (CH_2), 25.46 ($\text{NHCOCH}_2\text{CH}_2$), 27.13 (CH_2CH), 27.19 (CHCH_2), 29.07 (CH_2), 29.18 (CH_2), 29.22 (CH_2), 29.29 ($2 \times \text{CH}_2$), 29.49 (CH_2), 29.66 (CH_2), 29.73 (CH_2), 31.87 (CH_2), 34.55 ($\text{NHCOCH}_2\text{CH}_2$), 129.67 ($\text{CH}=\text{CH}$), 129.99 ($\text{CH}=\text{CH}$), 173.98 (NHCOCH₂).



N'-(4'-Hydroxy-3'-methoxybenzylidene)octadec-(9Z)-enohydrazide (73). General procedure X was applied to compound **72** (300 mg, 1.01 mmol), vanillin **1** (153 mg, 1.01 mmol), AcOH (60 μ L, 1.01 mmol) in MeOH (30 mL). Compound **73** was afforded after silica gel column chromatography (petroleum ether/EtOAc 6:4) as a colorless oil (1.32 g, 94%). The ^1H NMR analysis confirmed the presence of the *cis* isomer of the imine as a minor product. IR (ATR) ν = 3452, 3194, 2921, 2852, 1650, 1211 cm^{-1} . *Trans* isomer: ^1H NMR (400 MHz, CDCl_3) δ = 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.22–1.43 (m, 20H, CH_2), 1.69–1.78 (m, 2H, $\text{NHCOCH}_2\text{CH}_2$), 1.94–2.07 (m, 4H, CH_2CH , CHCH_2), 2.74 (t, 2H, J = 6.9 Hz, $\text{NHCOCH}_2\text{CH}_2$), 3.95 (s, 3H, CH_3O), 5.31–5.36 (m, 2H, $\text{CH}=\text{CH}$), 5.93 (br s, 1H, OH), 6.93 (d, 1H, J = 8.2 Hz, H_{Ar}), 7.10 (dd, 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.25 (d, 1H, J = 1.8 Hz, H_{Ar}), 7.69 (s, 1H, $\text{HC}=\text{NNH}$), 9.43 (s, 1H, NHCO). *Cis* isomer: ^1H NMR (400 MHz, CDCl_3) δ = 2.28 (t, 2H, J = 6.9 Hz, $\text{NHCOCH}_2\text{CH}_2$), 3.93 (s, 1H, CH_3OH), 5.36–5.39 (m, 2H, $\text{CH}=\text{CH}$), 5.97 (br s, 1H, OH), 6.89 (d, 1H, J = 8.2 Hz, H_{Ar}), 6.97 (dd, 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.49 (d, 1H, J = 1.8 Hz, H_{Ar}), 8.00 (s, 1H, $\text{HC}=\text{NNH}$), 8.62 (s, 1H, NHCO). The rest of signals are common to *trans* isomer. *Trans* isomer: ^{13}C NMR (101 MHz, CDCl_3) δ = 14.26 (CH_3), 22.82 ($\text{NHCOCH}_2\text{CH}_2$), 25.00 (CH_2), 27.34 (CH_2CH), 27.36 (CHCH_2), 29.35 (CH_2), 29.46 (CH_2), 29.46 (CH_2), 29.49 (CH_2), 29.61 (CH_2), 29.66 (CH_2), 29.84 (CH_2), 29.91 (CH_2), 32.04 (CH_2), 32.94 ($\text{NHCOCH}_2\text{CH}_2$), 56.08 (CH_3O), 108.06 (C_{Ar}), 114.63 (C_{Ar}), 122.32 (C_{Ar}), 126.54 (C_{Ar}), 129.88 ($\text{CH}=\text{CH}$), 130.13 ($\text{CH}=\text{CH}$), 143.54 ($\text{HC}=\text{NNH}$), 147.06 (C_{Ar}), 147.89 (C_{Ar}), 176.30 (NHCO). *Cis* isomer: ^{13}C NMR (101 MHz, CDCl_3) δ = 56.35 (CH_3O), 107.87 (C_{Ar}), 114.11 (C_{Ar}), 123.79 (C_{Ar}), 126.16 (C_{Ar}), 147.24 (C_{Ar}), 147.73 (C_{Ar}). The rest of signals are common to *trans* isomer. HR-MS (ESI^+), m/z : [M + Na] $^+$ calcd for $\text{C}_{52}\text{H}_{84}\text{N}_4\text{O}_6\text{Na}$, 883.6289; found 883.6286.

5.3. TRP Channels Assays. Assays of TRP-mediated elevation of $[\text{Ca}^{2+}]_i$ were performed as previously described.⁶⁰ HEK-293 (human embryonic kidney) cells wild-type or stably overexpressing recombinant human TRPV1 or rat TRPV2 were grown on 100 mm diameter

1992 Petri dishes as monolayers in Eagle's minimum essential medium
1993 (EMEM) supplemented with 1% nonessential amino acids, 10% fetal
1994 bovine serum (FBS), 50 U/mL penicillin plus 50 $\mu\text{g/mL}$
1995 streptomycin, and 2 mM glutamine, maintained under 5% CO_2 at
1996 37 $^\circ\text{C}$ and only for the overexpressing cells selected by G-418
1997 (Geneticin, 600 mg mL^{-1} ; Thermo-Fisher Scientific). On the day of
1998 the experiment, the cells were loaded for 1 h at 25 $^\circ\text{C}$ with the Ca^{2+}
1999 indicator Fluo-4-AM (Thermo-Fisher Scientific) 4 μM in DMSO
2000 containing 0.02% Pluronic F-127 (Thermo-Fisher Scientific) in
2001 EMEM without FBS. After loading, cells were washed twice in
2002 Tyrode's buffer (145 mM NaCl, 2.5 mM KCl, 1.5 mM CaCl_2 , 1.2 mM
2003 MgCl_2 , 10 mM D-glucose, and 10 mM HEPES, pH 7.4), resuspended
2004 in the same buffer, and transferred, about 100 000 cells for each
2005 determination, to the quartz cuvette of the spectrofluorimeter (λ_{ex} =
2006 488 nm; λ_{em} = 516 nm) PerkinElmer LS50B equipped with PTP-1
2007 fluorescence Peltier system (PerkinElmer Life and Analytical Sciences,
2008 Waltham, MA, USA) under continuous stirring at 25 $^\circ\text{C}$. Experiments
2009 were carried by measuring cell fluorescence before and after the
2010 addition of test compounds at various concentrations. The values of
2011 the effect on $[\text{Ca}^{2+}]_i$ in wild-type (i.e., not transfected with any TRP
2012 construct) HEK-293 cells were taken as baselines. Potency (EC_{50}
2013 values) was determined as the concentration of test compounds
2014 exerting a half-maximal agonist effect (i.e., half-maximal increases in
2015 $[\text{Ca}^{2+}]_i$). The efficacy of the agonists was determined by comparing
2016 their effect to the maximal effect on $[\text{Ca}^{2+}]_i$ observed with 4 μM
2017 ionomycin. Antagonist/desensitizing behavior was evaluated against
2018 the agonist capsaicin 0.1 μM (Sigma-Aldrich) for TRPV1 and the
2019 agonists lysophosphatidylcholine (LPC) (Sigma-Aldrich) 3 μM and
2020 cannabidiol (CBD) 2 μM (a kind gift by GW Pharmaceuticals) for
2021 TRPV2 by adding the test compounds in the quartz cuvette 5 min
2022 before stimulation of cells with the agonist. The effect on $[\text{Ca}^{2+}]_i$
2023 exerted by agonist alone was taken as 100%. Data are expressed as the
2024 concentration exerting a half-maximal inhibition of agonist-induced
2025 $[\text{Ca}^{2+}]_i$ elevation (IC_{50}). Concentration–response curves were fitted
2026 by a sigmoidal regression with variable slope. Curve fitting and
2027 parameter estimation were performed with GraphPad Prism (Graph-
2028 Pad Software Inc., San Diego, CA). Determinations were performed
2029 at least in triplicate. Statistical analysis of the data was performed by
2030 analysis of variance at each point using ANOVA followed by
2031 Bonferroni's test.

2032 ■ ASSOCIATED CONTENT

2033 ■ Supporting Information

2034 The Supporting Information is available free of charge on the
2035 ACS Publications website at DOI: 10.1021/acs.jmed-
2036 chem.8b00734.

- 2037 Tables S1 and S2 of TRPV1 activity and ^1H and ^{13}C
2038 NMR spectra (PDF)
2039 Molecular formula strings and some data (CSV)

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2051 Notes

2052 The authors declare no competing financial interest.

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2065 ■ ABBREVIATIONS USED

TRPV2, transient receptor potential vanilloid 2; TRPV1,
transient receptor potential vanilloid 1; EA, ethanolamide;
LPC, lysophosphatidylcholine; CBD, cannabidiol; PEA, palmi-
toyl ethanolamide; POEA, palmitoleoyl ethanolamide; OEA,
oleoyl ethanolamide; LEA, linoleoyl ethanolamide; AEA,
arachidonoyl ethanolamide; EPEA, eicosapentaenoyl ethanola-
mide; DHEA, docosahexaenoyl ethanolamide; PA, palmita-
mide; SA, stearamide; OA, oleamide; LA, linoleamide; ErA,
erucamide

2075 ■ REFERENCES

- (1) Iwata, Y.; Katanosaka, Y.; Arai, Y.; Komamura, K.; Miyatake, K.;
Shigekawa, M. A Novel Mechanism of Myocyte Degeneration
Involving the Ca^{2+} -Permeable Growth Factor-Regulated Channel.
J. Cell Biol. **2003**, 161 (5), 957–967.
- (2) Iwata, Y.; Katanosaka, Y.; Arai, Y.; Shigekawa, M.; Wakabayashi,
S. Dominant-Negative Inhibition of Ca^{2+} Influx via TRPV2
Ameliorates Muscular Dystrophy in Animal Models. *Hum. Mol.*
Genet. **2009**, 18 (5), 824–834.
- (3) Zanou, N.; Iwata, Y.; Schakman, O.; Lebacqz, J.; Wakabayashi, S.;
Gailly, P. Essential Role of TRPV2 Ion Channel in the Sensitivity of
Dystrophic Muscle to Eccentric Contractions. *FEBS Lett.* **2009**, 583
(22), 3600–3604.
- (4) Aguetaz, E.; Bois, P.; Cognard, C.; Sebillé, S. Stretch-Activated
TRPV2 Channels: Role in Mediating Cardiopathies. *Prog. Biophys.*
Mol. Biol. **2017**, 130, 273–280.
- (5) Everaerts, W.; Gevaert, T.; Nilius, B.; De Ridder, D. On the
Origin of Bladder Sensing: Tr(i)Ps in Urology. *Neurourol. Urodyn.*
2008, 27 (4), 264–273.
- (6) Nilius, B.; Owsianik, G.; Voets, T.; Peters, J. A. Transient
Receptor Potential Cation Channels in Disease. *Physiol. Rev.* **2007**, 87
(1), 165–217.
- (7) Prevarskaya, N.; Zhang, L.; Barritt, G. TRP Channels in Cancer.
Biochim. Biophys. Acta, Mol. Basis Dis. **2007**, 1772 (8), 937–946.
- (8) Lehen'kyi, V.; Prevarskaya, N. Study of TRP Channels in Cancer
Cells. In *TRP Channels*; Zhu, M. X., Ed.; CRC Press/Taylor &
Francis: Boca Raton, FL, 2011; pp 375–394.
- (9) Santoni, G.; Farfariello, V. TRP Channels and Cancer: New
Targets for Diagnosis and Chemotherapy. *Endocr., Metab. Immune*
Disord.: Drug Targets **2011**, 11 (1), 54–67.
- (10) Caprodossi, S.; Lucciarini, R.; Amantini, C.; Nabissi, M.;
Canesin, G.; Ballardini, P.; Di Spilimbergo, A.; Cardarelli, M. A.; Servi,
L.; Mammana, G.; et al. Transient Receptor Potential Vanilloid Type
2 (TRPV2) Expression in Normal Urothelium and in Urothelial
Carcinoma of Human Bladder: Correlation with the Pathologic Stage.
Eur. Urol. **2008**, 54 (3), 612–620.
- (11) Monet, M.; Lehen'kyi, V.; Gackiere, F.; Firlej, V.;
Vandenbergh, M.; Roudbaraki, M.; Gkika, D.; Pourtier, A.; Bidaux,
G.; Slomianny, C.; et al. Role of Cationic Channel TRPV2 in
Promoting Prostate Cancer Migration and Progression to Androgen
Resistance. *Cancer Res.* **2010**, 70 (3), 1225–1235.

- (12) Gkika, D.; Prevarskaya, N. TRP Channels in Prostate Cancer: The Good, the Bad and the Ugly? *Asian J. Androl.* **2011**, *13* (5), 673–676.
- (13) Ziglioli, F.; Frattini, A.; Maestroni, U.; Dinale, F. Vanilloid-Mediated Apoptosis in Prostate Cancer Cells through ATRPV-1 Dependent and a TRPV-1-Independent Mechanism. *Acta Bio Med. Atenei Parmensis* **2009**, *80* (1), 13–20.
- (14) Wang, C.; Hu, H.-Z.; Colton, C. K.; Wood, J. D.; Zhu, M. X. An Alternative Splicing Product of the Murine Trpv1 Gene Dominant Negatively Modulates the Activity of TRPV1 Channels. *J. Biol. Chem.* **2004**, *279* (36), 37423–37430.
- (15) Neeper, M. P.; Liu, Y.; Hutchinson, T. L.; Wang, Y.; Flores, C. M.; Qin, N. Activation Properties of Heterologously Expressed Mammalian TRPV2. *J. Biol. Chem.* **2007**, *282* (21), 15894–15902.
- (16) Juvin, V.; Penna, A.; Chemin, J.; Lin, Y.-L.; Rassendren, F.-A. Pharmacological Characterization and Molecular Determinants of the Activation of Transient Receptor Potential V2 Channel Orthologs by 2-Aminoethoxydiphenyl Borate. *Mol. Pharmacol.* **2007**, *72* (5), 1258–1268.
- (17) Qin, N.; Neeper, M. P.; Liu, Y.; Hutchinson, T. L.; Lubin, M. L.; Flores, C. M. TRPV2 Is Activated by Cannabidiol and Mediates CGRP Release in Cultured Rat Dorsal Root Ganglion Neurons. *J. Neurosci.* **2008**, *28* (24), 6231–6238.
- (18) De Petrocellis, L.; Ligresti, A.; Moriello, A. S.; Allarà, M.; Bisogno, T.; Petrosino, S.; Stott, C. G.; Di Marzo, V. Effects of Cannabinoids and Cannabinoid-Enriched Cannabis Extracts on TRP Channels and Endocannabinoid Metabolic Enzymes. *Br. J. Pharmacol.* **2011**, *163* (7), 1479–1494.
- (19) Bang, S.; Kim, K. Y.; Yoo, S.; Lee, S.-H.; Hwang, S. W. Transient Receptor Potential V2 Expressed in Sensory Neurons Is Activated by Probenecid. *Neurosci. Lett.* **2007**, *425* (2), 120–125.
- (20) Rohacs, T. Phosphoinositide Regulation of TRP Channels. In *Mammalian Transient Receptor Potential (TRP) Cation Channels*; Springer: Cham, Switzerland, 2014; pp 1143–1176.
- (21) Monet, M.; Gkika, D.; Lehen'kyi, V.; Pourtier, A.; Vanden Abeele, F.; Bidaux, G.; Juvin, V.; Rassendren, F.; Humez, S.; Prevarskaya, N. Lysophospholipids Stimulate Prostate Cancer Cell Migration via TRPV2 Channel Activation. *Biochim. Biophys. Acta, Mol. Cell Res.* **2009**, *1793* (3), 528–539.
- (22) De Petrocellis, L.; Ligresti, A.; Moriello, A. S.; Allarà, M.; Bisogno, T.; Petrosino, S.; Stott, C. G.; Di Marzo, V. Effects of Cannabinoids and Cannabinoid-Enriched Cannabis Extracts on TRP Channels and Endocannabinoid Metabolic Enzymes. *Br. J. Pharmacol.* **2011**, *163* (7), 1479–1494.
- (23) Peralvarez-Marín, A.; Donate-Macian, P.; Gaudet, R. What Do We Know about the Transient Receptor Potential Vanilloid 2 (TRPV2) Ion Channel? *FEBS J.* **2013**, *280* (21), 5471–5487.
- (24) Caterina, M. J.; Rosen, T. A.; Tominaga, M.; Brake, A. J.; Julius, D. A Capsaicin-Receptor Homologue with a High Threshold for Noxious Heat. *Nature* **1999**, *398*, 436.
- (25) Alexander, S.; Mathie, A.; Peters, J. Guide to Receptors and Channels (GRAC). *Br. J. Pharmacol.* **2011**, *164* (s1), S1–S2.
- (26) Leffler, A.; Linte, R. M.; Nau, C.; Reeh, P.; Babes, A. A High-Threshold Heat-Activated Channel in Cultured Rat Dorsal Root Ganglion Neurons Resembles TRPV2 and Is Blocked by Gadolinium. *Eur. J. Neurosci.* **2007**, *26* (1), 12–22.
- (27) Stotz, S. C.; Vriens, J.; Martyn, D.; Clardy, J.; Clapham, D. E. Citral Sensing by TRANSient Receptor Potential Channels in Dorsal Root Ganglion Neurons. *PLoS One* **2008**, *3* (5), e2082.
- (28) Nie, L.; Oishi, Y.; Doi, I.; Shibata, H.; Kojima, I. Inhibition of Proliferation of MCF-7 Breast Cancer Cells by a Blocker of Ca²⁺-Permeable Channel. *Cell Calcium* **1997**, *22* (2), 75–82.
- (29) Hisanaga, E.; Nagasawa, M.; Ueki, K.; Kulkarni, R. N.; Mori, M.; Kojima, I. Regulation of Calcium-Permeable TRPV2 Channel by Insulin in Pancreatic β -Cells. *Diabetes* **2009**, *58* (1), 174–184.
- (30) Aoyagi, K.; Ohara-Imaizumi, M.; Nishiwaki, C.; Nakamichi, Y.; Nagamatsu, S. Insulin/Phosphoinositide 3-Kinase Pathway Accelerates the Glucose-Induced First-Phase Insulin Secretion through TRPV2 Recruitment in Pancreatic β -Cells. *Biochem. J.* **2010**, *432* (2), 375–386.
- (31) Mihara, H.; Boudaka, A.; Shibasaki, K.; Yamanaka, A.; Sugiyama, T.; Tominaga, M. Involvement of TRPV2 Activation in Intestinal Movement through Nitric Oxide Production in Mice. *J. Neurosci.* **2010**, *30* (49), 16536–16544.
- (32) Lin, Y.; Sun, Z. Antiaging Gene Klotho Enhances Glucose-Induced Insulin Secretion by Up-Regulating Plasma Membrane Levels of TRPV2 in MIN6 β -Cells. *Endocrinology* **2012**, *153* (7), 3029–3039.
- (33) Iwata, Y.; Ohtake, H.; Suzuki, O.; Matsuda, J.; Komamura, K.; Wakabayashi, S. Blockade of Sarcolemmal TRPV2 Accumulation Inhibits Progression of Dilated Cardiomyopathy. *Cardiovasc. Res.* **2013**, *99* (4), 760–768.
- (34) Zanou, N.; Mondin, L.; Fuster, C.; Seghers, F.; Dufour, I.; de Clippele, M.; Schakman, O.; Tajeddine, N.; Iwata, Y.; Wakabayashi, S.; Voets, T.; Allard, B.; Gailly, P. Osmosensation in TRPV2 Dominant Negative Expressing Skeletal Muscle Fibres. *J. Physiol.* **2015**, *593* (17), 3849–3863.
- (35) Zubcevic, L.; Herzik, M. A., Jr; Chung, B. C.; Liu, Z.; Lander, G. C.; Lee, S.-Y. Cryo-Electron Microscopy Structure of the TRPV2 Ion Channel. *Nat. Struct. Mol. Biol.* **2016**, *23*, 180.
- (36) Huynh, K. W.; Cohen, M. R.; Jiang, J.; Samanta, A.; Lodowski, D. T.; Zhou, Z. H.; Moiseenkova-Bell, V. Y. Structure of the Full-Length TRPV2 Channel by Cryo-EM. *Nat. Commun.* **2016**, *7*, 11130.
- (37) Yang, F.; Vu, S.; Yarov-Yarovoy, V.; Zheng, J. Rational Design and Validation of a Vanilloid-Sensitive TRPV2 Ion Channel. *Proc. Natl. Acad. Sci. U. S. A.* **2016**, *113* (26), E3657–E3666.
- (38) Miao, X.; Malacea, R.; Fischmeister, C.; Bruneau, C.; Dixneuf, P. H. Ruthenium-Alkylidene Catalysed Cross-Metathesis of Fatty Acid Derivatives with Acrylonitrile and Methyl Acrylate: A Key Step toward Long-Chain Bifunctional and Amino Acid Compounds. *Green Chem.* **2011**, *13* (10), 2911–2919.
- (39) Malet, R.; Schäfer, H. J.; Quermann, R. Conversion of Unsaturated Fatty Acids — Nucleophilic Additions to Methyl (E)-12-Oxo-10-Octadecenoate [1]. *Lipid/Fett* **1996**, *98* (11), 370–379.
- (40) Stewart, I. C.; Bergman, R. G.; Toste, F. D. Phosphine-Catalyzed Hydration and Hydroalkoxylation of Activated Olefins: Use of a Strong Nucleophile to Generate a Strong Base. *J. Am. Chem. Soc.* **2003**, *125* (29), 8696–8697.
- (41) Straathof, A. J. J.; Adlercreutz, P. *Applied Biocatalysis*; CRC Press: London, 2000.
- (42) Carpino, L. A.; El-Faham, A.; Albericio, F. Racemization Studies during Solid-Phase Peptide Synthesis Using Azabenzotriazole-Based Coupling Reagents. *Tetrahedron Lett.* **1994**, *35* (15), 2279–2282.
- (43) Thalmann, A.; Oertle, K.; Gerlach, H. Ricinelaic Acid Lactone. In *Organic Syntheses*; Major Reference Works; John Wiley & Sons, Inc.: Hoboken, NJ, U.S., 2003; pp 192–192.
- (44) Collins, J. C.; Hess, W. W.; Frank, F. J. Dipyrindine-Chromium(VI) Oxide Oxidation of Alcohols in Dichloromethane. *Tetrahedron Lett.* **1968**, *9* (30), 3363–3366.
- (45) Müller, J.; Terfort, A. Synthesis of Pure Aromatic, Aliphatic, and Araliphatic Diselenides. *Inorg. Chim. Acta* **2006**, *359* (15), 4821–4827.
- (46) Salunke, D. B.; Connelly, S. W.; Shukla, N. M.; Hermanson, A. R.; Fox, L. M.; David, S. A. Design and Development of Stable, Water-Soluble, Human Toll-like Receptor 2 Specific Monoacyl Lipopeptides as Candidate Vaccine Adjuvants. *J. Med. Chem.* **2013**, *56* (14), 5885–5900.
- (47) Pedzisa, L.; Li, X.; Rader, C.; Roush, W. R. Assessment of Reagents for Selenocysteine Conjugation and the Stability of Selenocysteine Adducts. *Org. Biomol. Chem.* **2016**, *14* (22), 5141–5147.
- (48) López, S. E.; Salazar, J. Trifluoroacetic Acid: Uses and Recent Applications in Organic Synthesis. *J. Fluorine Chem.* **2013**, *156*, 73–100.
- (49) Agnihotri, G.; Crall, B. M.; Lewis, T. C.; Day, T. P.; Balakrishna, R.; Warshakoon, H. J.; Malladi, S. S.; David, S. A.

- 2253 Structure–Activity Relationships in Toll-Like Receptor 2-Agonists
2254 Leading to Simplified Monoacyl Lipopeptides. *J. Med. Chem.* **2011**, 54
2255 (23), 8148–8160.
- 2256 (50) Kumar, V.; Sharma, A.; Chander Sharma, P. Synthesis of Some
2257 Novel 2,5-Disubstituted Thiazolidinones from a Long Chain Fatty
2258 Acid as Possible Anti-Inflammatory, Analgesic and Hydrogen
2259 Peroxide Scavenging Agents. *J. Enzyme Inhib. Med. Chem.* **2011**, 26
2260 (2), 198–203.
- 2261 (51) Yang, F.; Zheng, J. Understand Spiciness: Mechanism of
2262 TRPV1 Channel Activation by Capsaicin. *Protein Cell* **2017**, 8 (3),
2263 169–177.
- 2264 (52) Gao, M.; Li, J.; Nie, C.; Song, B.; Yan, L.; Qian, H. Design,
2265 Synthesis and Biological Evaluation of Novel Hydrogen Sulfide
2266 Releasing Capsaicin Derivatives. *Bioorg. Med. Chem.* **2018**, 26 (9),
2267 2632–2639.
- 2268 (53) Walpole, C. S.; Wigglesworth, R.; Bevan, S.; Campbell, E. A.;
2269 Dray, A.; James, I. F.; Masdin, K. J.; Perkins, M. N.; Winter, J.
2270 Analogues of Capsaicin with Agonist Activity as Novel Analgesic
2271 Agents; Structure-Activity Studies. 2. The Amide Bond “B-Region”. *J.*
2272 *Med. Chem.* **1993**, 36 (16), 2373–2380.
- 2273 (54) Janusz, J. M.; Buckwalter, B. L.; Young, P. A.; LaHann, T. R.;
2274 Farmer, R. W.; Kasting, G. B.; Loomans, M. E.; Kerckaert, G. A.;
2275 Maddin, C. S.; Berman, E. F. Vanilloids. 1. Analogs of Capsaicin with
2276 Antinociceptive and Antiinflammatory Activity. *J. Med. Chem.* **1993**,
2277 36 (18), 2595–2604.
- 2278 (55) Lee, J.; Kang, S.-U.; Choi, H.-K.; Lee, J.; Lim, J.-O.; Kil, M.-J.;
2279 Jin, M.-K.; Kim, K.-P.; Sung, J.-H.; Chung, S.-J.; et al. Analysis of
2280 Structure–activity Relationships for the ‘B-Region’ of N -(3-Acyloxy-
2281 2-Benzylpropyl)- N ’ -[4-(Methylsulfonylamino)Benzyl]Thiourea
2282 Analogues as Vanilloid Receptor Antagonists: Discovery of an N
2283 -Hydroxythiourea Analogue with Potent Analgesic A. *Bioorg. Med.*
2284 *Chem. Lett.* **2004**, 14 (9), 2291–2297.
- 2285 (56) Cao, E.; Liao, M.; Cheng, Y.; Julius, D. TRPV1 Structures in
2286 Distinct Conformations Reveal Activation Mechanisms. *Nature* **2013**,
2287 504 (7478), 113–118.
- 2288 (57) Gavva, N. R.; Klionsky, L.; Qu, Y.; Shi, L.; Tamir, R.; Edenson,
2289 S.; Zhang, T. J.; Viswanadhan, V. N.; Toth, A.; Pearce, L. V.; et al.
2290 Molecular Determinants of Vanilloid Sensitivity in TRPV1. *J. Biol.*
2291 *Chem.* **2004**, 279 (19), 20283–20295.
- 2292 (58) Shibasaki, K. Physiological Significance of TRPV2 as a
2293 Mechanosensor, Thermosensor and Lipid Sensor. *J. Physiol. Sci.*
2294 **2016**, 66 (5), 359–365.
- 2295 (59) Appendino, G.; Minassi, A.; Morello, A. S.; De Petrocellis, L.;
2296 Di Marzo, V. N -Acylvanillamides: Development of an Expeditious
2297 Synthesis and Discovery of New Acyl Templates for Powerful
2298 Activation of the Vanilloid Receptor. *J. Med. Chem.* **2002**, 45 (17),
2299 3739–3745.
- 2300 (60) Moriello, A. S.; De Petrocellis, L. Assay of TRPV1 Receptor
2301 Signaling. In *Endocannabinoid Signaling: Methods and Protocols*; Press,
2302 H., Ed.; Springer: New York, NY, 2016; pp 65–76.