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Document downloaded from:

<http://hdl.handle.net/10459.1/72213>

The final publication is available at:

<https://doi.org/10.1017/S0007485312000193>

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5 **Cuticular hydrocarbons discriminate cryptic *Macrolophus***
6 **species (Hemiptera: Miridae)**

7

8 **César Gemeno^{1*}, Nerea Laserna¹, Magí Riba², Joan Valls³, Cristina**
9 **Castañé⁴ and Oscar Alomar⁴**

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12 ¹ *Department of Crop and Forest Science, University of Lleida, 25198 Lleida, Spain*

13 ² *Department of Chemistry, University of Lleida, 25198 Lleida, Spain*

14 ³ *Biostatistics Unit. Biomedical Research Institute (IRBLLEIDA), 25198 Lleida, Spain*

15 ⁴ *IRTA Cabrils, 08348 Barcelona, Spain*

16

17

18 *Corresponding author

19 Fax: +34 973-702690

20 Telephone: +34 973-702531

21 e-mail: cesar.gemeno@pvcf.udl.cat

22

23 *Macrolophus pygmaeus* is commercially employed in the biological control of
24 greenhouse and field vegetable pests. It is morphologically undistinguishable from the
25 cryptic species *M. melanotoma*, and this interferes with the evaluation of the biological
26 control activity of *M. pygmaeus*. We analysed the potential of cuticular hydrocarbon
27 composition as a method to discriminate the two *Macrolophus* species. A third species,
28 *M. costalis*, which is morphologically different from the other two species by having a
29 dark spot at the tip of the scutellum, served as a control. Sex, diet and species, all had
30 significant effects in the cuticular hydrocarbon profiles, but the variability associated to
31 sex or diet was smaller than among species. Discriminant quadratic analysis of cuticular
32 hydrocarbons confirmed the results of previous molecular genetic studies and showed,
33 using cross-validation methods, that *M. pygmaeus* can be discriminated from *M. costalis*
34 and *M. melanotoma* with prediction errors of 6.75% and 0%, respectively. Therefore,
35 cuticular hydrocarbons can be used to separate *M. pygmaeus* and *M. melanotoma*, while
36 *M. costalis* and *M. pygmaeus* can be readily distinguished from each other by the dark
37 tip of the scutellum of the former one.

38

39 **Keywords:** cuticular hydrocarbons, biological control, taxonomy

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41

42

Introduction

43

44 Several species in the genus *Macrolophus* (Hemiptera: Miridae) are efficient predators
45 of vegetable crops pests (e.g. whiteflies, aphids and thrips) (Alvarado et al., 1997;
46 Riudavets & Castañé 1998; Montserrat et al., 2000; Alomar et al., 2006; Hansen et al.,
47 1999; Athanassiou et al., 2003; Margaritopoulos et al., 2003). Control strategies that
48 incorporate *Macrolophus* species are based on both seasonal inoculative releases of
49 commercially produced individuals and habitat management to conserve natural
50 populations and enhance colonization (Alomar et al., 2002). In order to avoid the
51 incorrect use of predator reservoir plants it is essential to correctly identify the predator
52 species.

53

54 Several *Macrolophus* species are morphologically similar or have highly variable
55 taxonomic characters, which hampers correct identification and questions species-
56 identity status (Josifov, 1992; Kerzhner & Josifov, 1999). This is particularly true for
57 the two species that have been most cited as biological control agents, *M. pygmaeus*
58 (Rambur 1839), and *M. melanotoma* (Costa 1853) (the last one mostly cited as its junior
59 synonym *M. caliginosus* Wagner 1951), and has probably resulted in incorrect
60 attribution of biological control by *M. pygmaeus* to *M. melanotoma* (see Martínez-
61 Casales et al., 2006 for a historical review). DNA methods have been employed recently
62 to distinguish *M. pygmaeus* from *M. melanotoma* (Perdikis et al., 2003; Martínez-
63 Casales et al., 2006). Cuticular hydrocarbon analysis could provide an additional
64 method to separate morphologically similar species (Bagnères & Wicker-Thomas,
65 2010).

66

67 The insect cuticle is coated with a thin lipid layer containing a high percentage of
68 hydrocarbons (linear, branched, saturated and unsaturated), one of which main functions
69 is to prevent desiccation and pathogen entrance into the body (Howard & Blomquist,
70 2005). Cuticular hydrocarbons are usually present in high amounts and are easy to
71 extract by quickly rinsing the specimen in non-polar organic solvents (Blomquist et al.,
72 1987). In addition, most cuticular hydrocarbons are chemically stable and not very
73 volatile (Martin et al., 2009). The hydrocarbon blend is often species specific and
74 therefore it is a useful character in insect taxonomy (reviewed by Bagnères & Wicker-
75 Thomas, 2010).

76

77 In contrast to other insect groups, there are relatively few studies of cuticular
78 hydrocarbon taxonomy in Hemiptera. A recent publication reviews the hydrocarbons of
79 blood-sucking bugs (Juárez & Fernández, 2007), and isolated studies compare cuticular
80 hydrocarbons of *Orius* species (Anthocoridae) (Nakabou & Ohno, 2001), and aphids
81 (Clements et al., 2000). With regard to mirids, analysis of cuticular hydrocarbons has
82 been reported only for *Lygocoris pabulinus* (L.) (Drijfhout & Groot, 2001).

83

84 The purpose of the present study is to identify the cuticular hydrocarbons of three
85 *Macrolophus* species (*M. melanotoma*, *M. pygmaeus*, and *M. costalis*), as they may be
86 important in species recognition and mating behaviour. Since diet and sex may affect
87 insect cuticular hydrocarbon composition (Liang & Silverman, 2000; Thomas &
88 Simmons, 2008; Guerrieri et al., 2009), we also analysed the effect of diet (i.e., host
89 plant and associated prey) and sex on cuticular hydrocarbons. We then determine if
90 cuticular hydrocarbons can be used to distinguish adults of two *Macrolophus* species

91 (*M. melanotoma* and *M. pygmaeus*) that cannot be distinguished easily by external
92 morphological characters alone. A third predatory species, *M. costalis* Fieber, was
93 included in the study to confirm previous results on the taxonomic relationship of the
94 three species (Martínez-Cascales et al., 2006).

95

96 **Materials and methods**

97

98 *Insects*

99

100 Chemical analyses were performed on adults of both sexes. Wild individuals were
101 collected in the proximities of Cabrils (Barcelona, Spain) in the spring of 2008.
102 *Macrolophus melanotoma* was collected from *Dittrichia viscosa* (L.) Greuter
103 (Compositae), where it is abundant (O. A., personal observation). *Macrolophus*
104 *costalis*, which is readily distinguished by the dark spot on the scutellum, was collected
105 on *Cistus albidus* L. (Cistaceae), where it is commonly found (O.A., personal
106 observation). *M. pygmaeus* was obtained from a > 5-years-old laboratory colony
107 originated from tomato fields, and maintained on tobacco plants and frozen *Ephestia*
108 *kuehniella* Zeller (Lepidoptera: Pyralidae) egg prey. This colony is refreshed annually
109 with individuals collected in local tomato fields (Cabrils, Barcelona, Spain).
110 Examination of the length and shape of the respiratory horns of eggs (Perdikis et al.,
111 2003; Alomar & Goula, unpub. results) was used to confirm the identity of collected
112 females of *M. melanotoma* and *M. pygmaeus*. Some of those individuals were frozen (-
113 20°C) immediately or within 7 days after being collected (in which case they were
114 maintained in their respective diet), and are referred to as the “field” individuals.
115 Additionally, in order to evaluate the effect of diet on the cuticular hydrocarbon profile,
116 4th-instar *M. pygmaeus* nymphs taken from the colony, and the progeny of field-
117 collected *M. melanotoma* and *M. costalis*, were reared individually on tobacco leaves
118 and *E. kuehniella* eggs. The resulting adults, referred to as the “laboratory” individuals,
119 were frozen when 3 to 6 days old and analyzed at the same time as the “field”
120 individuals described above.

121
122 Cages used for oviposition and nymphal development (7-cm diameter x 3.5-cm high
123 with a ventilated lid) had a layer of 0.5% agar and a tobacco leaf disc placed on top of
124 it, with the abaxial surface facing upwards, and were kept at $25 \pm 2^\circ\text{C}$ and $85 \pm 5\%$ RH,
125 under a 16:8 light:dark photoregime. Individuals were moved to new cages as needed
126 and were provided with frozen *E. kuehniella* egg prey twice a week.

127

128 *Chemical Analyses*

129

130 Individuals were taken out of the freezer one at a time, let defrost, and submerged in 20
131 μl of hexane (HPLC grade, Sigma-Aldrich, Madrid, Spain) containing 50 ng of
132 pentadecane (98% pure, Sigma-Aldrich, Madrid, Spain) in a conical-bottom hexane-
133 rinsed glass vial. The use of internal standard allows minimization of any differences in
134 injection volume and in the daily response of the equipment. Previous analyses
135 indicated that pentadecane is not present in significant amounts in cuticular extracts of
136 these species. Five minutes after immersion in the solvent the insect was removed and
137 the solution was either analysed immediately or returned to the freezer (-20°C) for later
138 analysis.

139

140 For chemical analyses the volume of the extract was reduced immediately before
141 injection to 2-4 μl with a gentle nitrogen stream and 1 μl was injected manually in a gas
142 chromatograph (GC) in the splitless mode (split valve opened after 1 min). Samples
143 were analysed in either a GC-FID or a GC-MSD (Agilent Technologies 6890N GC, and
144 5973 Network quadrupole MSD), each equipped with a DB-5 column (30 m x of 0.32
145 mm ID x 0.25 μm , Agilent Technologies), and run with the same temperature program:

146 start at 60°C for 1 min, then increase to 320°C at 10 °C/min, and maintain at this
147 temperature for 25 min. Carrier gas was Helium at constant flow (1 ml/min), the
148 injector was set a 250°C and the detector at 280°C.

149

150 A total of 119 insects were analysed by GC-FID, and these are the samples used in the
151 statistical comparisons [10 individuals of each species, sex and diet (laboratory versus
152 field), except for N=9 field *M. melanotoma* males]. Two additional insects of each sex
153 and species were analysed by GC-MSD, and these data were used in the chemical
154 identification of the compounds. Straight-chain alkanes were identified by comparison
155 of retention times and mass spectra of authentic standards (Fluka, alkane standard
156 solutions 04070 (C₈-C₂₀) and 04071 (C₂₁-C₄₀), Sigma-Aldrich, Madrid, Spain).
157 Retention indices (RIs) were calculated according to van den Dool & Kratz (1963).
158 Linear and methyl branched alkanes were tentatively identified by comparison of their
159 RI and the characteristic ion fragments with those reported in the literature (Blomquist
160 et al., 1987; Juárez & Blomquist, 1993; Carlson et al., 1998; Juárez et al., 2001; Mullen
161 et al., 2008; Gomes et al., 2008; Dall'Aglio-Holvorcem et al., 2009).

162

163 Chain length of the selected compounds ranged between C₂₄ and more than C₄₀. Since
164 our largest alkane standard was C₄₀ we fitted a curve to the straight chain alkane
165 standards C₂₇ to C₄₀ to extrapolate the retention times of C₄₁ and C₄₂ ($R^2 = 0.99$, $y =$
166 $125.161 - 0.066 * x + 1.105 * 10^{-5} * x^2$, where y = estimated RT and x = chain length), and
167 then estimated the RI of sample peaks 27 and 28 (table 1).

168

169

Statistical Analyses

170

171 Percent relative abundance [(area of the target peak/area of the internal standard peak) x
172 100] was the parameter used in all the group comparisons. We chose those compounds
173 that showed a high relative abundance combined with a moderate coefficient of
174 variation, relative to other compounds, and that occurred consistently in at least one
175 class (i.e., species, sex, or diet). This resulted in the selection of 28 peaks, some of
176 which were abundant in one sex or species but below threshold level in others. A zero
177 value was assigned to undetected compounds. A Kolmogorov-Smirnov test was used to
178 assess normality of the data.

179

180 Two statistical analyses were performed. The first analysis was carried out to determine
181 the effect of diet, sex and species on the cuticular hydrocarbons. First we performed a
182 multivariate analysis of variance (MANOVA) in order to study how the 28 cuticular
183 hydrocarbons altogether vary with species, sex and diet. To measure the variability that
184 could be explained by each of these factors we computed the Pillai-Bartlett statistic
185 (Hand and Taylor, 1987), which provides a measure of the ratio of effect variance to
186 error variance. To assess the effect of individual cuticular hydrocarbons, the three
187 species were compared, compound by compound, males and females separately, using
188 ANOVA followed by a *Tukey's* mean comparison test. In addition, the effect of sex and
189 diet on each cuticular hydrocarbon compound was analysed with *t-tests*. *Bonferroni*
190 method was used to correct for multiple testing so that the threshold of significance was
191 set at $0.05/28 = 0.0017$ in the ANOVA and *t-tests*.

192

193 The second statistical analysis was performed to build a model to predict species
194 identity. We performed *Principal component analysis* (PCA), using the correlation
195 matrix of untransformed data, to obtain uncorrelated scores, visualize aggregation

196 patterns of the different insect groups, and to determine which compounds had the
197 strongest effect in this pattern. To build a prediction model for the species, the main
198 principal components (those adding at least 78% total variability) were used in a
199 *Quadratic discriminant analysis* (QDA), and the prediction error of discriminant
200 functions was evaluated using a 10-fold cross-validation method. The statistical tests
201 were performed with R software (R Development Core Team, 2008). Analysis codes of
202 the prediction model are available as Supplementary Material.

203

204

205 **Results**

206

207 Representative chromatograms indicating the 28 diagnostic peaks (by order of elution)
208 are shown in fig. 1. Normality assumption was accepted for all samples ($P < 0.0017$).
209 Significant differences in cuticular hydrocarbon profiles for diet, sex and species were
210 observed (MANOVA, $P < 0.001$, table 1). The differences between sexes or diets were
211 much smaller than those between species. Using the approximated F-statistic we found
212 that variability among species is almost 4 times higher than between males and females
213 ($26.8/6.9 = 3.87$) and almost 3 times higher than between diets ($26.8/9.1 = 2.91$).

214

215 Statistical comparison of individual compounds among the three species confirmed the
216 pattern shown in fig. 1: a large proportion of compounds were similar between *M.*
217 *pygmaeus* and *M. costalis*, and different from *M. melanotoma* (fig. 2, ANOVA, $P <$
218 0.0017). Roughly the first half (compounds 2 to 15) were more abundant in *M.*
219 *melanotoma* than in the other two species, and the second half (compounds 16 to 28)
220 were less abundant in *M. melanotoma* than in the other species. Therefore *M.*
221 *melanotoma* is different from the other two species by having a larger quantity of the
222 early eluting compounds, and a smaller quantity of the late eluting compounds (fig. 2).
223 Sex differences in cuticular hydrocarbons were numerous in *M. melanotoma*, scarce in
224 *M. pygmaeus* and lacking in *M. costalis* (fig. 2, *t-test*, $P < 0.0017$). The cuticular
225 hydrocarbon blends of *M. melanotoma* males and females were clearly different (figs. 1
226 and 2). Individuals reared in the laboratory in isolation with a homogeneous diet of
227 tobacco and moth eggs had a higher quantity of some hydrocarbons than their “field”
228 relatives (fig. 3, *t-test*, $P < 0.0017$), but the species-specific cuticular hydrocarbon
229 pattern was not altered (fig. 3), thus indicating a weak effect by diet.

230

231 Linear alkanes, from C₂₁ to C₃₅, were identified in the cuticular samples of the 3
232 species, but in very small quantities, and only one of them (C₂₄ or tetracosane,
233 compound 1, table 2) was abundant enough to be included among the 28 selected
234 compounds. The rest of the hydrocarbons consisted of unbranched monoenes and
235 alkanes with one or more methyl branches (table 2). Ion fragmentation suggests that
236 some peaks may contain more than one compound. A characteristic compound of *M.*
237 *melanotoma* is 2-methyldotriacontane (compound 7). The largest difference of *M.*
238 *melanotoma* females with the other two species, besides compound 7, was a larger
239 quantity of compounds 12 and 13 (11, 15-dimethylpentatriacontane and 11, 15, 19-
240 trimethyl pentatriacontane). For *M. melanotoma* males the largest difference with the
241 other two species was a larger quantity of early eluting compounds 2 to 5. *M.*
242 *melanotoma* males and females differed in 25 of the 28 hydrocarbons: the first 6, by
243 order of elution, were more abundant in males, and the other 19 were more abundant in
244 females (fig. 2). To name some examples, males had more hentriacontene and
245 triacontene (compounds 3 and 6) than females, and females had more 11, 15-
246 dimethylpentatriacontane and 11, 15, 19-trimethylheptatriacontane (compounds 12 and
247 20) than males. Compound 7 (2-methyldotriacontane), which is characteristic of this
248 species, was not different between sexes.

249

250 PC1 and PC2 explained 46.28 % and 21.73 % of the variability, respectively, and
251 clearly segregated *M. melanotoma* from the other two species, whereas sex differences
252 were only apparent in *M. melanotoma* (fig. 4). Segregation between field and laboratory
253 individuals was weak and only apparent in *M. melanotoma* (fig. 4), thus indicating that
254 diet did not affect the species-specific cuticular hydrocarbon blend. Different

255 compounds contributed in varying degrees to the PCs (table 2). PC1, which separated
256 *M. melanotoma* males from the other two species and from the females (fig. 4), was
257 mainly influenced by compounds that were characteristically small in *M. melanotoma*
258 males (compounds 17, 20, 22, 23 and 25, fig. 2). PC2, which separated *M. melanotoma*
259 females from males and from the other two species, was strongly influenced by
260 compounds 11 to 15 (table 2), which were more abundant in *M. melanotoma* females
261 than in the other insects (fig. 2).

262

263 With regard to the prediction model, a QDA based on PC1, PC2 and PC3 (which
264 together explained 78% of total variability) was used to predict the species of each test
265 individual (see Supplementary Material files). QDA was able to correctly classify all *M.*
266 *melatonoma* individuals, but 5 *M. costalis* individuals were classified as *M. pygmaeus*
267 and 2 *M. pygmaeus* were classified as *M. costalis* (table 3). Therefore the global error
268 was 5.88% when comparing these three species. To further assess the error prediction
269 level, we first run a training and test method using 3 quarters of the sample (90
270 individuals chosen at random) to build a QDA, and the remaining quarter of the sample
271 (29 individuals chosen at random) to test the prediction accuracy, which resulted in a
272 3.5% global error. Next, we used a 10-fold cross-validation method which indicated a
273 global prediction error of 6.75% (ranging from 0% to 25%), attributed to the separation
274 between *M. pygmaeus* and *M. costalis*, given that the error rate when discriminating *M.*
275 *melatonoma* from *M. pygmaeus* or *M. costalis* was 0% in all cases. We also performed
276 QDA separately for each sex, reaching an improved global error of 1.7% in both cases
277 (only one insect in each sex was erroneously classified, tables S1 and S2,
278 Supplementary Materials). Scripts of the model are available in R as Supplementary
279 Material and can be modified to include other species and compounds.

281 **Discussion**

282

283 Our study shows that the cuticular hydrocarbon profiles of *M. pygmaeus* and *M.*
284 *melanotoma* are markedly different from each other and can be used as a phenotypic
285 character to discriminate adults of these otherwise practically indistinguishable species.

286 Inclusion of a third species, *M. costalis*, which has a cuticular hydrocarbon profile that
287 is very similar to that of *M. pygmaeus*, but different from *M. melanotoma*, strengthens

288 the validity of the cuticular hydrocarbon method for species separation. Our results

289 agree with sequence variation of *mtDNA* where *M. melanotoma* and *M. pygmaeus*

290 separate in different clusters, whereas *M. pygmaeus* and *M. costalis* group in the same

291 cluster, and are therefore considered as sister species (Martínez-Cascales et al., 2006).

292 Crossing experiments between *M. pygmaeus* and *M. melanotoma* individuals collected

293 from the same host plants as in this study produced no progeny (O.A. & C. C., personal

294 observation), which is in accordance with the species status of the two taxa.

295

296 The prediction model results in 100% prediction of *M. melanotoma*, but the same model

297 is less successful at separating *M. pygmaeus* from *M. costalis*, as the hydrocarbon

298 profile of these two species are more similar to each other than to *M. melanotoma*.

299 However, *M. costalis* has a dark spot on the scutellum that distinguishes it from *M.*

300 *pygmaeus*, thus complete separation of all three species is possible. The same level of

301 discrimination between *M. pygmaeus* and *M. melanotoma* than the one we have

302 obtained is also achieved with *mtDNA* sequences (Martínez-Cascales et al., 2006), so

303 both methods are equally suitable to distinguish between the two species. Although

304 there were sex differences in cuticular hydrocarbon profiles, making a separate

305 prediction model for each sex reduces only slightly the prediction error for *M. pygmaeus*

306 and *M. costalis*, and thus it is not necessary to analyse males and females separately.
307 However, the two sexes are easily distinguished by eye, and so they could be analysed
308 separately if finer discrimination is needed. Diet also had an effect on the cuticular
309 hydrocarbons, but this effect was smaller than the differences among species, and thus
310 diet should not affect the ability of the model to predict species identity.

311

312 To our knowledge there is only one other report of cuticular hydrocarbons in mirids
313 (Drijfhout & Groot, 2001; Dridjfhout et al., 2003). We did not find any of the reported
314 *L. pabulinus* compounds in *Macrolophus* cuticular extracts, neither are they present in
315 several haematophagous triatomide heteroptera (Juárez & Fernández, 2007), however
316 there are similarities between the cuticular hydrocarbons of triatomide bugs and
317 *Macrolophus*. For example, the majority of compounds in both groups of insects are
318 mostly between 20 and 40 carbons, alkenes are few, and mono and methyl branched
319 alkanes are frequent, often with positions in carbons 11, 13, and 15. In contrast to the
320 pattern observed in triatomide bugs and *Macrolophus*, in three species of the antocorid
321 bug *Orius*, straight-chain alkanes, alkenes and single mono-methylated alkanes between
322 25 and 33 carbons in length comprise the most abundant cuticular hydrocarbons
323 (Nakabou & Ohno, 2001).

324

325 *M. melanotoma* males and females had very different cuticular hydrocarbon profiles,
326 but the hydrocarbon profiles of males and females of the other two *Macrolophus* species
327 were very similar. Sexual differences are also relatively small in the heteropterans
328 *Oncopeltus fasciatus* (Lygaeidae) and *Rhodnius prolixus* (Reduviidae) (Jackson, 1983;
329 Juárez et al., 2001). Whether sexual dimorphism of cuticular hydrocarbons plays a role
330 in mediating sexual recognition in *Macrolophus* is unknown, however there are many

331 examples in other insects where this is the case (reviewed in Thomas & Simmons,
332 2008). In the mirid *L. pabulinus* the leg hydrocarbons (Z)-9-pentacosene and (Z)-9-
333 heptacosene are produced in sex-specific amounts and could be involved in mate
334 location (Drijfhout & Groot, 2001; Drijfhout et al., 2003).

335

336 The effect of diet on cuticular hydrocarbon composition is well documented in social
337 omnivorous insects such as ants and cockroaches (Liang & Silverman, 2000; Guerrieri
338 et al., 2009). Although diet had a significant effect on the total quantity of cuticular
339 hydrocarbons of *Macrolophus*, the species- and sex-specific patterns of cuticular
340 hydrocarbons were not altered by the diet, suggesting that the cuticular hydrocarbon
341 composition of *Macrolophus* is largely under genetic control and not strongly
342 influenced by diet.

343

344

Acknowledgements

345

346 We thank Montse Lloveras for technical assistance with the chemical analysis. This
347 research was funded in part by the Spanish Department of Science and Innovation
348 (MICINN), projects AGL2005-03768, and AGL2006-08726 to OA and CC, and
349 AGL2007-62366 to CG.

350

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458 Press.
- 459

460 Table 1. Results from MANOVA using Pillai-Bartlett's statistic when analysing
461 variability due to species, sex and diet for 28 cuticular hydrocarbons of *Macrolophus*.

462

	df	Pillai statistic	Approx. F	Numerator df	Denominator df	p-value
Species	1	1.790	26.806	56	176	0
Sex	1	0.689	6.910	28	87	1.52E-09
Diet	1	0.747	9.198	28	87	5.40E-13
Residuals	114					

463

464

465 Table 2. Chemical structure of 28 cuticular hydrocarbons of three *Macrolophus* species.
 466 deduced from the characteristic ions (after GC-MSD) and the retention indices (RI)
 467 relative to straight chain hydrocarbons. PC1 and PC2: percent contribution of each
 468 compound to the principal components of the PCA shown in fig. 4.
 469

Peak number	RT (min)	RI	Tentative chemical structure	Diagnostic ions	PC1	PC2
1	22.1	2400	<i>n</i> -tetracosane	338	0.17	0.32
2	23.34	2557	11, 15-dimethylpentacosane	168, 211, 239	0.34	0.00
3	26.82	3069	hentriacontene	434 (M+)	3.46	0.41
4	27.2	3129	11-, 13-, 15-methylhentriacontane	168, 196, 224, 252, 280, 308	4.49	0.06
5	27.39	3157	11, 19-dimethylhentriacontane	168, 196, 295, 323	5.02	0.00
6	28.02	3248	triacontene	462 (M+)	3.95	0.47
7	28.1	3267	2-methyldotriacontane	421 (M-43), 449 (M-15)	4.12	5.68
8	28.58	3332	11-, 13-, 15-methyltriacontane	168, 196, 224, 252, 280, 308, 336	4.86	0.07
9	28.77	3355	mixture of di and tri-methyltriacontanes	168, 196, 181, 211, 239, 252, 267, 323, 356	4.80	0.00
10	29.71	3465	2-methyltetracontane	449 (M-43), 477 (M-15)	5.42	0.03
11	29.98	3492	2, 24-dimethyltetracontane	168, 336, 365, 463 (M-43), 491 (M-15)	1.21	11.57
12	30.58	3552	11, 15-dimethylpentatriacontane	168, 211, 239, 280, 308, 379	0.98	12.96
13	30.81	3570	11, 15, 19-trimethylpentatriacontane	168, 239, 252, 309, 323, 393	0.58	13.20
14	31.43	3652	12, 16-, 12, 18-dimethylhexatriacontane	182, 253, 280, 308, 351, 379	0.05	9.59
15	31.68	3669	3-methylhexatriacontane	491 (M-29)	0.10	15.23
16	31.97	3692	4, 10- and 4, 12-dimethylhexatriacontane	168, 197, 225, 336, 364, 393	3.04	6.94
17	32.17	3711	4, 10, 14-trimethylhexatriacontane	168, 239, 336, 407	5.82	0.21
18	32.63	3729	11-, 13-, 15-methylheptatriacontane	168, 196, 224, 280, 308, 336, 364	1.87	8.22
19	33.03	3751	11, 15-, and 11, 19-dimethylheptatriacontane	168, 239, 280, 295, 407	2.44	8.90
20	33.35	3771	11, 15, 19-trimethylheptatriacontane	168, 239, 280, 309, 351	6.73	0.17
21	33.61	3789	11, 15, 23-trimethylheptatriacontane	168, 224, 239	5.15	0.84
22	34.15	3837	unknown		6.31	0.14
23	34.48	3849	12, 16-dimethyloctatriacontane	182, 407, 253, 336, 309	6.04	1.21
24	34.84	3868	unknown	182, 225	4.85	0.65
25	36.31	3953	11, 21-dimethylnonatriacontane	168, 280, 295, 323, 407, 435	5.47	0.99
26	36.7	3973	11, 15, 19-trimethylnonatriacontane	168, 239, 309, 379, 449	5.33	0.64
27	37.89	4072	12, 16-dimethyltetracontane	182, 253, 378, 449	3.63	1.12
28	40.85	4186	11, 15, 19-trimethylhentetracontane	168, 239, 280, 308	3.78	0.38
Total					100	100

470

471

472

473 Table 3. Prediction errors in the classification of three *Macrolophus* species based on a
 474 quadratic discriminant analysis (QDA) using the three main principal components of the
 475 cuticular hydrocarbons of males and females. The number in each cell represents the
 476 number of individuals of the expected species (column) assigned to the three predicted
 477 species (rows). A total 119 insects were analysed.

Predicted species by QDA	Expected species			Total
	<i>M. melanotoma</i>	<i>M. pygmaeus</i>	<i>M. costalis</i>	
<i>M. melanotoma</i>	39	0	0	39
<i>M. pygmaeus</i>	0	38	2	40
<i>M. costalis</i>	0	5	35	40
Total	39	43	37	119
Prediction error (%)	0	13.16	5.4	5.88

478

479

480 Fig. 1. Representative gas chromatograph traces of *Macrolophus* cuticular
481 hydrocarbons. Each trace corresponds to a different single individual. The most
482 representative peaks were numbered 1 to 28, corresponding with the column elution
483 time. Notice the large difference between *M. melanotoma* and the other two species as
484 well as between *M. melanotoma* males and females. Arrow indicates the characteristic
485 dark spot on the scutellum of *M. costalis*.

486

487 Fig. 2. Relative abundance of male and female cuticular hydrocarbons in three
488 *Macrolophus* species. Abundance is the percentage relative to the internal standard.
489 Asterisks above each bar pair indicate significant difference between males (grey bars)
490 and females (empty bars) (t-test, $P < 0.0017$). Different letters for a given compound
491 indicate significant differences among species in females (capital letters) or males
492 (small letters) (ANOVA followed by Tukey's test, $P < 0.0017$ in both).

493

494 Fig. 3. Effect of diet on *Macrolophus* cuticular hydrocarbons. One group ("field"
495 individuals) fed on their natural diet (white bars), and another group ("laboratory"
496 individuals) were reared in a standard tobacco and moth egg diet (black bars).
497 Abundance is the percentage relative to the internal standard. Asterisks above a bar pair
498 indicate significant difference between diets (t-test, $P < 0.0017$).

499

500 Fig. 4. Contribution of the two largest principal components (PC) from a PCA of
501 *Macrolophus* cuticular hydrocarbons. PC1 explains 46 % of the variability and
502 separates *M. melanotoma* males (white circles) from the females (black circles), and
503 from the other two species (*M. pygmaeus*, triangles, *M. costalis*, squares). PC2 explains
504 22% of the variability and separates *M. melanotoma* females from the males and from

505 the other two species. Laboratory individuals marked with a dot, field individuals
506 unmarked.

1

2

3

4

5 **Cuticular hydrocarbons discriminate cryptic *Macrolophus***

6 **species (**Hemiptera**: Miridae)**

7

8 **César Gemeno^{1*}, Nerea Laserna¹, Magí Riba², Joan Valls³, Cristina**

9 **Castañé⁴ and Oscar Alomar⁴**

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11

12 ¹ *Department of Crop and Forest Science, University of Lleida, 25198 Lleida, Spain*

13 ² *Department of Chemistry, University of Lleida, 25198 Lleida, Spain*

14 ³ *Biostatistics Unit. Biomedical Research Institute (IRBLLEIDA), 25198 Lleida, Spain*

15 ⁴ *IRTA Cabrils, 08348 Barcelona, Spain*

16

17

18 *Corresponding author

19 Fax: +34 973-702690

20 Telephone: +34 973-702531

21 e-mail: cesar.gemeno@pvcf.udl.cat

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23 *Macrolophus pygmaeus* is commercially employed in the biological control of
24 greenhouse and field vegetable pests. It is morphologically undistinguishable from the
25 cryptic species *M. melanotoma*, and this interferes with the evaluation of the biological
26 control activity of *M. pygmaeus*. We analysed the potential of cuticular hydrocarbon
27 composition as a method to discriminate the two *Macrolophus* species. A third species,
28 *M. costalis*, which is morphologically different from the other two species by having a
29 dark spot at the tip of the scutellum, served as a control. **Sex, diet and species, all had**
30 **significant effects in the cuticular hydrocarbon profiles, but the variability associated to**
31 **sex or diet was smaller than among species.** Discriminant quadratic analysis of cuticular
32 hydrocarbons confirmed the results of previous molecular genetic studies and showed,
33 using cross-validation methods, that *M. pygmaeus* can be discriminated from *M. costalis*
34 and *M. melanotoma* with prediction errors of 6.75% and 0%, respectively. Therefore,
35 cuticular hydrocarbons can be used to separate *M. pygmaeus* and *M. melanotoma*, while
36 *M. costalis* and *M. pygmaeus* can be readily distinguished from each other by the dark
37 tip of the scutellum of the former one.

38

39 **Keywords:** cuticular hydrocarbons, biological control, taxonomy

40

41

Introduction

42

43

44 Several species in the genus *Macrolophus* (Hemiptera: Miridae) are efficient predators
45 of vegetable crops pests (e.g. whiteflies, aphids and thrips) (Alvarado et al., 1997;
46 Riudavets & Castañé 1998; Montserrat et al., 2000; Alomar et al., 2006; Hansen et al.,
47 1999; Athanassiou et al., 2003; Margaritopoulos et al., 2003). Control strategies that
48 incorporate *Macrolophus* species are based on both seasonal inoculative releases of
49 commercially produced individuals and habitat management to conserve natural
50 populations and enhance colonization (Alomar et al., 2002). In order to avoid the
51 incorrect use of predator reservoir plants it is essential to correctly identify the predator
52 species.

53

54 Several *Macrolophus* species are morphologically similar or have highly variable
55 taxonomic characters, which hampers correct identification and questions species-
56 identity status (Josifov, 1992; Kerzhner & Josifov, 1999). This is particularly true for
57 the two species that have been **most** cited as biological control agents, *M. pygmaeus*
58 (Rambur 1839), and *M. melanotoma* (Costa 1853) (the last one mostly cited as its junior
59 synonym *M. caliginosus* Wagner 1951), and has probably resulted in incorrect
60 attribution of biological control by *M. pygmaeus* to *M. melanotoma* (see Martínez-
61 Casales et al., 2006 for a historical review). DNA methods have been employed recently
62 to distinguish *M. pygmaeus* from *M. melanotoma* (Perdikis et al., 2003; Martínez-
63 Casales et al., 2006). Cuticular hydrocarbon analysis could provide an additional
64 method to separate morphologically similar species (Bagnères & Wicker-Thomas,
65 2010).

66

67 The insect cuticle is coated with a thin lipid layer containing a high percentage of
68 hydrocarbons (linear, branched, saturated and unsaturated), one of which main functions
69 is to prevent desiccation and pathogen entrance into the body (Howard & Blomquist,
70 2005). Cuticular hydrocarbons are usually present in high amounts and are easy to
71 extract by quickly rinsing the specimen in non-polar organic solvents (Blomquist et al.,
72 1987). In addition, most cuticular hydrocarbons are chemically stable and not very
73 volatile (Martin et al., 2009). The hydrocarbon blend is often species specific and
74 therefore it is a useful character in insect taxonomy (reviewed by Bagnères & Wicker-
75 Thomas, 2010).

76

77 In contrast to other insect groups, there are relatively few studies of cuticular
78 hydrocarbon taxonomy in **Hemiptera**. A recent publication reviews the hydrocarbons of
79 blood-sucking bugs (Juárez & Fernández, 2007), and isolated studies compare cuticular
80 hydrocarbons of *Orius* species (Anthocoridae) (Nakabou & Ohno, 2001), and aphids
81 (Clements et al., 2000). With regard to mirids, analysis of cuticular hydrocarbons has
82 been reported only for *Lygocoris pabulinus* (L.) (Drijfhout & Groot, 2001).

83

84 The purpose of the present study is to **identify the cuticular hydrocarbons of three**
85 ***Macrolophus* species (*M. melanotoma*, *M. pygmaeus*, and *M. costalis*), as they may be**
86 **important in species recognition and mating behaviour. Since diet and sex may affect**
87 **insect cuticular hydrocarbon composition (Liang & Silverman, 2000; Thomas &**
88 **Simmons, 2008; Guerrieri et al., 2009), we also analysed the effect of diet (i.e., host**
89 **plant and associated prey) and sex on cuticular hydrocarbons. We then determine if**
90 cuticular hydrocarbons can be used to distinguish adults of two *Macrolophus* species

91 (*M. melanotoma* and *M. pygmaeus*) that cannot be distinguished easily by external
92 morphological characters alone. A third predatory species, *M. costalis* Fieber, was
93 included in the study to confirm previous results on the taxonomic relationship of the
94 three species (Martínez-Cascales et al., 2006).

95

96 **Materials and methods**

97

98 *Insects*

99

100 Chemical analyses were performed on adults of both sexes. Wild individuals were
101 collected in the proximities of Cabrils (Barcelona, Spain) in the spring of 2008.
102 *Macrolophus melanotoma* was collected from *Dittrichia viscosa* (L.) Greuter
103 (Compositae), where it is abundant (O. A., personal observation). *Macrolophus*
104 *costalis*, which is readily distinguished by the dark spot on the scutellum, was collected
105 on *Cistus albidus* L. (Cistaceae), where it is commonly found (O.A., personal
106 observation). *M. pygmaeus* was obtained from a > 5-years-old laboratory colony
107 originated from tomato fields, and maintained on tobacco plants and frozen *Ephestia*
108 *kuehniella* Zeller (Lepidoptera: Pyralidae) egg prey. This colony is refreshed annually
109 with individuals collected in local tomato fields (Cabrils, Barcelona, Spain).
110 Examination of the length and shape of the respiratory horns of eggs (Perdikis et al.,
111 2003; Alomar & Goula, unpub. results) was used to confirm the identity of collected
112 females of *M. melanotoma* and *M. pygmaeus*. Some of those individuals were frozen (-
113 20°C) immediately or within 7 days after being collected (in which case they were
114 maintained in their respective diet), and are referred to as the “field” individuals.
115 Additionally, in order to evaluate the effect of diet on the cuticular hydrocarbon profile,
116 4th-instar *M. pygmaeus* nymphs taken from the colony, and the progeny of field-
117 collected *M. melanotoma* and *M. costalis*, were reared individually on tobacco leaves
118 and *E. kuehniella* eggs. The resulting adults, referred to as the “laboratory” individuals,
119 were frozen when 3 to 6 days old and analyzed at the same time as the “field”
120 individuals described above.

121
122 Cages used for oviposition and nymphal development (7-cm diameter x 3.5-cm high
123 with a ventilated lid) had a layer of 0.5% agar and a tobacco leaf disc placed on top of
124 it, with the abaxial surface facing upwards, and were kept at $25 \pm 2^\circ\text{C}$ and $85 \pm 5\%$ RH,
125 under a 16:8 light:dark photoregime. Individuals were moved to new cages as needed
126 and were provided with frozen *E. kuehniella* egg prey twice a week.

127

128 *Chemical Analyses*

129

130 Individuals were taken out of the freezer one at a time, let defrost, and submerged in 20
131 μl of hexane (HPLC grade, Sigma-Aldrich, Madrid, Spain) containing 50 ng of
132 pentadecane (98% pure, Sigma-Aldrich, Madrid, Spain) in a conical-bottom hexane-
133 rinsed glass vial. The use of internal standard allows minimization of any differences in
134 injection volume and in the daily response of the equipment. Previous analyses
135 indicated that pentadecane is not present in significant amounts in cuticular extracts of
136 these species. Five minutes after immersion in the solvent the insect was removed and
137 the solution was either analysed immediately or returned to the freezer (-20°C) for later
138 analysis.

139

140 For chemical analyses the volume of the extract was reduced immediately before
141 injection to 2-4 μl with a gentle nitrogen stream and 1 μl was injected manually in a gas
142 chromatograph (GC) in the splitless mode (split valve opened after 1 min). Samples
143 were analysed in either a GC-FID or a GC-MSD (Agilent Technologies 6890N GC, and
144 5973 Network quadrupole MSD), each equipped with a DB-5 column (30 m x of 0.32
145 mm ID x 0.25 μm , Agilent Technologies), and run with the same temperature program:

146 start at 60°C for 1 min, then increase to 320°C at 10 °C/min, and maintain at this
147 temperature for 25 min. Carrier gas was Helium at constant flow (1 ml/min), the
148 injector was set a 250°C and the detector at 280°C.

149

150 A total of 119 insects were analysed by GC-FID, and these are the samples used in the
151 statistical comparisons [10 individuals of each species, sex and diet (laboratory versus
152 field), except for N=9 field *M. melanotoma* males]. Two additional insects of each sex
153 and species were analysed by GC-MSD, and these data were used in the chemical
154 identification of the compounds. Straight-chain alkanes were identified by comparison
155 of retention times and mass spectra of authentic standards (Fluka, alkane standard
156 solutions 04070 (C₈-C₂₀) and 04071 (C₂₁-C₄₀), Sigma-Aldrich, Madrid, Spain).

157 Retention indices (RIs) were calculated according to van den Dool & Kratz (1963).

158 Linear and methyl branched alkanes were tentatively identified by comparison of their
159 RI and the characteristic ion fragments with those reported in the literature (Blomquist
160 et al., 1987; Juárez & Blomquist, 1993; Carlson et al., 1998; Juárez et al., 2001; Mullen
161 et al., 2008; Gomes et al., 2008; Dall’Aglia-Holvorcem et al., 2009).

162

163 Chain length of the selected compounds ranged between C₂₄ and more than C₄₀. Since
164 our largest alkane standard was C₄₀ we fitted a curve to the straight chain alkane
165 standards C₂₇ to C₄₀ to extrapolate the retention times of C₄₁ and C₄₂ ($R^2 = 0.99$, $y =$
166 $125.161 - 0.066 * x + 1.105 * 10^{-5} * x^2$, where $y =$ estimated RT and $x =$ chain length), and
167 then estimated the RI of sample peaks 27 and 28 (table 1).

168

169

Statistical Analyses

170

171 Percent relative abundance [(area of the target peak/area of the internal standard peak) x
172 100] was the parameter used in all the group comparisons. We chose those compounds
173 that showed a high relative abundance combined with a moderate coefficient of
174 variation, relative to other compounds, and that occurred consistently in at least one
175 class (i.e., species, sex, or diet). This resulted in the selection of 28 peaks, some of
176 which were abundant in one sex or species but below threshold level in others. A zero
177 value was assigned to undetected compounds. A Kolmogorov-Smirnov test was used to
178 assess normality of the data.

179

180 Two statistical analyses were performed. The first analysis was carried out to determine
181 the effect of diet, sex and species on the cuticular hydrocarbons. First we performed a
182 multivariate analysis of variance (MANOVA) in order to study how the 28 cuticular
183 hydrocarbons altogether vary with species, sex and diet. To measure the variability that
184 could be explained by each of these factors we computed the Pillai-Bartlett statistic
185 (Hand and Taylor, 1987), which provides a measure of the ratio of effect variance to
186 error variance. To assess the effect of individual cuticular hydrocarbons, the three
187 species were compared, compound by compound, males and females separately, using
188 ANOVA followed by a Tukey's mean comparison test. In addition, the effect of sex and
189 diet on each cuticular hydrocarbon compound was analysed with *t*-tests. Bonferroni
190 method was used to correct for multiple testing so that the threshold of significance was
191 set at $0.05/28 = 0.0017$ in the ANOVA and *t*-tests.

192

193 The second statistical analysis was performed to build a model to predict species
194 identity. We performed *Principal component analysis* (PCA), using the correlation
195 matrix of untransformed data, to obtain uncorrelated scores, visualize aggregation

196 patterns of the different insect groups, and to determine which compounds had the
197 strongest effect in this pattern. To build a prediction model for the species, the main
198 principal components (those adding at least 78% total variability) were used in a
199 *Quadratic discriminant analysis (QDA)*, and the prediction error of discriminant
200 functions was evaluated using a 10-fold cross-validation method. The statistical tests
201 were performed with R software (R Development Core Team, 2008). Analysis codes of
202 the prediction model are available as Supplementary Material.

203

204

205 **Results**

206

207 **Representative chromatograms indicating the 28 diagnostic peaks (by order of elution)**
208 **are shown in fig. 1. Normality assumption was accepted for all samples ($P < 0.0017$).**
209 **Significant differences in cuticular hydrocarbon profiles for diet, sex and species were**
210 **observed (MANOVA, $P < 0.001$, table 1). The differences between sexes or diets were**
211 **much smaller than those between species. Using the approximated F-statistic we found**
212 **that variability among species is almost 4 times higher than between males and females**
213 **($26.8/6.9 = 3.87$) and almost 3 times higher than between diets ($26.8/9.1 = 2.91$).**

214

215 Statistical comparison of individual compounds among the three species confirmed the
216 pattern shown in fig. 1: a large proportion of compounds were similar between *M.*
217 *pygmaeus* and *M. costalis*, and different from *M. melanotoma* (fig. 2, ANOVA, $P <$
218 0.0017). Roughly the first half (compounds 2 to 15) were more abundant in *M.*
219 *melanotoma* than in the other two species, and the second half (compounds 16 to 28)
220 were less abundant in *M. melanotoma* than in the other species. Therefore *M.*
221 *melanotoma* is different from the other two species by having a larger quantity of the
222 early eluting compounds, and a smaller quantity of the late eluting compounds (fig. 2).
223 Sex differences in **cuticular hydrocarbons** were numerous in *M. melanotoma*, scarce in
224 *M. pygmaeus* and lacking in *M. costalis* (fig. 2, *t-test*, $P < 0.0017$). **The cuticular**
225 **hydrocarbon blends of *M. melanotoma* males and females were clearly different (figs. 1**
226 **and 2). Individuals reared in the laboratory in isolation with a homogeneous diet of**
227 **tobacco and moth eggs had a higher quantity of some hydrocarbons than their “field”**
228 **relatives (fig. 3, *t-test*, $P < 0.0017$), but the species-specific cuticular hydrocarbon**
229 **pattern was not altered (fig. 3), thus indicating a weak effect by diet.**

230

231 Linear alkanes, from C₂₁ to C₃₅, were identified in the cuticular samples of the 3
232 species, but in very small quantities, and only one of them (C₂₄ or tetracosane,
233 compound 1, table 2) was abundant enough to be included among the 28 selected
234 compounds. The rest of the hydrocarbons consisted of unbranched monoenes and
235 alkanes with one or more methyl branches (table 2). Ion fragmentation suggests that
236 some peaks may contain more than one compound. A characteristic compound of *M.*
237 *melanotoma* is 2-methyltriacontane (compound 7). The largest difference of *M.*
238 *melanotoma* females with the other two species, besides compound 7, was a larger
239 quantity of compounds 12 and 13 (11, 15-dimethylpentatriacontane and 11, 15, 19-
240 trimethyl pentatriacontane). For *M. melanotoma* males the largest difference with the
241 other two species was a larger quantity of early eluting compounds 2 to 5. *M.*
242 *melanotoma* males and females differed in 25 of the 28 hydrocarbons: the first 6, by
243 order of elution, were more abundant in males, and the other 19 were more abundant in
244 females (fig. 2). To name some examples, males had more hentriacontene and
245 triacontene (compounds 3 and 6) than females, and females had more 11, 15-
246 dimethylpentatriacontane and 11, 15, 19-trimethylheptatriacontane (compounds 12 and
247 20) than males. Compound 7 (2-methyltriacontane), which is characteristic of this
248 species, was not different between sexes.

249

250 PC1 and PC2 explained 46.28 % and 21.73 % of the variability, respectively, and
251 clearly segregated *M. melanotoma* from the other two species, whereas sex differences
252 were only apparent in *M. melanotoma* (fig. 4). Segregation between field and laboratory
253 individuals was weak and only apparent in *M. melanotoma* (fig. 4), thus indicating that
254 diet did not affect the species-specific cuticular hydrocarbon blend. Different

255 compounds contributed in varying degrees to the PCs (table 2). PC1, which separated
256 *M. melanotoma* males from the other two species and from the females (fig. 4), was
257 mainly influenced by compounds that were characteristically small in *M. melanotoma*
258 males (compounds 17, 20, 22, 23 and 25, table 2, fig. 2). PC2, which separated *M.*
259 *melanotoma* females from males and from the other two species, was strongly
260 influenced by compounds 11 to 15 (table 2), which were more abundant in *M.*
261 *melanotoma* females than in the other insects (fig. 2).

262

263 **With regard to the prediction model**, a QDA based on PC1, PC2 and PC3 (which
264 together explained 78% of total variability) was used to predict the species of each test
265 individual (see Supplementary Material files). QDA was able to correctly classify all *M.*
266 *melatonoma* individuals, but 5 *M. costalis* individuals were classified as *M. pygmaeus*
267 and 2 *M. pygmaeus* were classified as *M. costalis* (table 3). Therefore the global error
268 was 5.88% when comparing these three species. To further assess the error prediction
269 level, we first run a training and test method using 3 quarters of the sample (90
270 individuals chosen at random) to build a QDA, and the remaining quarter of the sample
271 (29 individuals chosen at random) to test the prediction accuracy, which resulted in a
272 3.5% global error. Next, we used a 10-fold cross-validation method which indicated a
273 global prediction error of 6.75% (ranging from 0% to 25%), attributed to the separation
274 between *M. pygmaeus* and *M. costalis*, given that the error rate when discriminating *M.*
275 *melatonoma* from *M. pygmaeus* or *M. costalis* was 0% in all cases. We also performed
276 QDA separately for each sex, reaching an improved global error of 1.7% in both cases
277 (only one insect in each sex was erroneously classified, tables S1 and S2,
278 Supplementary Materials). Scripts of the model are available in R as Supplementary
279 Material and can be modified to include other species and compounds.

281 **Discussion**

282

283 Our study shows that the cuticular hydrocarbon profiles of *M. pygmaeus* and *M.*
284 *melanotoma* are markedly different from **each other** and can be used as a phenotypic
285 character to discriminate adults of these otherwise practically indistinguishable species.

286 **Inclusion of a third species, *M. costalis*, which has a cuticular hydrocarbon profile that**
287 **is very similar to that of *M. pygmaeus*, but different from *M. melanotoma*, strengthens**
288 **the validity of the cuticular hydrocarbon method for species separation. Our results**

289 agree with sequence variation of *mtDNA* where *M. melanotoma* and *M. pygmaeus*
290 separate in different clusters, whereas *M. pygmaeus* and *M. costalis* group in the same
291 cluster, and are therefore considered as sister species (Martínez-Cascales et al., 2006).

292 Crossing experiments between *M. pygmaeus* and *M. melanotoma* individuals collected
293 from the same host plants as in this study produced **no** progeny (O.A. & C. C., personal
294 observation), which is in accordance with the species status of the two taxa.

295

296 The prediction model results in 100% prediction of *M. melanotoma*, but the same model
297 is less successful at separating *M. pygmaeus* from *M. costalis*, as the hydrocarbon
298 profile of these two species are more similar to each other than to *M. melanotoma*.

299 **However, *M. costalis* has a dark spot on the scutellum that distinguishes it from *M.***
300 ***pygmaeus*, thus complete separation of all three species is possible. The same level of**

301 discrimination between *M. pygmaeus* and *M. melanotoma* **than the one we have**
302 **obtained is also** achieved with *mtDNA* sequences (Martínez-Cascales et al., 2006), so

303 both methods are equally suitable to distinguish **between the two** species. **Although**

304 **there were sex differences in cuticular hydrocarbon profiles**, making a separate

305 prediction model for each sex reduces **only slightly** the prediction error for *M. pygmaeus*

306 and *M. costalis*, and thus it is not necessary to analyse males and females separately.
307 However, the two sexes are easily distinguished by eye, and so they could be analysed
308 separately if finer discrimination is needed. Diet also had an effect on the cuticular
309 hydrocarbons, but this effect was smaller than the differences among species, and thus
310 diet should not affect the ability of the model to predict species identity.

311

312 To our knowledge there is only one other report of cuticular hydrocarbons in mirids
313 (Drijfhout & Groot, 2001; Dridjfhout et al., 2003). We did not find any of the reported
314 *L. pabulinus* compounds in *Macrolophus* cuticular extracts, neither are they present in
315 several haematophagous triatomide heteroptera (Juárez & Fernández, 2007), however
316 there are similarities between the cuticular hydrocarbons of triatomide bugs and
317 *Macrolophus*. For example, the majority of compounds in both groups of insects are
318 mostly between 20 and 40 carbons, alkenes are few, and mono and methyl branched
319 alkanes are frequent, often with positions in carbons 11, 13, and 15. In contrast to the
320 pattern observed in triatomide bugs and *Macrolophus*, in three species of the antocorid
321 bug *Orius*, straight-chain alkanes, alkenes and single mono-methylated alkanes between
322 25 and 33 carbons in length comprise the most abundant cuticular hydrocarbons
323 (Nakabou & Ohno, 2001).

324

325 *M. melanotoma* males and females had very different cuticular hydrocarbon profiles,
326 but the hydrocarbon profiles of males and females of the other two *Macrolophus* species
327 were very similar. Sexual differences are also relatively small in the heteropterans
328 *Oncopeltus fasciatus* (Lygaeidae) and *Rhodnius prolixus* (Reduviidae) (Jackson, 1983;
329 Juárez et al., 2001). Whether sexual dimorphism of cuticular hydrocarbons plays a role
330 in mediating sexual recognition in *Macrolophus* is unknown, however there are many

331 examples in other insects where this is the case (reviewed in Thomas & Simmons,
332 2008). In the mirid *L. pabulinus* the leg hydrocarbons (Z)-9-pentacosene and (Z)-9-
333 heptacosene are produced in sex-specific amounts and could be involved in mate
334 location (Drijfhout & Groot, 2001; Drijfhout et al., 2003).

335

336 The effect of diet on cuticular hydrocarbon composition is well documented in social
337 omnivorous insects such as ants and cockroaches (Liang & Silverman, 2000; Guerrieri
338 et al., 2009). Although diet had a significant effect on the total quantity of cuticular
339 hydrocarbons of *Macrolophus*, the species- and sex-specific patterns of cuticular
340 hydrocarbons were not altered by the diet, suggesting that the cuticular hydrocarbon
341 composition of *Macrolophus* is largely under genetic control and not strongly
342 influenced by diet.

343

344

Acknowledgements

345

346 We thank Montse Lloveras for technical assistance with the chemical analysis. This
347 research was funded in part by the Spanish Department of Science and Innovation
348 (MICINN), projects AGL2005-03768, and AGL2006-08726 to OA and CC, and
349 AGL2007-62366 to CG.

350

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- 459

460 Table 1. Results from MANOVA using Pillai-Bartlett's statistic when analysing
461 variability due to species, sex and diet for 28 cuticular hydrocarbons of *Macrolophus*.

462

	df	Pillai statistic	Approx. F	Numerator df	Denominator df	p-value
Species	1	1.790	26.806	56	176	0
Sex	1	0.689	6.910	28	87	1.52E-09
Diet	1	0.747	9.198	28	87	5.40E-13
Residuals	114					

463

464

465 **Table 2.** Chemical structure of 28 cuticular hydrocarbons of three *Macrolophus* species.
 466 deduced from the characteristic ions (after GC-MSD) and the retention indices (RI)
 467 relative to straight chain hydrocarbons. PC1 and PC2: percent contribution of each
 468 compound to the principal components of the PCA shown in fig. 4.

469

Peak number	RT (min)	RI	Tentative chemical structure	Diagnostic ions	PC1	PC2
1	22.1	2400	<i>n</i> -tetracosane	338	0.17	0.32
2	23.34	2557	11, 15-dimethylpentacosane	168, 211, 239	0.34	0.00
3	26.82	3069	hentriacontene	434 (M+)	3.46	0.41
4	27.2	3129	11-, 13-, 15-methylhentriacontane	168, 196, 224, 252, 280, 308	4.49	0.06
5	27.39	3157	11, 19-dimethylhentriacontane	168, 196, 295, 323	5.02	0.00
6	28.02	3248	tritriacontene	462 (M+)	3.95	0.47
7	28.1	3267	2-methyldotriacontane	421 (M-43), 449 (M-15)	4.12	5.68
8	28.58	3332	11-, 13-, 15-methyltritriacontane	168, 196, 224, 252, 280, 308, 336	4.86	0.07
9	28.77	3355	mixture of di and tri-methyltritriacontanes	168, 196, 181, 211, 239, 252, 267, 323, 356	4.80	0.00
10	29.71	3465	2-methyltetracontane	449 (M-43), 477 (M-15)	5.42	0.03
11	29.98	3492	2, 24-dimethyltetracontane	168, 336, 365, 463 (M-43), 491 (M-15)	1.21	11.57
12	30.58	3552	11, 15-dimethylpentatriacontane	168, 211, 239, 280, 308, 379	0.98	12.96
13	30.81	3570	11, 15, 19-trimethylpentatriacontane	168, 239, 252, 309, 323, 393	0.58	13.20
14	31.43	3652	12, 16-, 12, 18-dimethylhexatriacontane	182, 253, 280, 308, 351, 379	0.05	9.59
15	31.68	3669	3-methylhexatriacontane	491 (M-29)	0.10	15.23
16	31.97	3692	4, 10- and 4, 12-dimethylhexatriacontane	168, 197, 225, 336, 364, 393	3.04	6.94
17	32.17	3711	4, 10, 14-trimethylhexatriacontane	168, 239, 336, 407	5.82	0.21
18	32.63	3729	11-, 13-, 15-methylheptatriacontane	168, 196, 224, 280, 308, 336, 364	1.87	8.22
19	33.03	3751	11, 15-, and 11, 19-dimethylheptatriacontane	168, 239, 280, 295, 407	2.44	8.90
20	33.35	3771	11, 15, 19-trimethylheptatriacontane	168, 239, 280, 309, 351	6.73	0.17
21	33.61	3789	11, 15, 23-trimethylheptatriacontane	168, 224, 239	5.15	0.84
22	34.15	3837	unknown		6.31	0.14
23	34.48	3849	12, 16-dimethyloctatriacontane	182, 407, 253, 336, 309	6.04	1.21
24	34.84	3868	unknown	182, 225	4.85	0.65
25	36.31	3953	11, 21-dimethylnonatriacontane	168, 280, 295, 323, 407, 435	5.47	0.99
26	36.7	3973	11, 15, 19-trimethylnonatriacontane	168, 239, 309, 379, 449	5.33	0.64
27	37.89	4072	12, 16-dimethyltetracontane	182, 253, 378, 449	3.63	1.12
28	40.85	4186	11, 15, 19-trimethylhentetracontane	168, 239, 280, 308	3.78	0.38
Total					100	100

470

471

472

473 **Table 3.** Prediction errors in the classification of three *Macrolophus* species based on a
 474 quadratic discriminant analysis (QDA) using the three main principal components of the
 475 cuticular hydrocarbons of males and females. The number in each cell represents the
 476 number of individuals of the expected species (column) assigned to the three predicted
 477 species (rows). A total 119 insects were analysed.

Predicted species by QDA	Expected species			Total
	<i>M. melanotoma</i>	<i>M. pygmaeus</i>	<i>M. costalis</i>	
<i>M. melanotoma</i>	39	0	0	39
<i>M. pygmaeus</i>	0	38	2	40
<i>M. costalis</i>	0	5	35	40
Total	39	43	37	119
Prediction error (%)	0	13.16	5.4	5.88

478

479

480 Fig. 1. Representative gas chromatograph traces of *Macrolophus* cuticular
481 hydrocarbons. Each trace corresponds to a different single individual. The most
482 representative peaks were numbered 1 to 28, corresponding with the column elution
483 time. Notice the large difference between *M. melanotoma* and the other two species as
484 well as between *M. melanotoma* males and females. Arrow indicates the characteristic
485 dark spot on the scutellum of *M. costalis*.

486

487 Fig. 2. Relative abundance of male and female cuticular hydrocarbons in three
488 *Macrolophus* species. Abundance is the percentage relative to the internal standard.
489 Asterisks above each bar pair indicate significant difference between males (grey bars)
490 and females (empty bars) (t-test, $P < 0.0017$). Different letters for a given compound
491 indicate significant differences among species in females (capital letters) or males
492 (small letters) (ANOVA followed by Tukey's test, $P < 0.0017$ in both).

493

494 Fig. 3. Effect of diet on *Macrolophus* cuticular hydrocarbons. One group ("field"
495 individuals) fed on their natural diet (white bars), and another group ("laboratory"
496 individuals) were reared in a standard tobacco and moth egg diet (black bars).
497 Abundance is the percentage relative to the internal standard. Asterisks above a bar pair
498 indicate significant difference between diets (t-test, $P < 0.0017$).

499

500 Fig. 4. Contribution of the two largest principal components (PC) from a PCA of
501 *Macrolophus* cuticular hydrocarbons. PC1 explains 46 % of the variability and
502 separates *M. melanotoma* males (white circles) from the females (black circles), and
503 from the other two species (*M. pygmaeus*, triangles, *M. costalis*, squares). PC2 explains
504 22% of the variability and separates *M. melanotoma* females from the males and from

505 the other two species. Laboratory individuals marked with a dot, field individuals
506 unmarked.

Figure 1

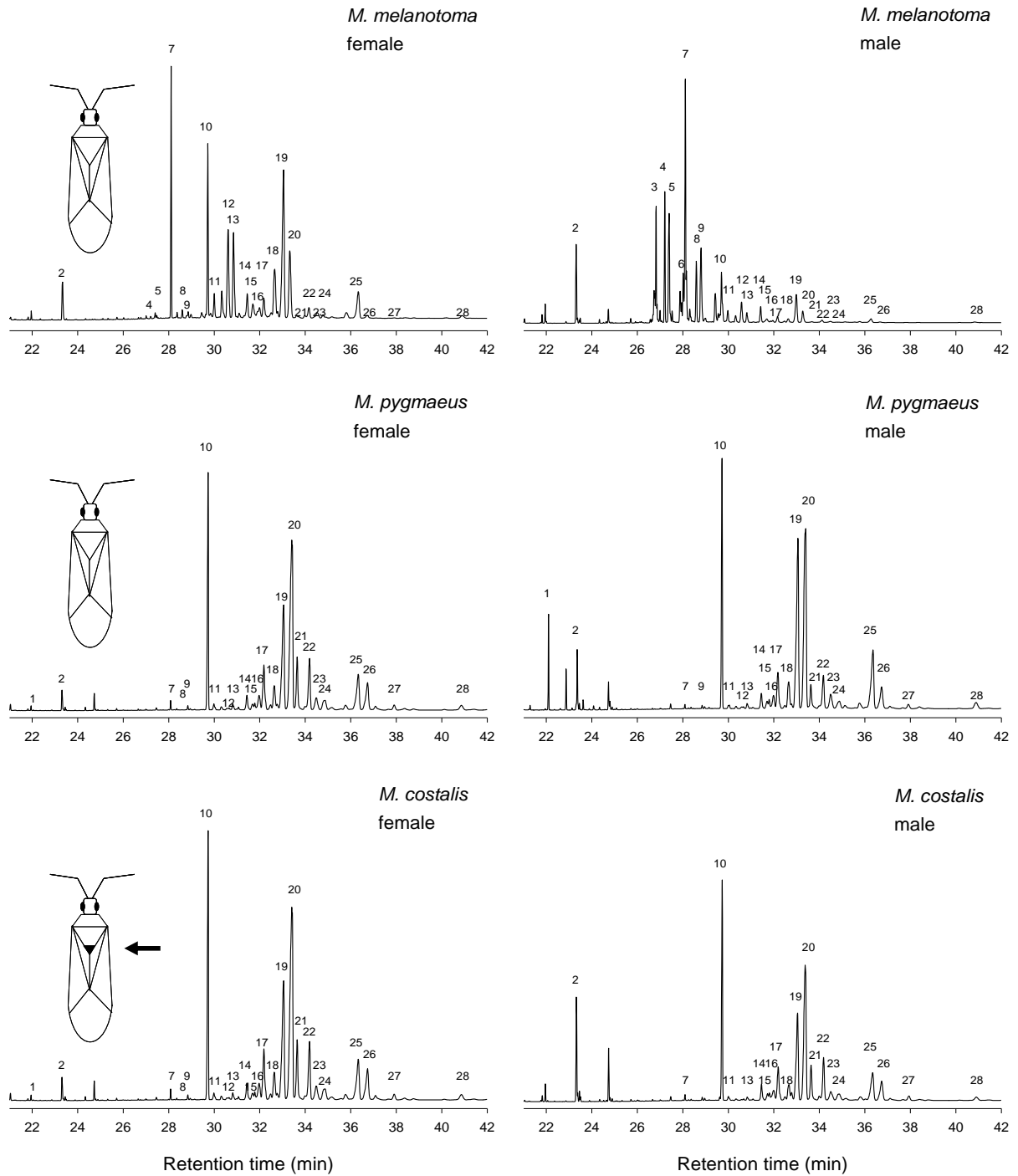


Figure 2

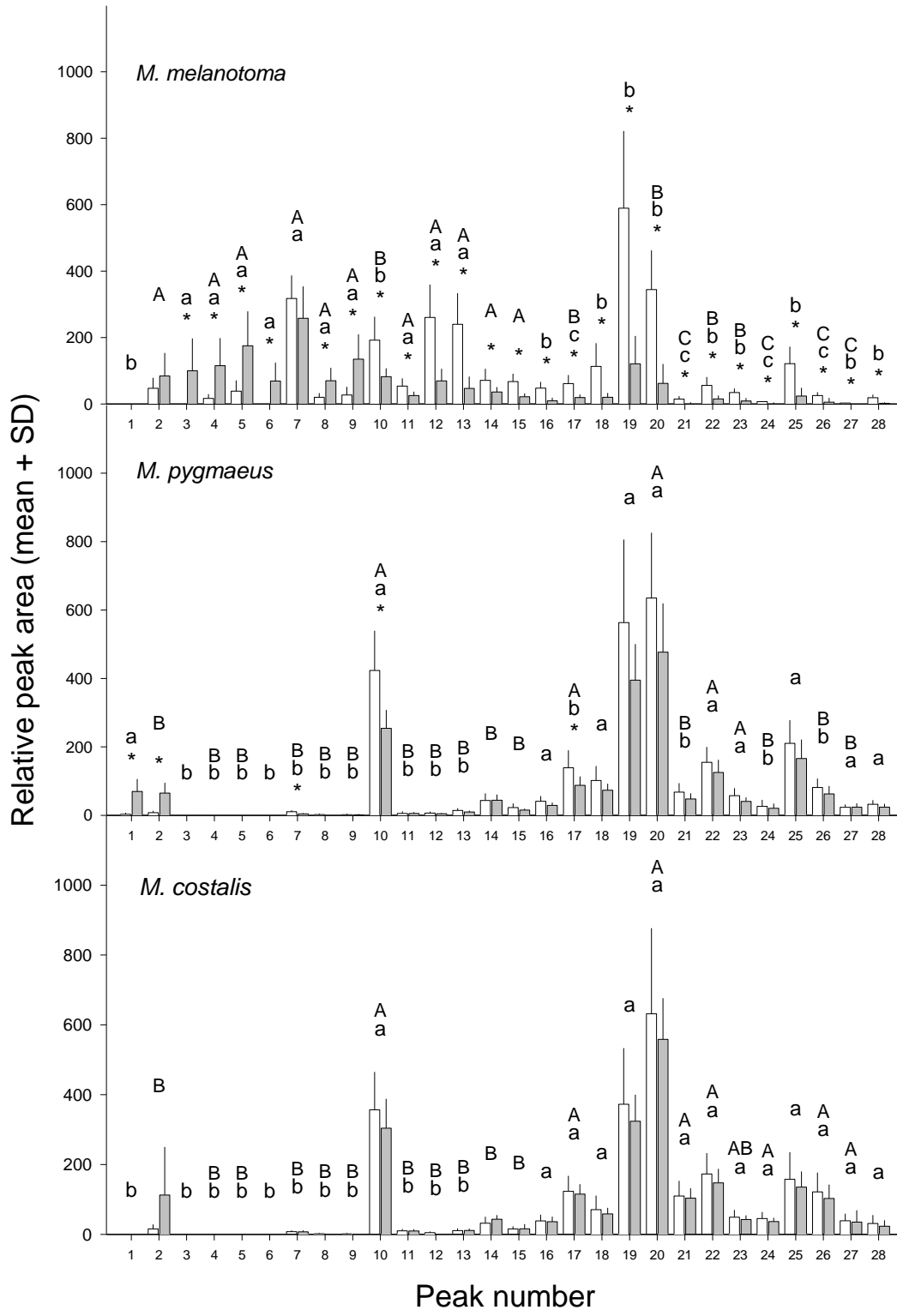


Figure3

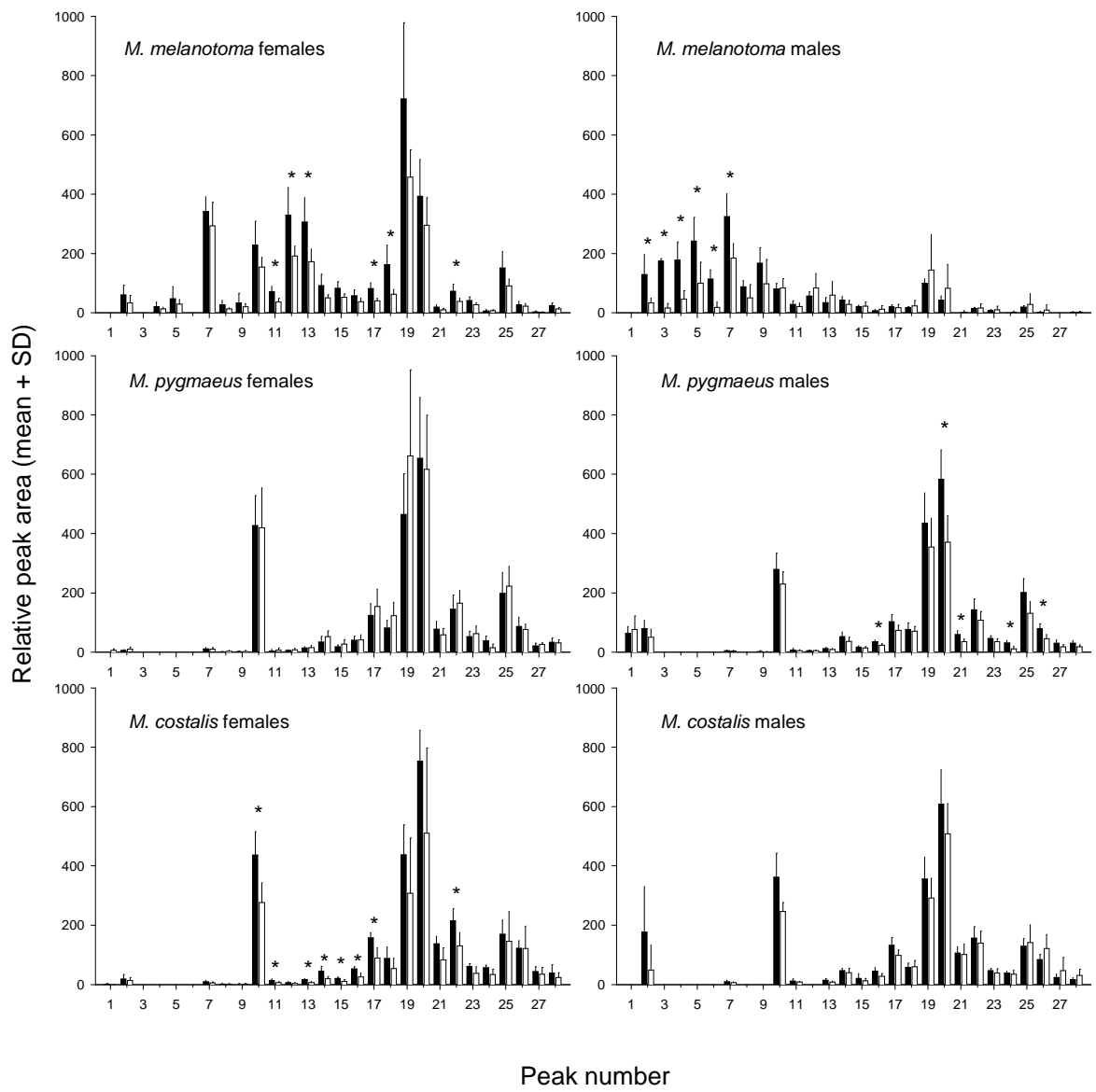
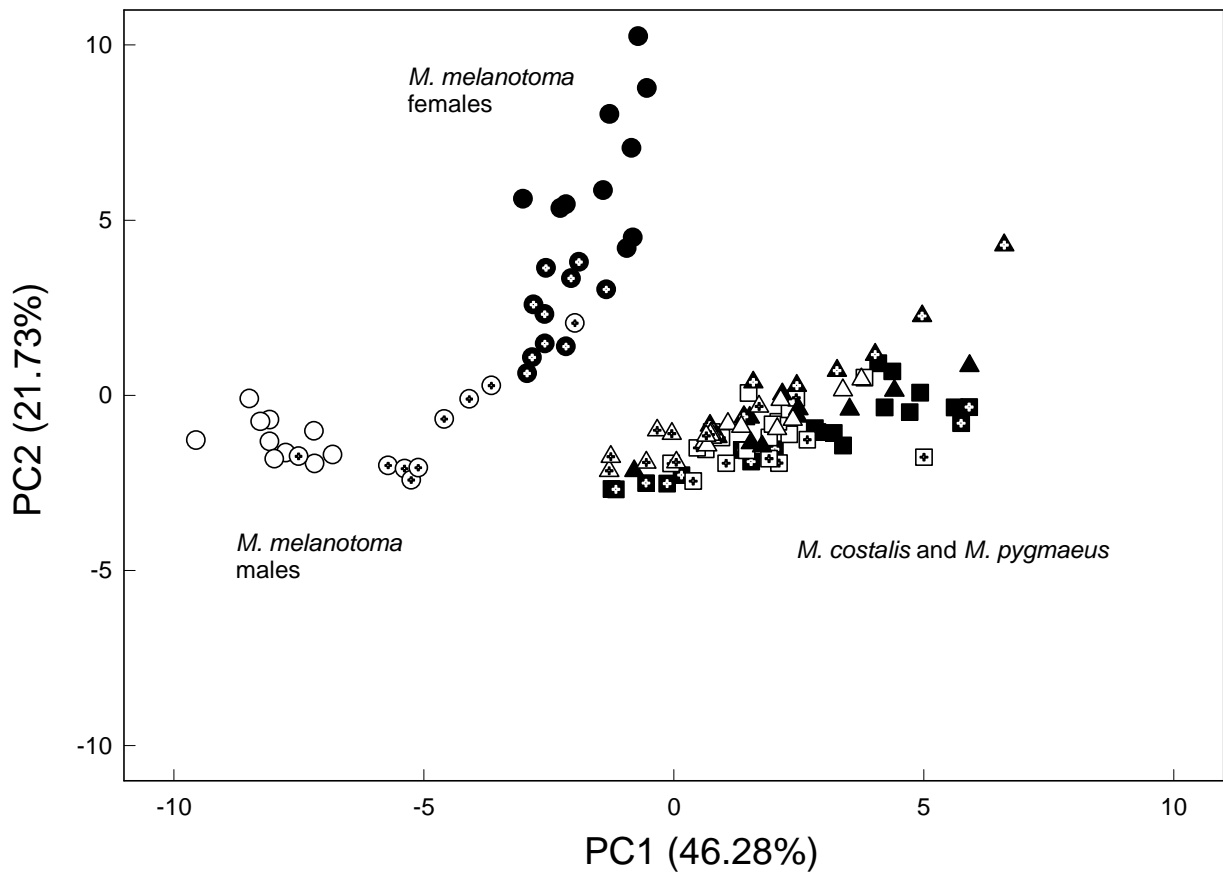


Figure4



Supplementary Material for

Cuticular hydrocarbons discriminate isomorphic *Macrolophus* species (Heteroptera: Miridae)

**César Gemenó^{1*}, Nerea Laserna¹, Magí Riba², Joan Valls³, Cristina Castañé⁴
and Oscar Alomar⁴**

¹ *Department of Crop and Forest Science, University of Lleida, 25198 Lleida, Spain*

² *Department of Chemistry, University of Lleida, 25198 Lleida, Spain*

³ *Biostatistics Unit. Biomedical Research Institute (IRBLLEIDA), 25198 Lleida, Spain*

⁴ *IRTA Cabrils, 08348 Barcelona, Spain*

1. Supplementary Tables

Males Predicted species by QDA	True species			Total
	<i>M. melanotoma</i>	<i>M. pygmaeus</i>	<i>M. costalis</i>	
<i>M. melanotoma</i>	19	0	0	19
<i>M. pygmaeus</i>	0	20	0	20
<i>M. costalis</i>	0	1	19	20
Total	19	21	19	59
Prediction error (%)	0	4.76	0	1.7

Table S1. Prediction errors in the classification of *Macrolophus* species based on a quadratic discriminant analysis (QDA) using the three main principal components of the cuticular hydrocarbons of males. The number in each cell represents the number of individuals of a true species (column) assigned to the three predicted species (rows). A total 59 insects was analysed.

Females Predicted species by QDA	True species			Total
	<i>M. melanotoma</i>	<i>M. pygmaeus</i>	<i>M. costalis</i>	
<i>M. melanotoma</i>	20	0	0	20
<i>M. pygmaeus</i>	0	20	1	20
<i>M. costalis</i>	0	0	20	20
Total	20	20	21	60
Prediction error (%)	0	0	4.76	1.7

Table S2. Prediction errors in the classification of *Macrolophus* species based on a quadratic discriminant analysis (QDA) using the three main principal components of the cuticular hydrocarbons of females. The number in each cell represents the number of individuals of a true species (column) assigned to the three predicted species (rows). A total 60 insects was analysed.

2. Functions for *Macrolophus* cuticular hydrocarbon QDA computation with R

Three files are available as supplementary material, as described below:

- 1) **S2.Macrolophus.data.TXT** Contains the relative areas of the 28 cuticular hydrocarbons from 119 insects corresponding to three species: *M. melatonoma* (Me), *M. pygmaeus* (Py), and *M. Costalis* (Co), two sexes (m, f), and two origins: laboratory (L) and field (F), analyzed in the main paper.
- 2) **S3.Macrolophus.QDA.R** Contains the script (in R code) that allows reproducing the results shown in the main paper.
- 3) **S4.Macrolophus.predict.R** Contains the code (in R) of the function predict.QDA(), which allows to compute the predicted probabilities for new individuals, using our data as basis.

To reproduce the results shown in the main paper in R, the script (**S3.Macrolophus.QDA.R**) has to be executed using the datafile provided “**S2.Macrolophus.data.TXT**”. This script can also be used to predict the species for a set of new individuals. Below we show, step by step, how we obtained the results of the main paper with R code. We also analyze males and females separately. In addition we analyze a male and a female *Macrolophus sp.* collected on tomato plants in Cabrils (Barcelona, Spain), by using the script contained in the file “**S4.Macrolophus.predict.R**”.

2.1 First, the data has to be read, using the data file “S2.Macrolophus.data.TXT”

```
> #####
> # 1 Data reading #
> #####

> macrolophus <- read.table("S2.Macrolophus.data.TXT", sep="\t", header=T)
> summary(macrolophus)
species sex origin p1 p2 p3 p4
Me:39 f:60 F:59 Min. : 0.000 Min. : 1.63 Min. : 0.00 Min. : 0.00
Co:40 m:59 L:60 1st Qu.: 0.000 1st Qu.: 11.42 1st Qu.: 0.00 1st Qu.: 0.00
Py:40 Median : 0.000 Median : 29.20 Median : 0.00 Median : 0.00
Mean : 12.320 Mean : 55.08 Mean : 15.91 Mean : 21.17
3rd Qu.: 2.005 3rd Qu.: 74.72 3rd Qu.: 0.00 3rd Qu.: 11.88
Max. :150.710 Max. :547.65 Max. :318.30 Max. :259.51

p5 p6 p7 p8 p9
Min. : 0.00 Min. : 0.00 Min. : 0.00 Min. : 0.00 Min. : 0.00
1st Qu.: 0.00 1st Qu.: 0.00 1st Qu.: 5.73 1st Qu.: 0.00 1st Qu.: 0.00
Median : 0.00 Median : 0.00 Median : 9.29 Median : 1.24 Median : 1.35
Mean : 34.44 Mean : 10.99 Mean : 99.57 Mean : 15.14 Mean : 26.82
3rd Qu.: 25.82 3rd Qu.: 0.00 3rd Qu.:235.72 3rd Qu.: 14.39 3rd Qu.: 17.43
Max. :393.66 Max. :157.52 Max. :476.77 Max. :157.47 Max. :294.53

p10 p11 p12 p13 p14
Min. : 52.81 Min. : 0.00 Min. : 0.00 Min. : 2.460 Min. : 0.00
1st Qu.:172.97 1st Qu.: 4.93 1st Qu.: 2.64 1st Qu.: 9.275 1st Qu.: 30.93
Median :252.68 Median : 9.60 Median : 5.64 Median : 15.670 Median : 42.01
Mean :270.35 Mean :18.13 Mean : 57.47 Mean : 55.422 Mean : 44.99
3rd Qu.:355.94 3rd Qu.:23.34 3rd Qu.: 54.49 3rd Qu.: 30.025 3rd Qu.: 55.42
Max. :677.64 Max. :91.32 Max. :483.56 Max. :419.810 Max. :131.52

p15 p16 p17 p18 p19
Min. : 0.00 Min. : 2.62 Min. : 4.73 Min. : 6.86 Min. : 48.15
1st Qu.: 13.29 1st Qu.:18.12 1st Qu.: 52.55 1st Qu.: 45.41 1st Qu.: 262.06
Median : 18.85 Median :32.79 Median : 91.42 Median : 64.00 Median : 377.50
Mean : 26.40 Mean :33.88 Mean : 91.38 Mean : 73.33 Mean : 396.46
3rd Qu.: 26.71 3rd Qu.:47.68 3rd Qu.:129.88 3rd Qu.: 92.42 3rd Qu.: 479.30
Max. :126.30 Max. :86.88 Max. :288.27 Max. :277.26 Max. :1374.73

p20 p21 p22 p23 p24
Min. : 24.01 Min. : 0.00 Min. : 3.92 Min. : 2.07 Min. : 0.000
1st Qu.: 285.81 1st Qu.: 16.29 1st Qu.: 53.40 1st Qu.: 24.36 1st Qu.: 5.505
Median : 467.93 Median : 49.50 Median :118.32 Median : 40.79 Median :19.320
Mean : 454.72 Mean : 57.84 Mean :112.74 Mean : 39.03 Mean :22.953
3rd Qu.: 634.28 3rd Qu.: 88.03 3rd Qu.:156.94 3rd Qu.: 50.22 3rd Qu.:36.525
Max. :1140.66 Max. :176.75 Max. :283.98 Max. :112.66 Max. :69.310

p25 p26 p27 p28
Min. : 5.98 Min. : 0.00 Min. : 0.00 Min. : 0.00
```

```

1st Qu.: 85.47    1st Qu.: 24.89    1st Qu.: 1.70    1st Qu.: 11.93
Median :131.88   Median : 63.68   Median : 20.09   Median : 20.32
Mean   :136.66   Mean   : 66.90   Mean   : 20.85   Mean   : 21.96
3rd Qu.:179.68   3rd Qu.: 96.83   3rd Qu.: 30.48   3rd Qu.: 28.04
Max.   :354.38   Max.   :273.62   Max.   :167.80   Max.   :117.71
> dim(macrolophus)
[1] 119 31
> head(macrolophus)
  species sex origin p1    p2 p3    p4    p5 p6    p7    p8    p9    p10   p11   p12   p13
1      Me   f     L  0  14.45  0  13.67 35.07  0 283.19 21.36 19.44 147.78 61.97 342.83 272.28
2      Me   f     L  0  26.25  0  4.90 10.70  0 285.29  8.70  7.17 217.82 41.70 227.25 269.16
3      Me   f     L  0  22.27  0  7.21 22.22  0 354.04 12.90 13.83 211.20 83.76 249.88 419.81
4      Me   f     L  0 112.70  0 35.17 66.40  0 291.57 48.99 26.85 136.85 79.55 381.80 341.69
5      Me   f     L  0  83.21  0 41.98 79.83  0 426.06 44.55 61.78 287.90 66.09 422.08 259.76
6      Me   f     L  0  59.55  0 27.08 62.57  0 407.93 25.29 43.05 147.60 91.32 289.20 321.04
      p14   p15   p16   p17   p18   p19   p20   p21   p22   p23   p24   p25   p26   p27   p28
1  99.85  75.50 45.42  57.17 111.53 590.21 311.86  9.65 62.45 32.59  7.02  98.23 16.11  1.70 11.86
2  74.01  66.68 52.13  59.52 136.79 634.74 395.99 28.36 71.38 39.22  6.80 157.56 30.79  4.69 25.41
3  68.35  70.34 76.38  66.94  58.73 428.78 449.71 23.21 33.47 27.32  8.73  76.08 25.21  0.82 14.48
4 128.09 103.03 73.26  88.76 145.39 831.35 474.16 34.49 80.34 49.89 10.00 171.57 29.66  2.92 25.51
5 131.52  97.00 50.55 103.16 277.26 1090.02 429.99  8.62 93.84 54.69  7.25 239.37 48.04  4.74 34.73
6  99.89  64.28 53.26  69.80  88.64 462.01 287.52 18.29 54.26 26.29  5.21  85.66 17.36  1.60 13.18

```

2.2 Next, the code below can be used to perform the principal component analysis.

```

> #####
> # 2 Principal components analysis #
> #####

> library(MASS)
> macrolophus.pca <- princomp(macrolophus[,4:31],cor=T)
> summary(macrolophus.pca)
Importance of components:

Standard deviation   3.5997770  2.4667929  1.6782470  1.14622797  1.04554207  0.89193104  0.72343036
Proportion of Variance 0.4627998  0.2173238  0.1005897  0.04692281  0.03904136  0.02841218  0.01869112
Cumulative Proportion 0.4627998  0.6801236  0.7807134  0.82763620  0.86667756  0.89508974  0.91378086

Comp.8      Comp.9      Comp.10     Comp.11     Comp.12     Comp.13
Standard deviation  0.66406272  0.65833720  0.6079244  0.46722859  0.426682075  0.365307627
Proportion of Variance 0.01574926  0.01547885  0.0131990  0.00779652  0.006502057  0.004766059
Cumulative Proportion 0.92953012  0.94500898  0.9582080  0.96600450  0.972506554  0.977272613

Comp.14     Comp.15     Comp.16     Comp.17     Comp.18     Comp.19
Standard deviation  0.336951113  0.301806189  0.287428171  0.267444218  0.251891725  0.228047493
Proportion of Variance 0.004054859  0.003253106  0.002950534  0.002554515  0.002266051  0.001857345
Cumulative Proportion 0.981327472  0.984580579  0.987531113  0.990085627  0.992351679  0.994209024

Comp.20     Comp.21     Comp.22     Comp.23     Comp.24     Comp.25
Standard deviation  0.200466700  0.188514348  0.1508854859  0.1484273877  0.1166391642  0.0933369246
Proportion of Variance 0.001435246  0.001269202  0.0008130868  0.0007868103  0.0004858820  0.0003111351
Cumulative Proportion 0.995644270  0.996913472  0.9977265590  0.9985133694  0.9989992513  0.9993103864

Comp.26     Comp.27     Comp.28
Standard deviation  0.0920458179  0.0809715804  0.0654244002
Proportion of Variance 0.0003025869  0.0002341570  0.0001528697
Cumulative Proportion 0.9996129733  0.9998471303  1.0000000000
> macrolophus.pca
Call:
princomp(x = macrolophus[, 4:31], cor = T)

Standard deviations:
  Comp.1   Comp.2   Comp.3   Comp.4   Comp.5   Comp.6   Comp.7   Comp.8   Comp.9
3.59977704 2.46679290 1.67824699 1.14622797 1.04554207 0.89193104 0.72343036 0.66406272 0.65833720
  Comp.10  Comp.11  Comp.12  Comp.13  Comp.14  Comp.15  Comp.16  Comp.17  Comp.18
0.60792436 0.46722859 0.42668208 0.36530763 0.33695111 0.30180619 0.28742817 0.26744422 0.25189173
  Comp.19  Comp.20  Comp.21  Comp.22  Comp.23  Comp.24  Comp.25  Comp.26  Comp.27
0.22804749 0.20046670 0.18851435 0.15088549 0.14842739 0.11663916 0.09333692 0.09204582 0.08097158
  Comp.28
0.06542440

28 variables and 119 observations.

```

2.3 Quadratic discriminant analysis is then applied to the first three principal components.

```

> #####
> # 3 Quadratic discriminant analysis #
> #####

> macrolophus$PC1 <- macrolophus.pca$scores[,1]
> macrolophus$PC2 <- macrolophus.pca$scores[,2]
> macrolophus$PC3 <- macrolophus.pca$scores[,3]
> macrolophus.qda <- qda(species~PC1+PC2+PC3,data=macrolophus)
> macrolophus.qda
Call:
qda(species ~ PC1 + PC2 + PC3, data = macrolophus)

```



```

67 2.029885e-27 1.000000e+00 5.895113e-15
68 6.929522e-35 9.952248e-01 4.775237e-03
69 3.708216e-24 1.000000e+00 2.527683e-30
70 1.437963e-88 1.000000e+00 1.754505e-08
71 2.382960e-36 7.018986e-01 2.981014e-01
72 1.256957e-40 9.946113e-01 5.388716e-03
73 1.705737e-22 1.000000e+00 3.943261e-11
74 8.435407e-17 4.496036e-01 5.503964e-01
75 9.077042e-44 7.998866e-01 2.001134e-01
76 4.778201e-38 9.981186e-01 1.881379e-03
77 7.973526e-28 9.566672e-01 4.333277e-02
78 1.025495e-23 4.383259e-01 5.616741e-01
79 2.659344e-22 8.470499e-01 1.529501e-01
80 3.705368e-38 1.769242e-01 8.230758e-01
81 1.010249e-52 8.249005e-01 1.750995e-01
82 5.759876e-85 5.612043e-01 4.387957e-01
83 9.417009e-27 2.606889e-02 9.739311e-01
84 2.647101e-37 7.191369e-02 9.280863e-01
85 8.396146e-62 3.318818e-01 6.681182e-01
86 6.991704e-32 3.233243e-02 9.676676e-01
87 1.114155e-11 1.428729e-01 8.571271e-01
88 4.022769e-32 3.385770e-01 6.614230e-01
89 4.779310e-30 3.028359e-01 6.971641e-01
90 3.111415e-42 3.854323e-03 9.961457e-01
91 1.171852e-23 1.657259e-02 9.834274e-01
92 5.539828e-50 1.182253e-03 9.988177e-01
93 9.494014e-20 2.341961e-02 9.765804e-01
94 9.529373e-59 4.491360e-05 9.999551e-01
95 4.270801e-72 4.650132e-09 1.000000e+00
96 1.803989e-26 1.047406e-01 8.952594e-01
97 4.936870e-34 8.475899e-03 9.915241e-01
98 1.408292e-25 2.692238e-02 9.730776e-01
99 3.773500e-22 4.667346e-02 9.533265e-01
100 1.117377e-19 3.564483e-02 9.643552e-01
101 2.813980e-32 5.480513e-02 9.451949e-01
102 3.803048e-37 1.518908e-01 8.481092e-01
103 1.613566e-34 2.271748e-01 7.728252e-01
104 4.896069e-50 1.984771e-02 9.801523e-01
105 5.417494e-47 6.799428e-02 9.320057e-01
106 1.376361e-21 1.002046e-01 8.997954e-01
107 1.551351e-23 3.468337e-01 6.531663e-01
108 3.035439e-21 9.710385e-02 9.028962e-01
109 1.807461e-26 1.511359e-01 8.488641e-01
110 1.650914e-11 3.619455e-02 9.638055e-01
111 2.187144e-08 4.318655e-02 9.568134e-01
112 1.198214e-07 4.263977e-02 9.573601e-01
113 6.485381e-20 2.052001e-02 9.794800e-01
114 3.303492e-14 1.771733e-01 8.228267e-01
115 2.671738e-11 3.316938e-02 9.668306e-01
116 6.112121e-18 4.239882e-02 9.576012e-01
117 4.653508e-14 2.692003e-02 9.730800e-01
118 1.015974e-19 2.691500e-02 9.730850e-01
119 1.355224e-25 8.503637e-03 9.914964e-01

```

```

> macrolophus$QDA.prob.Me <- macrolophus.predict$posterior[,1]
> macrolophus$QDA.prob.Co <- macrolophus.predict$posterior[,2]
> macrolophus$QDA.prob.Py <- macrolophus.predict$posterior[,3]
> macrolophus$QDA.predicted.species <- macrolophus.predict$class

```

The following data frame contains the predictions for the insects in the dataset.

```

> results <- macrolophus[,c("species", "QDA.prob.Me", "QDA.prob.Co", "QDA.prob.Py", "QDA.predicted.species")]
> results[,2:4] <- round(results[,2:4],2)
> results
  species QDA.prob.Me QDA.prob.Co QDA.prob.Py QDA.predicted.species
1      Me           1           0.00         0.00                Me
2      Me           1           0.00         0.00                Me
3      Me           1           0.00         0.00                Me
4      Me           1           0.00         0.00                Me
5      Me           1           0.00         0.00                Me
6      Me           1           0.00         0.00                Me
7      Me           1           0.00         0.00                Me
8      Me           1           0.00         0.00                Me
9      Me           1           0.00         0.00                Me
10     Me           1           0.00         0.00                Me
11     Me           1           0.00         0.00                Me
12     Me           1           0.00         0.00                Me
13     Me           1           0.00         0.00                Me
14     Me           1           0.00         0.00                Me
15     Me           1           0.00         0.00                Me
16     Me           1           0.00         0.00                Me
17     Me           1           0.00         0.00                Me
18     Me           1           0.00         0.00                Me
19     Me           1           0.00         0.00                Me
20     Me           1           0.00         0.00                Me
21     Me           1           0.00         0.00                Me
22     Me           1           0.00         0.00                Me

```

23	Me	1	0.00	0.00	Me
24	Me	1	0.00	0.00	Me
25	Me	1	0.00	0.00	Me
26	Me	1	0.00	0.00	Me
27	Me	1	0.00	0.00	Me
28	Me	1	0.00	0.00	Me
29	Me	1	0.00	0.00	Me
30	Me	1	0.00	0.00	Me
31	Me	1	0.00	0.00	Me
32	Me	1	0.00	0.00	Me
33	Me	1	0.00	0.00	Me
34	Me	1	0.00	0.00	Me
35	Me	1	0.00	0.00	Me
36	Me	1	0.00	0.00	Me
37	Me	1	0.00	0.00	Me
38	Me	1	0.00	0.00	Me
39	Me	1	0.00	0.00	Me
40	Co	0	0.31	0.69	Py
41	Co	0	0.97	0.03	Co
42	Co	0	0.28	0.72	Py
43	Co	0	0.95	0.05	Co
44	Co	0	1.00	0.00	Co
45	Co	0	0.99	0.01	Co
46	Co	0	0.95	0.05	Co
47	Co	0	0.99	0.01	Co
48	Co	0	1.00	0.00	Co
49	Co	0	0.98	0.02	Co
50	Co	0	1.00	0.00	Co
51	Co	0	0.83	0.17	Co
52	Co	0	0.78	0.22	Co
53	Co	0	1.00	0.00	Co
54	Co	0	0.49	0.51	Py
55	Co	0	0.95	0.05	Co
56	Co	0	0.84	0.16	Co
57	Co	0	0.70	0.30	Co
58	Co	0	0.59	0.41	Co
59	Co	0	0.74	0.26	Co
60	Co	0	1.00	0.00	Co
61	Co	0	1.00	0.00	Co
62	Co	0	1.00	0.00	Co
63	Co	0	0.89	0.11	Co
64	Co	0	0.96	0.04	Co
65	Co	0	0.94	0.06	Co
66	Co	0	0.92	0.08	Co
67	Co	0	1.00	0.00	Co
68	Co	0	1.00	0.00	Co
69	Co	0	1.00	0.00	Co
70	Co	0	1.00	0.00	Co
71	Co	0	0.70	0.30	Co
72	Co	0	0.99	0.01	Co
73	Co	0	1.00	0.00	Co
74	Co	0	0.45	0.55	Py
75	Co	0	0.80	0.20	Co
76	Co	0	1.00	0.00	Co
77	Co	0	0.96	0.04	Co
78	Co	0	0.44	0.56	Py
79	Co	0	0.85	0.15	Co
80	Py	0	0.18	0.82	Py
81	Py	0	0.82	0.18	Co
82	Py	0	0.56	0.44	Co
83	Py	0	0.03	0.97	Py
84	Py	0	0.07	0.93	Py
85	Py	0	0.33	0.67	Py
86	Py	0	0.03	0.97	Py
87	Py	0	0.14	0.86	Py
88	Py	0	0.34	0.66	Py
89	Py	0	0.30	0.70	Py
90	Py	0	0.00	1.00	Py
91	Py	0	0.02	0.98	Py
92	Py	0	0.00	1.00	Py
93	Py	0	0.02	0.98	Py
94	Py	0	0.00	1.00	Py
95	Py	0	0.00	1.00	Py
96	Py	0	0.10	0.90	Py
97	Py	0	0.01	0.99	Py
98	Py	0	0.03	0.97	Py
99	Py	0	0.05	0.95	Py
100	Py	0	0.04	0.96	Py
101	Py	0	0.05	0.95	Py
102	Py	0	0.15	0.85	Py
103	Py	0	0.23	0.77	Py
104	Py	0	0.02	0.98	Py
105	Py	0	0.07	0.93	Py
106	Py	0	0.10	0.90	Py
107	Py	0	0.35	0.65	Py
108	Py	0	0.10	0.90	Py
109	Py	0	0.15	0.85	Py
110	Py	0	0.04	0.96	Py

111	Py	0	0.04	0.96	Py
112	Py	0	0.04	0.96	Py
113	Py	0	0.02	0.98	Py
114	Py	0	0.18	0.82	Py
115	Py	0	0.03	0.97	Py
116	Py	0	0.04	0.96	Py
117	Py	0	0.03	0.97	Py
118	Py	0	0.03	0.97	Py
119	Py	0	0.01	0.99	Py

The table below compares the true species with the predicted ones. An error of 5.88% is obtained.

```
>
table(macrolophus$species,macrolophus$QDA.predicted.species,dnn=c("real","predicted"))
> correct.prediction
      predicted
real Me Co Py
Me 39  0  0
Co  0 35  5
Py  0  2 38
> prop.table(correct.prediction,1)
      predicted
real  Me    Co    Py
Me 1.000 0.000 0.000
Co 0.000 0.875 0.125
Py 0.000 0.050 0.950
> prop.table(correct.prediction,2)
      predicted
real  Me    Co    Py
Me 1.00000000 0.00000000 0.00000000
Co 0.00000000 0.94594595 0.11627907
Py 0.00000000 0.05405405 0.88372093
>
> total.correct <- sum(diag(correct.prediction))/sum(correct.prediction)
> total.correct
[1] 0.9411765

> total.error <- 1-total.correct
> total.error
[1] 0.05882353
```

2.4 Finally, sections 5.1 and 5.2 in the script (“S3.Macrolophus.QDA.R”) contain the code for performing the same analysis above but for each sex individually. The result is shown in supplementary tables 1 and 2 above.

2.5 Here we illustrate the prediction for two *Macrolophus sp.* individuals, one of each sex, of unknown species status, which were collected on tomato plants (“tomatomale” and “tomatofemale”).

```
> #####
> # 4 Prediction for new individuals #
> #####

> library(mnormt)
> tomatomale <- c(9.3359225,35.24040853,0,0,0,2.620230036,4.143500323,0,227.0959237,6.803160605,
1.675141163,5.174382195,27.78788589,33.50457571,21.56733939,80.30059764,62.56136815,290.5071812,451.0842287,
81.21811111,161.8425264,35.04168524,11.66117306,149.6672654,57.47777861,23.24498038,20.68211748)
> tomatofemale <- c(0.465840529,33.08057566,0,1.69478331,0,2.434254523,13.35527135,3.219487532,0.805361245,
623.4437673,21.48072989,5.886884465,11.75026023,17.20756204,32.65718085,44.08958108,133.3946882,100.2624269,6
68.2018557,775.8341467,86.4138592,143.781386,65.40976156,43.59786875,237.3767564,101.9367407,24.22607705,38.8
2679428)
> macrolophus.new <- rbind(tomatomale,tomatofemale)
> source("S4.Macrolophus.predict.R")
> predict.QDA(macrolophus.pca,macrolophus.new,macrolophus,macrolophus$species)
      prob.Ca.cond.X.star prob.Co.cond.X.star prob.Py.cond.X.star
tomatomale                0                0.335                0.665
tomatofemale              0                0.074                0.926
```

The results clearly show that the female can be assigned to *M. pygmaeus* (with a probability of 0.92). However, the prediction for the male is less clear. Although it is not a *M. melatonoma* (probability 0), it could be either a *M. costalis* (probability 0.075) or *M. pygmaeus* (probability 0.665). Because the male lacked the characteristic dark spot on the scutellum, it was classified as *M. pygmaeus*. So the model demonstrated that the two insects collected on tomato belong to this last species.

species	sex	origin	p1	p2	p3	p4	p5	p6	p7	p8	p9	p10
	p11	p12	p13	p14	p15	p16	p17	p18	p19	p20	p21	p22
	p23	p24	p25	p26	p27	p28						
Me	f	L	0.00	14.45	0.00	13.67	35.07	0.00	283.19	21.36	19.44	
	147.78	61.97	342.83	272.28	99.85	75.50	45.42	57.17	111.53	590.21	311.86	9.65
	62.45	32.59	7.02	98.23	16.11	1.70	11.86					
Me	f	L	0.00	26.25	0.00	4.90	10.70	0.00	285.29	8.70	7.17	
	217.82	41.70	227.25	269.16	74.01	66.68	52.13	59.52	136.79	634.74	395.99	28.36
	71.38	39.22	6.80	157.56	30.79	4.69	25.41					
Me	f	L	0.00	22.27	0.00	7.21	22.22	0.00	354.04	12.90	13.83	
	211.20	83.76	249.88	419.81	68.35	70.34	76.38	66.94	58.73	428.78	449.71	23.21
	33.47	27.32	8.73	76.08	25.21	0.82	14.48					
Me	f	L	0.00	112.70	0.00	35.17	66.40	0.00	291.57	48.99	26.85	
	136.85	79.55	381.80	341.69	128.09	103.03	73.26	88.76	145.39	831.35	474.16	34.49
	80.34	49.89	10.00	171.57	29.66	2.92	25.51					
Me	f	L	0.00	83.21	0.00	41.98	79.83	0.00	426.06	44.55	61.78	
	287.90	66.09	422.08	259.76	131.52	97.00	50.55	103.16	277.26	1090.02		
	429.99	8.62	93.84	54.69	7.25	239.37	48.04	4.74	34.73			
Me	f	L	0.00	59.55	0.00	27.08	62.57	0.00	407.93	25.29	43.05	
	147.60	91.32	289.20	321.04	99.89	64.28	53.26	69.80	88.64	462.01	287.52	18.29
	54.26	26.29	5.21	85.66	17.36	1.60	13.18					
Me	f	L	0.00	78.24	0.00	11.94	29.35	0.00	350.30	34.72	63.41	
	328.09	87.65	403.34	358.25	0.00	99.90	71.20	106.12	195.55	887.85	475.61	23.58
	89.63	49.39	7.49	175.05	33.25	4.30	22.37					
Me	f	L	0.00	60.74	0.00	48.21	143.60	0.00	304.93	40.71	97.21	
	171.24	88.38	483.56	412.75	121.16	126.30	86.88	107.93	192.98	1154.90		
	624.05	17.25	96.79	59.61	11.09	225.98	41.94	4.61	39.05			
Me	f	L	0.00	59.74	0.00	5.96	12.66	0.00	356.84	10.31	0.00	
	311.84	45.79	192.94	150.29	107.10	51.43	26.15	68.99	207.29	514.11	197.88	9.13
	102.20	36.93	0.00	142.65	20.27	8.66	35.32					
Me	f	L	0.00	89.47	0.00	4.47	12.12	0.00	353.91	19.62	6.70	
	333.17	66.03	303.83	264.59	84.85	70.81	40.99	84.37	215.15	619.46	283.57	19.14
	44.50	38.28	0.00	139.39	15.95	0.00	18.66					
Me	f	F	0.00	45.78	0.00	17.73	27.74	0.00	361.18	16.36	19.56	
	195.88	46.49	214.43	209.15	55.64	68.24	51.32	46.49	80.08	621.04	419.46	13.72
	37.60	32.72	6.40	122.97	29.12	1.52	16.67					
Me	f	F	0.00	13.92	0.00	9.04	17.65	0.00	189.65	11.71	15.27	
	128.94	31.78	150.95	151.25	42.47	44.00	33.94	40.18	59.86	373.44	284.39	16.97
	38.91	29.57	10.27	85.28	24.57	1.78	10.35					
Me	f	F	0.00	21.06	0.00	19.55	63.33	0.00	271.98	16.49	46.79	
	136.53	62.16	245.48	229.28	64.45	65.08	48.01	55.16	52.58	460.07	329.52	12.01
	50.00	34.73	11.92	81.96	25.73	1.70	9.48					
Me	f	F	0.00	28.40	0.00	17.82	35.95	0.00	246.09	14.72	17.74	
	121.76	28.35	144.47	132.55	41.50	40.02	25.73	26.12	52.34	368.55	229.88	7.47
	28.22	19.44	4.06	81.04	17.30	1.44	14.59					
Me	f	F	0.00	13.33	0.00	14.28	30.97	0.00	249.04	14.80	23.11	
	157.94	35.64	178.32	212.80	55.70	61.94	54.72	49.77	64.82	534.58	461.10	19.99
	54.09	37.29	11.29	123.75	41.00	5.89	20.76					
Me	f	F	0.00	14.26	0.00	10.33	19.35	0.00	235.02	8.86	12.66	
	119.60	23.25	184.30	162.62	33.03	52.67	36.17	32.81	44.52	398.05	254.53	6.27
	27.25	22.48	6.04	72.24	19.44	1.54	11.19					

Me	f	F	0.00	13.07	0.00	12.93	32.10	0.00	258.68	13.22	18.93			
			125.76	36.98	211.37	173.17	48.78	55.95	38.49	40.00	48.93	461.17	289.80	5.95
			31.71	24.20	5.80	78.10	22.88	1.46	12.00					
Me	f	F	0.00	30.29	0.00	15.38	37.20	0.00	380.60	16.64	27.48			
			172.18	35.46	194.73	190.71	59.51	48.72	35.41	36.82	71.50	440.15	266.52	8.47
			37.40	21.82	3.82	85.20	15.67	1.50	10.74					
Me	f	F	0.00	75.10	0.00	8.57	23.90	0.00	451.20	12.42	12.53			
			212.83	38.53	229.36	175.35	61.12	52.51	35.77	47.29	94.49	575.30	287.22	5.61
			52.71	34.67	9.62	119.64	23.45	1.95	17.43					
Me	f	F	0.00	77.92	0.00	3.68	7.36	0.00	295.29	5.17	5.66			
			173.75	22.16	158.78	86.07	42.58	32.24	15.05	31.83	59.40	341.88	134.97	0.00
			28.90	17.07	3.02	61.22	9.14	1.12	8.76					
Me	m	L	0.00	131.25	189.04	177.01	281.07	80.71	306.90	90.25	193.02	89.89		
			51.46	70.39	49.25	36.37	23.84	10.46	31.25	14.23	106.41	50.89	0.00	9.11
			5.84	0.00	17.22	0.00	0.00	4.27						
Me	m	L	0.00	96.12	178.61	151.41	334.51	157.52	367.91	75.48	236.28	96.12		
			42.79	82.85	75.53	68.57	31.58	14.93	28.15	15.14	113.42	71.39	0.00	20.53
			11.00	0.00	20.23	6.41	0.00	0.00						
Me	m	L	0.00	171.27	172.28	200.78	235.16	97.48	289.94	109.00	180.10	69.31		
			24.65	52.82	29.46	46.90	20.51	6.26	17.55	16.60	79.93	37.90	0.00	14.48
			7.83	0.00	24.04	0.00	0.00	5.03						
Me	m	L	0.00	117.25	211.30	259.51	156.99	105.95	288.27	96.89	90.89	97.27		
			16.76	37.49	20.19	48.04	18.85	4.18	16.82	26.19	100.70	28.07	0.00	17.19
			6.05	0.00	19.44	0.00	0.00	0.00						
Me	m	L	0.00	126.69	143.50	190.06	239.21	82.46	476.77	119.53	200.88	92.99		
			36.04	62.10	28.93	40.67	21.14	7.36	21.72	16.12	96.88	40.92	0.00	12.76
			8.38	1.75	17.88	0.00	0.00	3.12						
Me	m	L	0.00	43.69	160.06	134.80	216.29	89.29	274.29	46.36	98.94	57.47		
			17.22	43.54	21.26	35.05	20.00	5.30	14.95	15.71	108.28	33.18	0.00	13.48
			6.67	0.00	16.36	0.00	0.00	2.53						
Me	m	L	0.00	77.15	114.72	123.47	158.48	101.55	250.81	90.32	151.90	74.44		
			17.66	47.79	19.36	25.79	17.04	3.18	12.39	10.61	81.02	29.67	0.00	6.82
			4.49	0.00	11.85	0.00	0.00	0.00						
Me	m	L	0.00	116.81	318.30	245.20	393.66	155.95	421.11	84.12	203.80	55.68		
			25.25	56.17	37.21	40.08	24.59	8.88	22.99	21.50	122.93	60.64	0.00	17.14
			9.21	1.10	29.38	6.73	0.00	0.00						
Me	m	L	0.00	295.56	35.67	58.80	154.98	154.15	240.99	98.38	214.04			
			109.43	34.47	67.82	36.42	52.74	17.36	6.89	25.29	15.49	95.56	39.83	0.00
			16.65	9.76	0.00	19.60	0.00	0.00	0.00					
Me	m	L	0.00	116.79	224.29	231.85	256.51	117.33	329.81	67.69	105.49	63.70		
			16.87	42.53	27.97	34.38	17.02	5.69	16.94	16.40	91.32	34.00	0.00	14.18
			7.53	1.42	17.90	0.00	0.00	3.69						
Me	m	F	0.00	69.94	45.77	66.94	119.85	45.34	180.34	60.61	116.66	65.12		
			10.08	47.78	26.24	19.60	16.78	5.39	9.08	14.40	76.96	40.01	0.00	7.58
			4.63	0.00	17.09	5.07	0.00	0.00						
Me	m	F	0.00	41.84	4.95	12.95	20.65	0.00	181.64	14.07	17.11			
			102.41	33.64	148.21	116.11	37.22	31.93	17.74	22.98	36.01	228.81	116.19	2.46
			20.27	9.87	1.17	29.64	4.62	0.00	2.71					
Me	m	F	0.00	30.64	3.27	11.81	39.06	0.00	236.42	17.08	53.89	87.08		
			32.98	116.11	89.63	39.91	26.60	14.92	24.82	26.39	168.04	86.53	0.00	18.49
			7.31	0.98	19.04	3.70	0.00	2.72						

Me	m	F	0.00	29.20	6.86	61.61	106.17	6.81	186.24	32.63	54.16	67.78		
			31.58	79.48	51.37	41.77	20.93	10.28	24.54	28.79	143.14	71.57	1.78	21.21
			6.81	0.00	21.16	3.66	0.00	3.38						
Me	m	F	0.00	27.40	38.34	73.90	111.85	24.84	148.46	56.67	100.56	85.06		
			10.77	47.89	29.57	16.63	12.94	4.78	6.95	12.81	86.11	45.12	0.00	8.86
			5.51	0.91	14.03	0.00	0.00	0.00						
Me	m	F	0.00	38.34	7.27	36.27	90.75	16.52	109.25	63.21	109.77	62.39		
			10.84	39.16	19.10	11.40	9.90	2.62	4.73	7.53	48.15	24.01	0.00	3.92
			2.07	0.00	5.98	1.03	0.00	0.00						
Me	m	F	0.00	24.28	14.44	88.75	253.53	49.05	180.09	157.47	294.53	77.01		
			15.06	75.05	30.12	21.33	13.28	5.10	9.53	13.03	74.25	42.22	0.00	8.48
			5.10	0.00	20.47	3.50	0.00	0.00						
Me	m	F	0.00	24.41	5.95	19.15	33.46	6.92	278.82	19.89	38.16			
			156.53	32.35	163.41	146.52	47.44	54.82	41.07	40.47	63.41	421.14	283.20	15.60
			50.99	43.93	13.84	122.47	57.41	0.00	9.55					
Me	m	F	0.00	19.45	18.59	44.85	125.74	16.49	154.81	27.93	98.09	52.81		
			17.02	35.22	29.93	18.26	10.39	8.39	9.49	6.86	50.10	40.51	0.00	5.58
			2.53	0.00	6.77	1.91	0.00	0.00						
Co	f	L	0.61	45.07	0.00	0.00	0.00	0.00	10.52	0.00	0.00			
			535.32	23.42	12.30	18.95	59.04	26.83	58.28	173.78	114.28	542.28	789.23	
			108.94	247.15	69.21	55.89	207.22	110.21	31.66	29.37				
Co	f	L	1.16	37.17	0.00	0.00	0.00	0.00	12.66	1.98	1.30			
			503.87	15.37	5.51	18.51	48.53	21.17	66.84	174.82	96.18	477.09	937.31	
			176.75	193.86	68.73	69.31	204.01	152.59	23.15	34.22				
Co	f	L	0.00	22.33	0.00	0.00	0.00	0.00	15.50	0.00	0.00			
			423.31	17.59	7.50	17.32	83.12	24.93	53.93	182.17	172.23	481.50	670.14	
			113.02	235.42	60.63	62.64	190.69	99.29	35.82	25.09				
Co	f	L	1.32	33.26	0.00	0.00	0.00	0.00	7.28	2.87	0.00			
			399.67	13.79	4.30	15.83	37.95	21.62	56.21	137.84	52.56	393.16	727.47	
			134.25	133.98	66.69	59.79	137.18	107.45	37.95	117.71				
Co	f	L	0.00	7.64	0.00	0.00	0.00	0.00	6.21	0.00	0.00			
			560.52	7.00	4.78	13.85	36.32	22.23	49.43	160.52	108.33	630.01	915.62	
			133.12	283.98	74.42	60.23	239.23	146.82	79.65	31.49				
Co	f	L	4.81	6.60	0.00	0.00	0.00	0.00	12.92	4.68	0.00			
			463.37	11.99	6.73	15.86	41.00	19.52	50.00	146.88	99.96	453.57	762.34	
			148.22	241.76	64.75	63.90	201.16	147.77	51.65	48.40				
Co	f	L	2.16	5.20	0.00	0.00	0.00	0.00	9.67	3.93	0.00			
			352.63	11.49	5.79	13.35	36.48	16.99	40.26	151.69	65.24	367.89	618.75	
			106.82	207.56	49.93	45.26	142.17	105.50	47.47	31.91				
Co	f	L	0.00	8.18	0.00	0.00	0.00	0.00	2.65	0.00	0.00			
			327.30	7.82	0.00	9.26	24.95	12.99	40.65	142.39	67.59	400.42	717.26	
			121.41	199.34	51.47	48.77	167.59	151.65	55.86	27.78				
Co	f	L	0.25	11.65	0.00	0.00	0.00	0.00	10.34	5.58	7.68			
			433.30	11.42	8.31	19.93	46.74	21.76	55.28	131.20	51.57	296.07	650.79	
			162.73	211.80	49.36	53.07	90.79	86.97	37.00	17.57				
Co	f	L	0.00	4.37	0.00	0.00	0.00	0.00	8.67	0.62	0.69			
			367.48	15.12	7.28	15.67	34.95	17.20	52.43	170.11	48.13	338.77	746.39	
			165.53	195.08	50.83	52.64	114.49	111.79	35.64	20.87				
Co	f	F	0.64	10.73	0.00	0.00	0.00	0.00	6.20	0.00	0.00			
			359.75	8.88	4.39	10.94	30.70	16.54	40.71	131.44	96.04	581.96	864.46	
			117.18	220.57	73.58	60.57	331.27	273.62	85.88	51.84				

Co	f	F	0.00	22.64	0.00	0.00	0.00	0.00	10.51	0.00	0.00			
			334.19	11.37	3.53	6.84	31.16	11.96	29.48	133.97	42.07	267.35	467.93	88.32
			152.63	33.24	30.57	96.15	88.32	21.88	37.05					
Co	f	F	0.00	7.94	0.00	0.00	0.00	0.00	6.75	2.41	1.98			
			230.37	2.83	1.42	2.46	17.81	5.24	14.41	58.24	33.54	187.53	339.96	55.13
			120.97	24.52	22.39	104.16	96.50	23.48	14.27					
Co	f	F	0.00	9.51	0.00	0.00	0.00	0.00	4.32	5.37	7.38			
			379.04	7.74	3.17	9.29	16.64	20.67	52.18	134.10	129.31	677.28	1140.66	
			175.98	174.47	76.41	67.99	315.45	228.88	56.61	44.15				
Co	f	F	0.00	13.87	0.00	0.00	0.00	0.00	6.04	0.00	0.00			
			201.76	3.29	1.98	3.20	12.30	5.54	13.38	50.59	23.24	133.06	243.87	48.24
			85.90	18.42	16.67	52.25	43.87	11.80	7.66					
Co	f	F	0.00	37.83	0.00	0.00	0.00	0.00	4.57	1.54	3.09			
			307.94	7.60	8.69	8.40	12.46	13.31	27.89	71.94	46.97	303.09	524.29	92.51
			105.14	45.89	42.80	156.51	151.37	27.71	21.66					
Co	f	F	0.00	5.79	0.00	0.00	0.00	0.00	5.13	0.56	0.75			
			283.91	7.39	5.04	8.14	24.75	12.24	27.44	114.35	72.56	361.18	557.27	87.34
			155.11	42.49	32.94	158.26	113.65	52.99	26.64					
Co	f	F	0.00	6.28	0.00	0.00	0.00	0.00	3.96	1.24	1.34			
			231.31	4.85	1.24	3.12	19.98	6.82	18.05	70.13	39.86	228.78	377.25	65.23
			107.76	26.81	23.89	108.36	94.71	27.25	16.57					
Co	f	F	0.00	8.94	0.00	0.00	0.00	0.00	0.00	0.00	2.36	3.85		
			198.72	2.67	1.70	3.08	10.64	6.78	15.26	53.03	32.32	220.30	335.25	47.28
			89.00	23.07	19.32	91.01	77.49	22.61	12.38					
Co	f	F	0.00	3.60	0.00	0.00	0.00	0.00	5.81	3.18	1.66			
			239.83	5.95	0.93	2.90	15.91	4.01	12.59	72.34	25.17	112.59	251.18	48.55
			97.37	13.55	14.94	43.15	45.37	16.18	5.39					
Co	m	L	0.00	166.46	0.00	0.00	0.00	0.00	8.23	0.00	0.00			
			403.56	7.99	0.00	9.65	46.20	0.00	50.55	112.90	63.61	342.09	609.97	
			118.28	143.83	51.66	45.17	152.77	104.51	20.09	20.57				
Co	m	L	0.00	188.90	0.00	0.00	0.00	0.00	4.29	0.00	0.00			
			245.67	1.93	0.00	14.47	46.01	16.23	47.60	116.06	48.61	318.00	671.40	
			134.06	137.34	45.50	43.73	131.88	103.53	24.56	16.06				
Co	m	L	0.00	90.63	0.00	0.00	0.00	0.00	3.91	0.00	0.00			
			305.37	6.74	0.00	7.13	40.04	13.38	27.54	111.33	52.34	305.86	494.92	85.94
			171.29	38.28	34.08	128.61	88.18	41.21	17.38					
Co	m	L	0.00	67.78	0.00	0.00	0.00	0.00	8.25	0.00	0.00			
			440.85	8.85	0.00	9.91	49.17	21.86	33.43	124.43	75.42	390.85	549.32	78.90
			204.54	43.65	35.17	143.04	72.77	32.07	22.01					
Co	m	L	0.00	125.41	0.00	0.00	0.00	0.00	14.85	0.00	0.00			
			492.19	13.64	0.00	18.04	45.43	0.00	65.35	173.27	62.16	440.92	735.97	
			104.51	99.56	51.16	37.95	122.11	77.89	0.00	16.17				
Co	m	L	0.00	97.27	0.00	0.00	0.00	0.00	8.08	0.00	0.00			
			402.40	12.01	0.00	17.63	57.86	56.39	56.50	167.69	77.73	508.13	831.88	
			137.23	232.04	62.50	49.89	181.71	109.44	35.70	22.16				
Co	m	L	0.00	55.31	0.00	0.00	0.00	0.00	5.65	0.00	0.00			
			264.42	7.06	0.00	9.31	29.98	20.72	29.93	92.73	34.96	265.10	444.58	77.13
			127.21	44.58	35.43	106.65	63.68	18.68	11.46					
Co	m	L	0.00	335.43	0.00	0.00	0.00	0.00	19.91	0.00	0.00			
			339.82	27.93	0.00	22.56	58.97	26.50	54.81	142.54	66.46	365.71	616.58	
			118.09	134.97	42.17	34.82	118.09	66.01	14.31	17.26				

Co	m	L	0.00	90.65	0.00	0.00	0.00	0.00	10.23	0.00	0.00			
			414.85	12.76	0.00	16.23	46.94	29.06	42.14	151.17	46.30	304.30	605.62	
			114.34	172.71	42.64	38.72	103.79	81.43	24.32	15.67				
Co	m	L	0.00	547.65	0.00	0.00	0.00	0.00	0.00	8.94	0.00	0.00		
			309.61	11.94	0.00	15.02	43.41	17.79	41.93	133.08	48.95	323.56	522.62	92.27
			135.90	41.53	33.21	109.25	69.56	23.25	0.00					
Co	m	F	0.00	55.93	0.00	0.00	0.00	0.00	0.00	5.01	0.00	0.00		
			249.15	11.02	0.00	12.92	41.53	14.19	33.62	129.31	84.32	359.25	635.24	
			135.38	244.56	71.12	65.68	237.36	199.44	167.80	24.65				
Co	m	F	0.00	14.30	0.00	0.00	0.00	0.00	0.00	4.87	0.00	0.00		
			258.31	12.78	0.00	12.94	74.28	33.47	47.76	120.69	72.12	311.02	586.90	
			150.00	116.61	46.49	41.85	115.58	108.63	17.01	63.10				
Co	m	F	0.00	13.55	0.00	0.00	0.00	0.00	0.00	2.24	0.00	0.00		
			202.03	7.71	0.00	5.26	38.67	9.69	17.98	107.97	46.69	338.72	617.77	
			110.21	162.01	42.73	37.62	158.42	148.83	60.29	20.32				
Co	m	F	0.00	279.87	0.00	0.00	0.00	0.00	0.00	10.16	0.00	0.00		
			297.66	8.74	0.00	9.43	42.31	12.07	26.77	111.82	55.89	259.01	413.63	68.59
			139.52	32.58	25.60	93.45	70.25	29.21	13.19					
Co	m	F	0.00	10.04	0.00	0.00	0.00	0.00	0.00	4.27	0.00	0.00		
			241.76	6.38	0.00	4.44	28.32	7.81	17.85	77.85	53.19	252.01	345.84	46.70
			121.94	26.02	17.56	104.80	62.97	29.57	13.05					
Co	m	F	0.00	13.21	0.00	0.00	0.00	0.00	0.00	4.23	0.00	0.00		
			235.61	4.83	0.00	6.37	21.86	12.54	27.23	76.93	98.19	416.77	576.32	87.32
			116.23	49.56	39.24	257.14	182.09	40.71	48.42					
Co	m	F	0.00	66.92	0.00	0.00	0.00	0.00	0.00	5.99	0.00	0.00		
			282.87	9.52	0.00	9.78	40.14	0.00	32.99	85.59	56.21	255.14	543.75	
			138.22	106.42	37.41	38.27	136.90	154.63	33.84	35.03				
Co	m	F	0.00	12.77	0.00	0.00	0.00	0.00	0.00	5.97	0.00	0.00		
			239.00	6.98	0.00	5.24	42.40	8.08	26.69	88.98	33.81	241.80	528.34	
			113.83	144.56	31.47	32.06	109.14	105.51	30.09	13.05				
Co	m	F	0.00	20.41	0.00	0.00	0.00	0.00	0.00	6.90	0.00	0.00		
			204.55	6.90	0.00	9.17	43.79	14.21	29.31	96.76	64.00	290.62	416.34	63.45
			117.52	25.59	18.83	99.59	81.72	33.72	63.45					
Co	m	F	0.00	2.12	0.00	0.00	0.00	0.00	0.00	6.31	0.00	0.00		
			253.39	4.72	0.00	4.37	24.37	5.43	17.88	83.48	33.33	190.86	422.18	96.93
			123.72	21.36	26.90	97.70	102.12	24.42	12.63					
Py	f	L	0.00	4.13	0.00	0.00	0.00	0.00	0.00	11.79	0.00	0.00		
			370.73	5.01	3.73	12.08	39.69	17.68	43.42	124.36	73.28	463.56	737.03	86.15
			146.17	51.28	40.08	183.20	94.11	18.07	26.42					
Py	f	L	0.92	1.71	0.00	0.00	0.00	0.00	0.00	12.67	3.67	4.71		
			414.25	8.17	8.54	18.04	41.17	23.29	52.50	142.71	65.08	464.50	762.58	
			118.75	171.13	70.17	65.92	193.58	117.79	19.33	23.96				
Py	f	L	1.10	8.41	0.00	0.00	0.00	0.00	0.00	11.08	0.55	1.24		
			523.62	0.00	6.30	19.72	70.40	25.32	58.59	167.37	107.67	723.44	1043.24	
			119.76	242.88	84.28	59.79	326.56	138.69	38.60	52.02				
Py	f	L	0.00	5.15	0.00	0.00	0.00	0.00	0.00	8.53	2.24	2.07		
			504.40	0.00	5.23	9.59	34.74	13.54	28.41	100.09	121.68	415.26	436.41	44.42
			120.45	43.40	12.97	203.34	59.50	20.84	29.29					
Py	f	L	0.55	6.33	0.00	0.00	0.00	0.00	0.00	11.13	2.73	1.64		
			503.90	4.72	5.47	12.85	34.17	19.09	47.91	97.77	79.54	438.54	660.13	77.82
			85.63	55.10	39.40	226.24	93.56	11.64	38.73					

Py	f	L	0.00	6.00	0.00	0.00	0.00	0.00	12.47	1.63	1.47	
	484.87	0.00	6.67	17.36	39.99	21.76	45.63	141.02	114.46	619.66	827.64	87.73
	176.53	66.32	46.98	294.84	118.35	33.88	60.29					
Py	f	L	0.79	4.48	0.00	0.00	0.00	0.00	18.47	2.12	3.82	
	491.37	14.61	7.68	16.19	42.92	25.61	47.99	199.92	78.12	536.36	648.61	68.62
	160.73	41.39	28.23	122.25	56.25	13.53	17.43					
Py	f	L	0.58	3.95	0.00	0.00	0.00	0.00	5.86	0.00	0.00	
	194.60	0.00	2.21	5.28	15.44	7.95	17.18	65.24	40.34	233.43	316.31	33.08
	75.33	22.63	16.31	100.23	39.64	10.16	16.48					
Py	f	L	0.00	7.42	0.00	0.00	0.00	0.00	9.48	0.00	0.00	
	427.73	4.43	4.02	10.10	0.00	13.20	29.38	107.22	75.05	374.95	528.45	62.16
	133.61	43.51	34.02	173.20	71.96	17.94	40.21					
Py	f	L	0.00	8.25	0.00	0.00	0.00	0.00	7.72	0.29	0.41	
	358.07	0.00	5.71	12.07	24.27	14.29	34.58	96.30	53.06	372.98	578.97	70.35
	130.51	43.78	32.94	161.23	73.55	23.94	25.01					
Py	f	F	2.30	6.33	0.00	0.00	0.00	0.00	7.07	1.15	0.00	
	353.80	6.21	5.34	14.10	56.19	27.70	45.50	172.83	121.54	700.04	718.41	49.24
	169.34	60.50	32.31	221.91	81.59	29.22	45.62					
Py	f	F	11.39	15.43	0.00	0.00	0.00	0.00	9.29	4.71	7.06	
	370.49	20.09	9.67	16.35	57.59	23.75	38.00	139.81	100.17	530.22	523.79	55.40
	151.20	50.23	0.00	177.64	64.69	24.97	26.94					
Py	f	F	3.19	2.82	0.00	0.00	0.00	0.00	7.98	0.00	0.00	
	523.45	4.73	5.40	13.26	66.18	26.46	40.33	159.73	144.44	789.44	694.05	64.58
	161.39	77.29	26.09	305.03	99.75	23.27	45.73					
Py	f	F	1.85	1.63	0.00	0.00	0.00	0.00	5.24	0.00	0.00	
	293.00	3.43	3.61	7.97	34.65	16.25	25.28	100.35	81.51	448.26	423.38	36.06
	124.26	41.22	11.71	163.23	55.22	16.56	22.32					
Py	f	F	7.85	13.39	0.00	0.00	0.00	0.00	22.03	8.82	0.00	
	677.64	0.00	15.20	26.56	80.51	41.15	59.31	211.36	198.57	797.60	670.95	
	101.85	252.47	97.23	0.00	269.93	86.70	33.39	31.04				
Py	f	F	18.26	20.93	0.00	0.00	0.00	0.00	16.46	7.16	8.96	
	574.59	17.62	17.87	31.65	80.19	63.39	78.48	288.27	203.31	1374.73		
	996.11	79.26	209.93	112.66	25.07	354.38	110.95	30.43	43.14			
Py	f	F	3.57	9.45	0.00	0.00	0.00	0.00	7.49	1.61	2.90	
	305.65	5.22	3.76	9.45	44.12	15.61	30.67	122.78	90.35	445.45	546.82	51.49
	145.61	41.57	27.06	166.82	61.33	22.12	21.96					
Py	f	F	2.44	14.65	0.00	0.00	0.00	0.00	8.50	3.62	2.40	
	296.38	7.95	7.56	16.06	42.01	25.24	43.35	136.50	104.57	662.52	716.06	65.43
	161.34	60.51	13.70	215.35	77.99	31.02	27.83					
Py	f	F	2.32	11.19	0.00	0.00	0.00	0.00	7.10	0.00	1.41	
	324.10	5.26	4.28	9.46	38.34	16.05	29.71	105.65	97.61	449.76	512.09	54.47
	171.90	48.90	0.00	173.74	67.27	27.83	25.75					
Py	f	F	1.04	5.12	0.00	0.00	0.00	0.00	7.62	0.00	0.00	
	473.74	0.00	2.70	6.33	24.30	13.73	19.93	97.17	84.56	424.30	361.55	28.92
	107.66	38.91	11.69	171.91	56.35	23.26	23.35					
Py	m	L	59.02	43.27	0.00	0.00	0.00	0.00	3.27	0.00	0.00	
	252.68	2.93	2.83	8.49	25.51	13.85	31.32	73.80	58.73	381.95	472.73	46.10
	69.12	41.22	27.51	173.22	65.07	10.83	21.85					
Py	m	L	45.77	59.75	0.00	0.00	0.00	0.00	7.82	0.00	6.15	
	305.06	16.50	8.91	17.77	55.20	20.99	41.12	121.33	83.84	486.14	643.36	68.89
	158.77	50.20	34.96	200.86	75.04	27.72	24.73					

Py	m	L	78.41	68.19	0.00	0.00	0.00	0.00	7.75	0.00	6.26			
			312.73	9.60	7.58	15.45	52.25	16.60	38.24	111.70	77.26	384.88	633.33	75.28
			151.30	48.08	39.18	207.58	95.34	32.55	37.00					
Py	m	L	62.32	87.25	0.00	0.00	0.00	0.00	0.00	3.04	0.00	0.00		
			240.25	4.38	2.85	10.09	44.81	15.70	37.87	92.10	66.89	420.27	675.93	77.45
			145.29	46.43	34.71	209.51	97.15	30.54	30.35					
Py	m	L	122.26	120.53	0.00	0.00	0.00	0.00	0.00	3.92	0.00	2.12		
			234.09	9.25	5.64	12.70	80.72	24.61	42.95	145.14	123.75	623.67	720.53	68.73
			200.16	65.67	42.87	287.23	101.72	52.66	39.18					
Py	m	L	52.67	101.11	0.00	0.00	0.00	0.00	0.00	5.25	0.00	0.38		
			407.76	6.27	5.59	12.87	63.08	21.35	37.83	130.46	103.47	597.16	684.43	61.59
			187.08	57.30	34.80	272.63	95.90	35.81	49.45					
Py	m	L	58.39	74.35	0.00	0.00	0.00	0.00	0.00	3.85	0.00	0.69		
			249.22	3.20	2.77	6.27	50.82	12.46	30.75	67.86	61.98	344.68	494.90	51.82
			125.26	36.07	26.30	158.35	62.67	21.54	20.93					
Py	m	L	55.96	119.58	0.00	0.00	0.00	0.00	0.00	3.54	0.00	0.00		
			234.71	4.83	3.45	9.05	60.94	14.84	32.79	92.63	63.57	377.50	514.46	49.50
			124.37	41.36	28.20	182.19	73.43	18.14	24.61					
Py	m	L	51.05	63.45	0.00	0.00	0.00	0.00	0.00	5.13	0.00	1.35		
			299.81	4.29	3.45	7.74	34.64	12.45	25.31	91.42	64.01	334.50	452.91	43.73
			118.32	34.08	12.77	148.81	60.65	33.10	21.49					
Py	m	L	45.64	59.77	0.00	0.00	0.00	0.00	0.00	3.93	0.00	1.23		
			253.75	8.97	4.36	11.86	53.38	15.29	30.47	98.03	61.36	404.24	539.86	56.02
			146.74	40.79	26.90	169.53	66.77	33.97	27.89					
Py	m	F	32.69	41.64	0.00	0.00	0.00	0.00	0.00	6.07	0.00	3.76		
			231.16	6.11	6.80	9.41	44.91	15.83	22.36	65.85	79.41	345.68	318.06	28.28
			108.91	31.85	7.41	119.52	33.77	11.29	18.71					
Py	m	F	42.19	35.53	0.00	0.00	0.00	0.00	0.00	6.06	0.00	0.00		
			260.06	4.15	3.50	7.14	20.51	8.01	16.79	52.06	41.89	186.07	246.52	28.30
			64.43	18.65	0.00	69.93	34.44	9.52	11.99					
Py	m	F	14.86	17.73	0.00	0.00	0.00	0.00	0.00	3.12	0.00	0.00		
			203.34	0.00	2.58	6.14	26.26	11.37	18.20	59.64	60.88	265.31	266.47	22.30
			71.67	22.30	0.00	76.53	23.65	6.61	8.57					
Py	m	F	108.90	90.95	0.00	0.00	0.00	0.00	0.00	3.80	0.00	0.00		
			262.48	2.30	4.65	8.35	50.18	16.61	22.76	69.58	73.74	459.28	391.00	30.27
			142.07	39.72	19.81	169.38	51.63	29.46	20.06					
Py	m	F	99.91	27.47	0.00	0.00	0.00	0.00	0.00	2.16	0.00	0.38		
			189.71	1.46	2.03	6.23	16.21	9.85	20.63	66.61	53.75	319.66	388.78	38.25
			86.70	34.19	0.00	151.41	63.96	22.66	28.18					
Py	m	F	84.93	34.70	0.00	0.00	0.00	0.00	0.00	2.51	0.00	0.26		
			213.38	2.15	2.33	6.03	21.74	10.68	17.85	58.22	59.45	291.78	305.66	24.34
			90.68	26.89	8.36	113.79	39.13	16.76	15.89					
Py	m	F	150.71	91.42	0.00	0.00	0.00	0.00	0.00	2.37	0.00	0.00		
			176.84	8.06	3.71	10.26	37.82	17.90	30.83	83.74	67.31	461.36	515.91	44.16
			106.90	39.84	22.98	155.45	56.79	22.55	16.77					
Py	m	F	30.06	28.64	0.00	0.00	0.00	0.00	0.00	3.30	0.00	0.00		
			240.85	4.43	4.66	7.78	42.10	14.32	23.52	81.82	88.13	331.14	353.75	38.24
			124.20	35.57	10.80	111.76	34.89	12.84	12.10					
Py	m	F	73.96	59.13	0.00	0.00	0.00	0.00	0.00	3.11	0.00	0.34		
			203.91	3.53	2.78	7.58	43.36	13.09	24.72	78.15	80.74	389.41	438.56	44.87
			129.80	42.33	16.95	161.11	55.79	23.92	18.17					

Py	m	F	127.33	72.23	0.00	0.00	0.00	0.00	7.58	0.00	3.44	
	315.33	9.38	8.29	16.04	58.21	20.29	32.13	111.89	94.54	494.16	482.11	53.57
	151.01	56.08	20.24	186.03	60.61	22.20	25.15					

```
#####
# 1 Data reading #
#####

macrolophus <- read.table("S2.Macrolophus.data.TXT",sep="\t",header=T)
summary(macrolophus)
dim(macrolophus)
head(macrolophus)

#####
# 2 Principal components analysis #
#####

library(MASS)

macrolophus.pca <- princomp(macrolophus[,4:31],cor=T)
summary(macrolophus.pca)
macrolophus.pca

#####
# 3 Quadratic discriminant analysis #
#####

macrolophus$PC1 <- macrolophus.pca$scores[,1]
macrolophus$PC2 <- macrolophus.pca$scores[,2]
macrolophus$PC3 <- macrolophus.pca$scores[,3]
macrolophus.qda <- qda(species~PC1+PC2+PC3,data=macrolophus)
macrolophus.qda

macrolophus.predict <- predict(macrolophus.qda)
macrolophus.predict

macrolophus$QDA.prob.Me <- macrolophus.predict$posterior[,1]
macrolophus$QDA.prob.Co <- macrolophus.predict$posterior[,2]
macrolophus$QDA.prob.Py <- macrolophus.predict$posterior[,3]
macrolophus$QDA.predicted.species <- macrolophus.predict$class

results <-
macrolophus[,c("species","QDA.prob.Me","QDA.prob.Co","QDA.prob.Py","QDA.predicted.species")]
results[,2:4] <- round(results[,2:4],2)
results

correct.prediction <-
table(macrolophus$species,macrolophus$QDA.predicted.species,dnn=c("real","predicted"))
correct.prediction
prop.table(correct.prediction,1)
prop.table(correct.prediction,2)

total.correct <- sum(diag(correct.prediction))/sum(correct.prediction)
```

```

total.correct

total.error <- 1-total.correct
total.erro

#####
# 4 Quadratic discriminant analysis by sex #
#####

#4.1 males

macrolophus.male <- macrolophus[macrolophus$sex=="m",]

macrolophus.pca <- princomp(macrolophus.male[,4:31],cor=T)
summary(macrolophus.pca)
macrolophus.pca

macrolophus.male$PC1 <- macrolophus.pca$scores[,1]
macrolophus.male$PC2 <- macrolophus.pca$scores[,2]
macrolophus.male$PC3 <- macrolophus.pca$scores[,3]
macrolophus.qda <- qda(species~PC1+PC2+PC3,data=macrolophus.male)
macrolophus.qda

macrolophus.predict <- predict(macrolophus.qda)
macrolophus.predict

macrolophus.male$QDA.prob.Me <- macrolophus.predict$posterior[,1]
macrolophus.male$QDA.prob.Co <- macrolophus.predict$posterior[,2]
macrolophus.male$QDA.prob.Py <- macrolophus.predict$posterior[,3]
macrolophus.male$QDA.predicted.species <- macrolophus.predict$class

results <-
macrolophus.male[,c("species","QDA.prob.Me","QDA.prob.Co","QDA.prob.Py","QDA.predicted.species")]
results[,2:4] <- round(results[,2:4],2)
results

correct.prediction <-
table(macrolophus.male$species,macrolophus.male$QDA.predicted.species,dnn=c("real","predicted"))
correct.prediction
prop.table(correct.prediction,1)
prop.table(correct.prediction,2)

total.correct <- sum(diag(correct.prediction))/sum(correct.prediction)
total.correct

total.error <- 1-total.correct
total.error

#4.2 females

```

```

macrolophus.female <- macrolophus[macrolophus$sex=="f",]

macrolophus.pca <- princomp(macrolophus.female[,c(4:5,7:8,10:31)],cor=T)
summary(macrolophus.pca)
macrolophus.pca

macrolophus.female$PC1 <- macrolophus.pca$scores[,1]
macrolophus.female$PC2 <- macrolophus.pca$scores[,2]
macrolophus.female$PC3 <- macrolophus.pca$scores[,3]
macrolophus.qda <- qda(species~PC1+PC2+PC3,data=macrolophus.female)
macrolophus.qda

macrolophus.predict <- predict(macrolophus.qda)
macrolophus.predict

macrolophus.female$QDA.prob.Me <- macrolophus.predict$posterior[,1]
macrolophus.female$QDA.prob.Co <- macrolophus.predict$posterior[,2]
macrolophus.female$QDA.prob.Py <- macrolophus.predict$posterior[,3]
macrolophus.female$QDA.predicted.species <- macrolophus.predict$class

results <-
macrolophus.female[,c("species","QDA.prob.Me","QDA.prob.Co","QDA.prob.Py","QDA.predicted.species
")]
results[,2:4] <- round(results[,2:4],2)
results

correct.prediction <-
table(macrolophus.female$species,macrolophus.female$QDA.predicted.species,dnn=c("real","predicte
d"))
correct.prediction
prop.table(correct.prediction,1)
prop.table(correct.prediction,2)

total.correct <- sum(diag(correct.prediction))/sum(correct.prediction)
total.correct

total.error <- 1-total.correct
total.error

#####
# 5 Prediction for new individuals #
#####

library(mnormt)
tomatomale <-
c(9.3359225,35.24040853,0,0,0,2.620230036,4.143500323,0,227.0959237,6.803160605,
1.675141163,5.174382195,27.78788589,33.50457571,21.56733939,80.30059764,62.56136815,290.
5071812,451.0842287,

```

```
81.21811111,161.8425264,35.04168524,11.66117306,149.6672654,57.47777861,23.24498038,20.6
8211748)
tomatofemale <-
c(0.465840529,33.08057566,0,1.69478331,0,2.434254523,13.35527135,3.219487532,0.805361245,
623.4437673,21.48072989,5.886884465,11.75026023,17.20756204,32.65718085,44.08958108,133.
3946882,100.2624269,668.2018557,
775.8341467,86.4138592,143.781386,65.40976156,43.59786875,237.3767564,101.9367407,24.226
07705,38.82679428)
macrolophus.new <- rbind(tomatomale,tomatofemale)
source("S4.Macrolophus.predict.R")
predict.QDA(macrolophus.pca,macrolophus.new,macrolophus,macrolophus$species)
```

```

predict.QDA <- function(object.pca,new.individuals,data,species)
{
  data <- as.matrix(data[,4:31])
  gravity.center <- apply(data,2,mean)
  centered.data <- data -
matrix(gravity.center,byrow=T,nrow=nrow(data),ncol=ncol(data))
  variances <- apply(data,2,var) * (nrow(data)-1) / nrow(data)
  deviations <- sqrt(variances)
  centered.reduced.data <- centered.data /
matrix(deviations,byrow=T,nrow=nrow(data),ncol=ncol(data))

  X <- centered.reduced.data
  M <- diag(1,nrow=ncol(X))
  D <- diag(1/nrow(X),nrow=nrow(X))

  #svd decomposition
  eigen.X <- eigen(t(X) %*% D %*% X %*% M)

  #projections for individuals
  F.proof <- X %*% M %*% eigen.X$vectors

  #prediction for new individuals

  X.star <- new.individuals
  X.star <- X.star - matrix(gravity.center,byrow=T,nrow=nrow(X.star),ncol=ncol(X.star))
  X.star <- X.star / matrix(deviations,byrow=T,nrow=nrow(X.star),ncol=ncol(X.star))
  F.star <- X.star %*% M %*% eigen.X$vectors #projections for new individuals

  mu.Me.proof <- apply(F.proof[species=="Me",1:3],2,mean)
  sigma.Me.proof <- var(F.proof[species=="Me",1:3])
  mu.Co.proof <- apply(F.proof[species=="Co",1:3],2,mean)
  sigma.Co.proof <- var(F.proof[species=="Co",1:3])
  mu.Py.proof <- apply(F.proof[species=="Py",1:3],2,mean)
  sigma.Py.proof <- var(F.proof[species=="Py",1:3])

  prob.X.star.cond.Me <-
dmnorm(x=F.star[,1:3],mean=mu.Me.proof,varcov=sigma.Me.proof)
  prob.X.star.cond.Co <-
dmnorm(x=F.star[,1:3],mean=mu.Co.proof,varcov=sigma.Co.proof)
  prob.X.star.cond.Py <-
dmnorm(x=F.star[,1:3],mean=mu.Py.proof,varcov=sigma.Py.proof)

  prob.Me <- prop.table(table(macrolophus$species))["Me"]
  prob.Co <- prop.table(table(macrolophus$species))["Co"]
  prob.Py <- prop.table(table(macrolophus$species))["Py"]

  prob.X.star <- prob.Me * prob.X.star.cond.Me + prob.Co * prob.X.star.cond.Co + prob.Py
* prob.X.star.cond.Py

  prob.Me.cond.X.star <- prob.Me * prob.X.star.cond.Me /prob.X.star
  prob.Co.cond.X.star <- prob.Co * prob.X.star.cond.Co /prob.X.star
  prob.Py.cond.X.star <- prob.Py * prob.X.star.cond.Py /prob.X.star

```



```
results <- cbind(prob.Me.cond.X.star,prob.Co.cond.X.star,prob.Py.cond.X.star)
rownames(results) <- rownames(X.star)
results <- round(results,3)
results
```

```
}
```